Histology of spheroidal degeneration of the cornea in Labrador

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SUMMARY  Nine specimens of the corneas of patients from Labrador and Northern Newfoundland affected by spheroidal degeneration (climatic droplet keratopathy) have been examined microscopically. Histochemical stains confirmed studies of similar corneal degenerations from other geographical areas that the droplets contain a protein which does not have all the characteristic properties of elastic tissue. Staining compatible in some instances with fibrin and 'fibrinoid' was found. By immunoperoxidase techniques the droplets were located in the zones of greatest concentration of various plasma constituents, especially albumin and immunoglobulins G and A. Reasons are given why the abnormal deposits are not thought to be derived directly from corneal collagen.

It is suggested that some of the plasma proteins, which are known to be diffusing through the cornea from the limbal vessels under normal conditions, are acted upon by the ultraviolet light reflected from the ice of Labrador and degraded so that they accumulate in the superficial stroma.

There is current interest in a type of degeneration of the cornea which is now termed either droplet keratopathy (Freedman, 1973a) or spheroid (spheroidal) degeneration (Fraunfelder et al., 1972). Spheroidal or nodular deposits of abnormal hyaline material accumulate in Bowman's layer and the superficial stroma. At least 12 English-language papers have described the light or electron microscopy of this condition. They included material from regions where external climatic factors appeared to be the main aetiological agent (Klintworth, 1972; D'Alena and Wood, 1972; Garner et al., 1973; Johnson and Ghosh, 1975), cases from temperate regions where no definite aetiology was proposed (Etzine and Kaufmann, 1964; Christensen, 1973; Garner et al., 1976), and cases of spheroidal degeneration in association with a corneal dystrophy (Garner, 1970), and in association with other intraocular disease and trauma (Hanna and Fraunfelder, 1972; Brownstein et al., 1973; Rodrigues et al., 1975; Cursino and Fine, 1976).

Analysis of all the staining reactions reported shows a good general agreement between authors, who used tissue from different sources. Within the spheroidal bodies there is a protein, containing a considerable proportion of the amino-acids trypto-

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phan, tyrosine, cysteine, and cystine, which are not normally present in the corneal stroma in large amounts. The deposits also have some of the staining properties of ribonucleic acid. At one time the protein was thought to be related to keratin and was labelled 'keratinoid' (Garner, 1970). The most popular terms at present are 'elastotic degeneration' or 'elastosis', but Garner et al. (1976) have now shown that the deposits do not stain with 2 of the characteristic stains for elastic tissue. The precise chemical identity of the material has therefore not yet been determined. Most authors imply that it is formed in situ from structural components of the cornea. Klintworth (1972), however, proposed that degenerate elastotic material formed from collagen beneath the adjacent conjunctiva might be diffusing into the cornea.

The form of droplet keratopathy, or spheroidal degeneration, seen with such remarkable frequency in the people of Northern Newfoundland and Labrador was originally described in a classic paper by Freedman (1965) under the title 'Labrador Keratopathy'. Additional details of the clinical features and prevalence in that geographical area may be found in the reports by Gillan (1970), Freedman (1973a, 1973b), Wyatt (1973), Johnson and Ghosh (1975), and Young and Finlay (1975).

The electron microscopy of specimens from Northern Newfoundland and Labrador has already
been published (Johnson and Ghosh, 1975). This report describes additional histochemistry of corneal tissue from the same geographical region, and puts forward a new hypothesis for the origin of the material.

Materials and methods

The 9 patients from whom histological material came are listed in Table 1. The tissue was obtained by central lamellar keratotomy for visual indications, by keratotomy of a large pterygium or by superficial biopsy of peripheral cornea at the time of making a fornix-based flap as part of a cataract extraction. The tissue was fixed immediately in formalin-saline and after transportation embedded in Paramat (Gurr). In addition, 2 whole eyes were donated for grafting, from 1 of which a lamellar button was cut, so that these eyes were fixed 12 hours after death.

Six control donor eyes were obtained in England. The anterior segments of 3 were fixed in absolute ethanol 3 parts to glacial acetic acid 1 part for 18 hours, the remainder in formalin-saline. This control tissue was fixed 12 to 36 hours after death.

The pathological specimens were stained by the methods listed in Table 2 using standard techniques according to Pearse (1968) or Culling (1974). The stains for elastin, namely Verhoeff-van Gieson, orcein, Gomori’s aldehyde fuchsin, and rhodanil blue (McConaill and Gurr, 1964) were controlled by staining the elastic fibres of sections of temporal artery. Specimens were also examined with the Zeiss fluorescence microscope for natural fluorescence as well as after staining with thioflavin T using the BG 12 excitor filter with barrier filters 53 and 44.

In addition sections of both specimens and controls were tested by the antiperoxidase-peroxidase (PAP) method (Burns, 1975), using commercial unconjugated antihuman antibodies prepared in the rabbit against serum albumin, IgG, IgA, IgM, C3 complement, α2 macroglobulin, transferrin, and fibrinogen (DAKO—immunoglobulins). Since albumin or other adhesive was not used on the slides, many specimens washed away. Material was limited, and series of slides were obtained for 4 pathological corneas as well as 4 controls.

One specimen of formalin-fixed tissue (Case 5), originally embedded in Paramat, was dewaxed and

<table>
<thead>
<tr>
<th>Case no.</th>
<th>Race</th>
<th>Age</th>
<th>Sex</th>
<th>Grade of kerato-</th>
<th>Source of specimen</th>
<th>Associated conditions</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Caucasian</td>
<td>68</td>
<td>F</td>
<td>Peripheral</td>
<td>kerato-</td>
<td>Cataract: diabetes</td>
</tr>
<tr>
<td>2</td>
<td>Caucasian</td>
<td>58</td>
<td>M</td>
<td>Whole eyes (one</td>
<td>kerato-</td>
<td>—</td>
</tr>
<tr>
<td>3</td>
<td>Caucasian</td>
<td>62</td>
<td>M</td>
<td>Keratotomy</td>
<td>kerato-</td>
<td>Large pterygia</td>
</tr>
<tr>
<td>4</td>
<td>Caucasian</td>
<td>64</td>
<td>M</td>
<td>Peripheral</td>
<td>kerato-</td>
<td>Cataract</td>
</tr>
<tr>
<td>5</td>
<td>Caucasian</td>
<td>72</td>
<td>M</td>
<td>Central lamellar keratotomy</td>
<td>kerato-</td>
<td>Cataract</td>
</tr>
<tr>
<td>6</td>
<td>Caucasian</td>
<td>77</td>
<td>M</td>
<td>Central and peripheral</td>
<td>kerato-</td>
<td>Cataract</td>
</tr>
<tr>
<td>7</td>
<td>Caucasian</td>
<td>81</td>
<td>M</td>
<td>Peripheral</td>
<td>kerato-</td>
<td>Cataract</td>
</tr>
<tr>
<td>8</td>
<td>Eskimo</td>
<td>61</td>
<td>M</td>
<td>Peripheral</td>
<td>kerato-</td>
<td>Cataract</td>
</tr>
<tr>
<td>9</td>
<td>Eskimo</td>
<td>86</td>
<td>M</td>
<td>Peripheral</td>
<td>kerato-</td>
<td>Cataract</td>
</tr>
</tbody>
</table>

Table 2 Results of histochemical staining of spheroidal degeneration

<table>
<thead>
<tr>
<th>Stain</th>
<th>Cases tested</th>
<th>Staining of droplets</th>
</tr>
</thead>
<tbody>
<tr>
<td>Haematoxylin and eosin</td>
<td>1, 2, 4, 5</td>
<td>Small droplets eosinophilic; larger eosinophilic with basophilic centres</td>
</tr>
<tr>
<td>Periodic-acid-Schiff</td>
<td>2, 4</td>
<td>Negative</td>
</tr>
<tr>
<td>Toluidine blue</td>
<td>1, 2, 4, 5</td>
<td>Blue-green; no metachromasia</td>
</tr>
<tr>
<td>Feulgen</td>
<td>2, 4</td>
<td>Negative</td>
</tr>
<tr>
<td>Methyl green-pyronin</td>
<td>2, 4</td>
<td>Bright red; no change after RNA-ase</td>
</tr>
<tr>
<td>Gallocyanine-chromalum</td>
<td>2</td>
<td>Grey; no change after RNA-ase</td>
</tr>
<tr>
<td>Mallory’s PTAH</td>
<td>1–9</td>
<td>Purple (fibrinoid purple)</td>
</tr>
<tr>
<td>Martius-scarlet-blue</td>
<td>1–9</td>
<td>All red or orange-brown (fibrin red)</td>
</tr>
<tr>
<td>Masson 44/41</td>
<td>1, 2, 3, 4, 5, 7, 8, 9</td>
<td>Orange-brown (old fibrin red)</td>
</tr>
<tr>
<td>Rhodanil blue</td>
<td>2</td>
<td>Purplish-pink (fibrin deep red)</td>
</tr>
<tr>
<td>Verhoeff-van Gieson</td>
<td>1, 2, 4, 5</td>
<td>Almost black, but browner than blue-black of elastic fibres; some olive-brown</td>
</tr>
<tr>
<td>Orcein</td>
<td>1, 2, 4, 6, 8, 9</td>
<td>Negative (pale yellow)</td>
</tr>
<tr>
<td>Gomori’s aldehyde fuchsin</td>
<td>1, 2, 6, 8, 9</td>
<td>Variable pink, some colourless; elastic fibres very deep pink; deep pink material in pingueculum</td>
</tr>
<tr>
<td>Fontana silver</td>
<td>1, 2, 3, 5, 6, 7, 8, 9</td>
<td>Negative</td>
</tr>
<tr>
<td>Congo Red</td>
<td>1, 2, 4</td>
<td>Negative</td>
</tr>
<tr>
<td>Thioflavin T</td>
<td>5</td>
<td>Negative (fluorescence less than natural fluorescence)</td>
</tr>
</tbody>
</table>
embedded in Araldite, without further fixation or staining, for electron microscopy.

Results

The staining reactions of the droplets are summarised in Table 2. The results were very similar in all specimens tested.

With methyl green pyronin the spheroids stained bright pink. This was not altered by pretreatment with RNA-ase for 2 hours. Similarly, the pale grey staining with gallocyanin chromalum was unaffected by RNA-ase. These reactions are therefore not consistent with the material being ribonucleic acid.

The Martius-scarlet-blue method stains fibrin red. The red or orange-brown reaction of the material in these droplets indicated a staining property in common with fibrin (Fig. 8).

The size and distribution of individual droplets were shown up particularly well by Mallory’s phosphotungstic acid-haematoxylin method. At the extreme periphery, in the early stages of the keratopathy, there was characteristically a margin of cornea free of droplets (Fig. 1). The margin of the affected layer began abruptly with a dense accumulation of rather large spheroids. These became both smaller and sparser towards the centre of the cornea (Fig. 2). They were absent from the pupillary area in grade I. The deep purple of the droplets with this stain was suggestive of fibrinoid rather than of fibrin, which should appear a bright blue. In a more severe grade of keratopathy the periphery of the larger droplets stained darker than the centres (Fig. 3). The smaller deposits stained uniformly across their diameter. Paler staining in the centre of the larger droplets was also a feature of staining by haematoxylin and eosin (Fig. 6).

One percent rhodanile blue, pH 7.0 for 30 minutes (McConaill and Gurr, 1964) is said to stain fibrin deep red, elastic fibres blue, and collagen fibres colourless. The droplets in the 1 specimen so far tested were purplish-pink. The larger elastic fibres in the temporal artery were blue. The small elastic fibres which stain with Verhoeff, orcein, and aldehyde fuchsin were not stained with rhodanile blue.

The epithelium was normal in all cases except Case 9, a severe example of grade III keratopathy. In this specimen the epithelium was thinned to 2 or 3 layers of cells over the large irregular conglomerates.
of droplet material; Bowman's membrane was also destroyed in this specimen (Fig. 4).

The droplets in all the specimens showed an intense white-yellow natural fluorescence. The fluorescence was rather less pronounced after staining with thioflavin T. The Congo red reaction for amyloid was negative.

**IMMUNOPEROXIDASE METHOD**

The density of staining of the corneas by the immunoperoxidase method varied widely according to the plasma component against which the antibody was active, and also varied to some extent between the different specimens.

**Table 3**  *Results of reaction with immune sera*

<table>
<thead>
<tr>
<th>Anti-sera* against</th>
<th>Reaction in Epithelium</th>
<th>Bowman's</th>
<th>Concentrated in periphery of stroma anteriorly</th>
<th>Pathological specimens</th>
</tr>
</thead>
<tbody>
<tr>
<td>Albumin</td>
<td>+ +</td>
<td>0</td>
<td>+ (also deep periphery)</td>
<td>Droplets in zone of densest reaction in anterior stroma; have stained borders</td>
</tr>
<tr>
<td>IgG</td>
<td>± (basal cells)</td>
<td>+</td>
<td>±</td>
<td>Droplets in zone of darkest reactivity. Staining rims (Case 6)</td>
</tr>
<tr>
<td>Transferrin</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>Staining around droplets in Case 2. Suggestion of staining rims (Case 6)</td>
</tr>
<tr>
<td>Fibrinogen</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>Staining not especially concentrated around droplets</td>
</tr>
<tr>
<td>IgA</td>
<td>±</td>
<td>±</td>
<td>±</td>
<td>Concentration of antibody around droplets in anterior stroma (Case 6); staining droplet rims</td>
</tr>
<tr>
<td>Complement C₃</td>
<td>±</td>
<td>+</td>
<td>+</td>
<td>Not usually concentrated around droplets</td>
</tr>
<tr>
<td>IgM</td>
<td>Trace</td>
<td>±</td>
<td>±</td>
<td>Concentrated around droplets in one case</td>
</tr>
<tr>
<td>α-2 Macroglobulin</td>
<td>0</td>
<td>0</td>
<td>Trace</td>
<td>Slight staining of peripheral anterior stroma not especially around droplets</td>
</tr>
</tbody>
</table>

*Anti-sera listed in order of average overall density of staining of corneal sections*
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Fig. 6 Grade I (Case 2). Haematoxylin and eosin. Centres of droplets paler than periphery (×340)

Fig. 7 Grade III (Case 5). Orcein-van Gieson. Droplets, pale yellow, did not stain for elastin (×170)

Fig. 8 Grade III (Case 5). Martius-scarlet-blue method for fibrin (×170)

Fig. 9 Grade II (Case 8). Masson 44/41 method for old fibrin (×340)

Fig. 10 Periphery of cornea of Case 2 (grade I keratopathy) reacted with antibody-enzyme (PAP) method for human transferrin. The antibody is concentrated in the anterior part of the periphery of the cornea. The few droplets in this section (pale blue) are in the most superficial layer of this zone (×75)

Fig. 11 More central portion of cornea of Case 2 (Grade I) reacted by PAP method for IgG. The fine colourless droplets are in Bowman's membrane, which is also the most densely staining layer for antibody. This slide is from the same zone of cornea as Fig. 2 (×115)
densest staining in this specimen lay beneath the epithelium at the zone of transition from corneal to conjunctival epithelium (Figs. 10 and 12). Many of the droplets in these sections were deposited here, peripheral to the termination of Bowman’s membrane. Others were in the periphery of Bowman’s membrane itself (Fig. 11). Cases 6 and 9 (grade III keratopathy) had larger droplets more centrally situated. In all slides reacted with antialbumin the droplets were embedded in the zone of densest stain, appearing blue against the granular dark surrounding stroma. They appeared to be swimming in a sea of albumin. This concentration of staining around the droplets applied also to IgG and IgA (Fig. 11). It was not so pronounced with the antibodies to the other plasma components.

In no sections were the droplets stained across their whole diameter. In all cases exposed to the antibody for albumin there were distinct staining rims to the droplets, and in Case 6 with anti-IgG and anti-IgA as well. This appearance suggested that these proteins might be contributing to or were at least adhering to the outer shell of the droplets. There was also the suggestion of staining rims with antitransferrin.

Any aggregate of protein material in the cornea might be liable to be coated with plasma proteins. The staining rims do not necessarily prove that the spheroidal bodies are themselves derived from altered plasma proteins. The main conclusion from these tests is that these plasma constituents are present in the corneal stroma with a concentration gradient from the periphery to the centre, and that the droplets are spatially related to the areas of heaviest concentration, notably of albumin and to a less extent immunoglobulins and perhaps transferrin. It was also clear from Case 2 (grade I keratopathy), where the termination of Bowman’s membrane was conspicuous in some slides, that droplets were deposited further peripherally and not restricted to Bowman’s layer.

**ELECTRON MICROSCOPY**
Electron microscopy of the section fixed in formalin

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**Fig. 5** Section of normal control cornea near periphery, reacted with antiserum to human serum albumin, followed by the antibody-enzyme (PAP) method. Albumin appears to be concentrated in the superficial stroma, while Bowman’s membrane does not stain. (Magnification of negative × 160, of this print × 470.)

**Fig. 12** Drawing of Fig. 10 (section of peripheral cornea of Case 2) showing position of droplets in relation to density of staining after reaction with antibody to human transferrin followed by the antibody-enzyme (PAP) method. Appearance with anti-IgA and anti-IgM similar but paler; with anti-IgG staining extended more uniformly into corneal stroma.

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without any staining showed the droplets to be electron-dense, contrasting sharply with the surrounding stroma.

Discussion

Our results correspond in general with those of previous reports. Although the droplets stain with pyronin, they are not affected by RNA-ase. The Feulgen reaction has been universally negative in previous reports and in our tests. We do not think that nucleic acids are a main component of this material.

There is no evidence in any report or in these studies of a major carbohydrate component. Natural fluorescence in the case of amyloid is thought to reflect a high tryptophan and tyrosine content, and possibly an organised and orientated structure (Pearse, 1968). The explanation of the natural fluorescence of the corneal droplets from Labrador may be the same. However, the Congo red stain did not support the presence of amyloid, nor have previous electron micrographs (Johnson and Ghosh, 1975) given an impression of the fibrillar structure of amyloid.

The recent consensus has been to designate the spheroids as 'elastotic degeneration' or 'elastosis' (Brownstein et al., 1973; Rodrigues et al., 1975).

However, Garner and colleagues (1976) showed that spheroids in the cornea did not stain with either orcein or aldehyde fuchsin as if they had properties of elastic tissue, differing in this respect from elastotic material beneath pingueculae. On the other hand degenerated material from both sites stained with Verhoeff-van Gieson. This has now been confirmed in the tissue from Labrador (Fig. 7).

Gillman and his associates (1954) stressed the value of Mallory's phosphotungstic acid haematoxylin in distinguishing between 'fibrinoid', which stained purple as in our sections, and elastotic collagen (pale yellow or unstained). In a consideration of the class of protein which the droplets might contain the positive staining with Martius-scarlet-blue for a fibrin-like material becomes important. Lendrum et al. (1962) recommended this stain for the demonstration of fibrin; by staining both collagen and fibrin strong and different colours it shows their relationship with clarity (Fig. 8). In our sections there was no transitional zone between the surrounding collagen and the droplets.

However, the material is unlikely to consist predominantly of unaltered fibrin because of its consistent autofluorescence. Moreover, the immunoperoxidase method indicated that the droplets do not lie in the areas where a substance behaving antigenically as fibrinogen is most highly concentrated. Fibrin is known to change its staining properties as it ages, but the Masson 44/41 method did not give the blue-black stain characteristic of old fibrin (Fig. 9).

The term 'fibrinoid' may be used to describe material derived from collagen in a rheumatic nodule. The keratopathy droplets are unlikely to be fibrinoid in this sense because of the demonstration in previous studies of the high content of tryptophan. So-called 'vascular fibrinoid', on the other hand, has been found to be composed largely of fibrin. Lendrum and associates (1962) pointed out that fibrinogen escaping from the lumen of an artery and precipitated intramurally as fibrin forms a spongework or filter which traps and concentrates other substances leaking out from plasma. The deposited admixtures of fibrin and other proteins in this site are thought to bring about slight differences in the staining reaction of the adulterated fibrin.

Staining reactions for fibrin are, according to Lendrum et al. (1962) dependent on the 'fine structure' of the material rather than on a simple chemical reaction in the ordinary sense. This fine structure probably undergoes change with age. The droplets of climatic keratopathy are thought on clinical grounds to accumulate and increase in size very slowly over many years.

If the term fibrinoid is used to signify a material composed of plasma protein derivatives, including fibrin, then it might be applied to these droplets. From the staining reactions described above we should prefer at present to call the abnormal material of the droplets 'fibrinoid' rather than 'elastotic degeneration'.

Clinical Observation

Well-established clinical observations on the Labrador variety of this disease have not been sufficiently taken into account in discussions of the source and composition of the droplets. In his initial report Freedman (1965) clearly described the earliest stages, in which the droplets are first seen in the interpalpebral strip of exposed cornea at the temporal and nasal margins. The border of the zone of droplet deposition is separated from the nasal margin by a strip of clear cornea and is irregular, like a 'geographical coastline'. Towards the middle of the cornea the droplets become smaller and more sparsely arranged; only in the advanced stages do they reach the optical axis. Although the marginal strip is not completely free of droplets in every case, this general appearance (Fig. 13) has been confirmed by Johnson and Ghosh (1975) and Young and Finlay (1975). The clear zone is approximately 0.6 mm in width. Vascularisation
is not a feature of Labrador keratopathy, but occasionally, owing to incidental injury or disease, a blood vessel may be seen invading the cornea, as is represented on one side of Fig. 13. In an early grade of keratopathy there is characteristically a clear zone round such a vessel and its branches similar in width to the marginal clear zone.

All observers who have studied the disease in Labrador have no doubt that it is caused by exposure to a climatic factor. Demographic evidence (to be published separately) points to ultraviolet light as the essential agent. If the droplets were being formed from a structural component of the corneal stroma such as collagen, it would be expected that a uniform band of degeneration would extend across the whole width of the cornea even in the early stages. It might be expected that the same argument would apply to other theories of the origin of the material, such as from the turnover of the protein of the basement membrane or its derivation from disturbed kerocytes.

Another reason why the droplets cannot be formed from collagen without the accretion of extraneous protein material is their amino-acid composition. Type I collagen, of which the corneal stroma is composed, contains no tryptophan, cysteine, or cystine, very little tyrosine, and fairly small amounts of histidine. Methionine is present as less than 6 residues per 1000 (Lian et al., 1973). Tryptophan, tyrosine, and sulphur-containing amino-acids have all been demonstrated in considerable amounts in the droplets by other authors. The amino-acids which absorb ultraviolet light and undergo chemical degradation most easily are tryptophan and histidine, followed by cysteine, methionine, and tyrosine. As proteins go, therefore, collagen must be one of the most inert to ultraviolet. It is also extremely unlikely that changing the molecular configuration of collagen would make it dense enough, unstained, by electron micrography to be consistent with our result if it were the main component of the droplets.
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acted upon by light until they have travelled a measurable distance from the limbal vessels.

(2) Alternatively, protein which has undergone the initial stages of transformation may be re-absorbed from this zone before it can be precipitated.

(3) It is also possible that the rather clear-cut outer edge of the area of droplet deposition may coincide with the peripheral termination of Bowman's membrane. Unless it could be shown, however, that Bowman's layer was destroyed round an invading blood vessel, the clear zones observed round such vessels argue against this third explanation. In addition, some droplets were seen by the immunoperoxidase method to be present in an area peripheral to the clear-cut termination of Bowman's layer, even in grade I, and are evidently not dependent on some physical property of this membrane in order to accumulate.

The idea that the abnormal spheroidal deposits are formed from plasma constituents present in the cornea and subconjunctival tissue under normal conditions is attractive in that it offers explanations of how similar material could also be laid down beneath the conjunctiva and in the cornea in a variety of intraocular diseases, and with advancing age, as well as under the influence of external climatic factors.

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