Topical immunotherapy for pseudomonas keratitis in rabbits: use of antilipopolysaccharide plasma

N. H. WELSH,¹ A. J. RAUCH,² AND S. L. GAFFIN³

From the ¹Departments of Ophthalmology and ²Physiology, University of Natal, Medical School, Durban, South Africa

SUMMARY Pseudomonas keratitis is currently treated with antibiotics with a variable success rate. Part of the morbidity caused by pseudomonas is due to the action of lipopolysaccharide (LPS) present on the surface membrane of the bacteria. Specific IgG present in equine anti-LPS hyperimmune plasma has been found to bind to the LPS from a range of Gram-negative bacteria, including pseudomonas, and by activating complement it destroys these bacteria. Anti-LPS plasma was therefore used as a therapeutic agent in experimentally induced pseudomonas keratitis in rabbits. Thirty out of 15 (86.7%) anti-LPS treated eyes improved, whereas four out of 17 (23.5%) saline treated control eyes improved (χ²=12.76 p<0.001). No ill effects were noted when anti-LPS was administered to healthy rabbit or baboon eyes. Anti-LPS thus was protective in pseudomonas keratitis, and clinical trials appear to be warranted.

Pseudomonas aeruginosa is responsible for severe keratitis that may progress to panophthalmitis. It is particularly disastrous when it occurs as a postoperative complication of invasive surgery—for example, penetrating keratoplasty. Current therapy is based on antibiotics with antipseudomonal activity, such as gentamicin and tobramycin.¹ High systemic doses are required to obtain adequate intraocular drug levels. The use of these agents is associated with a high incidence of side effects such as ototoxicity and nephrotoxicity.²

The pathophysiology of pseudomonas infections involves a number of components and products of the bacterial cell.³ Enzymes are produced which have a destructive effect on the host cell—for example, proteases and elastases. The lipopolysaccharide (LPS, endotoxin) component of the cell wall has also been implicated in the disease.⁴ This has been shown to be the cause of non-infectious corneal ring formation, mediated by complement activation.⁵ Antibiotics exert a direct antibacterial effect only, and thus immunotherapy directed at other aspects of the condition has enjoyed increased attention.⁶

Gaffin and colleagues have developed an equine anti-LPS hyperimmune plasma containing specific IgG which binds to free LPS and also has bactericidal properties.⁷ This plasma has been used successfully to treat a variety of Gram-negative infections involving LPS in horses, dogs, cats, sheep, and rats.⁸ The diversity and cross-reactivity of the antibodies in the preparation enable it to kill a wide range of Gram-negative bacteria, including pseudomonas, klebsiella, Escherichia coli, proteus, shigella, and salmonella. Such serum or plasma has recently proved effective in the prophylaxis and therapy of endotoxaemia in the veterinary field, particularly the race horse industry of South Africa.⁹

This study has been conducted to determine whether anti-LPS hyperimmune equine plasma (anti-LPS) would be effective in treating corneal pseudomonas infections. Anti-LPS was first tested against in-vitro cultures of Pseudomonas aeruginosa and found to be bactericidal. The serum was then found safe to use topically on normal rabbit eyes, as a lavage. The anti-LPS was then used therapeutically on experimentally induced pseudomonas keratitis in rabbits.

Materials and methods

An untyped culture of Pseudomonas aeruginosa was obtained from a microbiology department of a university hospital. It was cultured in nutrient broth for
24 hours to a final concentration of 600–900 million organisms/ml.

Anti-LPS was prepared from horses by plasmapheresis, suitably immunised (Atox Pharmaceutical Co., 14 Old Main Road, 3600 Gillitts, South Africa). The plasma contained 1500 µg/ml of LPS precipitable antibodies. These antibodies could bind to endotoxins prepared from *Shigella flexneri*, five strains of *E. coli*, five species of *Salmonella*, *Klebsiella*, *Proteus*, and *Pseudomonas*.

Rabbits of mixed, non-inbred, strain, weighing 2·5–3·5 kg were used. All had normal corneas and anterior segments. The corneas were anaesthetised locally with 3 drops of oxybuprocaine HCl prior to inoculation.

**METHOD OF INOCULATION**

Two methods were used to show that anti-LPS was effective in both moderate (group A) and severe infections (group B).

**Group A.** Both corneas of rabbits were prepared by making three deep vertical and horizontal incisions into the stroma, in a cross-hatch pattern, with a sterile 20 gauge needle, according to Furguele. The epithelium was removed in a 5 × 5 mm area. Four drops of the bacterial inoculum were dropped on to the cornea and allowed to remain in contact with the cornea and inferior fornix for 30 seconds.

**Group B.** Rabbit eyes were infected with contaminated sutures. A virgin silk suture (8/0) was soaked for five minutes in a broth culture of *Pseudomonas*. The suture was then passed through the corneal stroma. The ends were cut, and the sutures were allowed to remain in the cornea for two days. The lids were not sutured together.

After inoculation the animals were examined daily. The eyes were photographed at the beginning and end of treatment and the lesions were assessed on slit-lamp biomicroscopy. The severity of the infection present was graded according to the following criteria: (a) area of cornea involved; (b) depth of lesion; (c) degree of vascularisation.

**Table 1 Method of scoring severity of experimental pseudomonas keratitis**

<table>
<thead>
<tr>
<th>% Surface of cornea affected</th>
<th>grade (a)</th>
<th>Density and depth</th>
<th>grade (b)</th>
<th>vascularisation</th>
<th>grade (c)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>(a) 0</td>
<td>None</td>
<td>(b) 0</td>
<td>None</td>
<td>(c) 0</td>
</tr>
<tr>
<td>1–33</td>
<td>(a) 1</td>
<td>Mild macula</td>
<td>(b) 1</td>
<td>Mild</td>
<td>(c) 1</td>
</tr>
<tr>
<td>34–66</td>
<td>(a) 2</td>
<td>Moderate leucomatous</td>
<td>(b) 2</td>
<td>Moderate</td>
<td>(c) 2</td>
</tr>
<tr>
<td>67–100</td>
<td>(a) 3</td>
<td>Severe leucomatos</td>
<td>(b) 3</td>
<td>Severe</td>
<td>(c) 3</td>
</tr>
</tbody>
</table>

Maximum score totally all factors a+b+c=9.

This system was modified from that of previous workers. Each grade varied from 0 to 3 points. Thus (a) 0, indicated no infection, (a) 1, up to 33% corneal area involved, (a) 2, 33 to 66%, (a) 3, 66 to 100% affected. If the opacification was nebulous, the score was (b) 0, if it was macula (b) 1, if it was moderate leucomatos, (b) 2, severe leucomatus (b) 3. If the cornea perforated, this was denoted as (b) 3p. Finally, if no vascularisation was present, it would be (c) 0, if mild (c) 1, if moderate (c) 2, if severe, (c) 3. Therefore a moderate keratitis involving 50% of the cornea, with mild opacity, and some vascularisation would be scored as (a) 2 (b) 1 (c) 1. The most severe stage would be (a) 3 (b) 3p (c) 3. In assessing overall severity the points were added together, so that a maximum of 9p was obtainable (Table 1).

A masked randomised trial was initiated. The rabbits were treated as follows. One infected eye received anti-LPS, and where possible (infections established in both eyes) the contralateral eye served as a control, receiving saline. Anti-LPS or saline was administered as a lavage at the rate of 40 drops/minute for 5 minutes 3 times a day. The treated and control eyes were chosen at random. Treatment started as soon as the infection was clearly established, that is, after two days. The final appearance of the eyes was evaluated after eight days, by which time, anti-LPS had neutralised the effect of the pseudomonas.

Since we wished to determine merely whether the serum could control and limit the infection, the final result would not necessarily be a reversal of the lesions but a quiet eye with scar formation. In five severe cases topical corticosteroid drops (dexamethasone disodium phosphate) were added to this regimen after eight days, as it has been reported that steroids can cause a recurrence of the infection.

Steroids were also administered to five control eyes.

**Results**

In group A rabbits an infection developed in 22 out of 32 eyes inoculated. It developed in two days or less and varied in severity. In group B a severe infection developed in 10/10 eyes inoculated.

Table 2 shows the results of treatment with anti-
keratitis and some with partial healing. Of the 17 saline treated infected eyes (23.5%) improved, 9 deteriorated, and 4 remained unchanged. The unchanged eyes were already in the most severe group, so further deterioration could only have led to perforation.

Table 3 shows the changes that occurred on the 1st, 4th, and 8th days. Five out of the 17 saline treated control eyes in groups A and B perforated by day 8, 4/17 improved spontaneously, 4 remained unchanged. Among anti-LPS treated eyes only 1 deteriorated, and 1 was unchanged.

Table 4 shows the complete scores on day 1 and day 8.

Fig. 1A shows the eye of rabbit 7 before treatment, and Fig. 1B shows the same eye after eight days of anti-LPS treatment. Five serum-treated eyes that received topical corticosteroid after the 8th day showed dramatic improvement, and three of the control eyes continued to deteriorate. These are not indicated in the tables.

Discussion

Our experimental model was similar to previous models used to assess the different modes of treatment for pseudomonas keratitis. Where possible the anti-LPS treated eye was compared with the contralateral eye of the same animal treated with saline. The eyes were allocated for treatment in a randomised

Table 4 Clinical evaluation of pseudomonas keratitis before and after treatment

<table>
<thead>
<tr>
<th>Control</th>
<th>1st day</th>
<th>8th day</th>
<th>Treated eye</th>
<th>1st day</th>
<th>8th day</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Surface</td>
<td>Depth</td>
<td>Vessels</td>
<td>Surface</td>
<td>Depth</td>
</tr>
<tr>
<td></td>
<td>(a)</td>
<td>(b)</td>
<td>(c)</td>
<td>(a)</td>
<td>(b)</td>
</tr>
<tr>
<td>Group A</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
</tr>
<tr>
<td>2</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
</tr>
<tr>
<td>3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
</tr>
<tr>
<td>4</td>
<td>2 2</td>
<td>1 1</td>
<td>0*</td>
<td>2 2</td>
<td>1 1</td>
</tr>
<tr>
<td>5</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
</tr>
<tr>
<td>6</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
</tr>
<tr>
<td>7</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
</tr>
<tr>
<td>8</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
</tr>
<tr>
<td>9</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
</tr>
<tr>
<td>10</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
</tr>
<tr>
<td>11</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
</tr>
<tr>
<td>12</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
<td>3 3</td>
</tr>
<tr>
<td>Group B</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>13</td>
<td>3 2 3</td>
<td>3 2 3</td>
<td>3 2 3</td>
<td>3 2 3</td>
<td>3 2 3</td>
</tr>
<tr>
<td>14</td>
<td>3 2 3</td>
<td>3 2 3</td>
<td>3 2 3</td>
<td>3 2 3</td>
<td>3 2 3</td>
</tr>
<tr>
<td>15</td>
<td>3 2 3</td>
<td>3 2 3</td>
<td>3 2 3</td>
<td>3 2 3</td>
<td>3 2 3</td>
</tr>
<tr>
<td>16</td>
<td>3 2 3</td>
<td>3 2 3</td>
<td>3 2 3</td>
<td>3 2 3</td>
<td>3 2 3</td>
</tr>
<tr>
<td>17</td>
<td>3 2 3</td>
<td>3 2 3</td>
<td>3 2 3</td>
<td>3 2 3</td>
<td>3 2 3</td>
</tr>
</tbody>
</table>

*Indicates controlled eyes that improved. †Indicates treated eyes that deteriorated or did not improve. 3p Indicates perforation.
Topical immunotherapy for pseudomonas keratitis in rabbits: use of antilipopolysaccharide plasma

10/15 eyes the plasma was used on eyes severely infected—that is with morbidity indices of six or more —and the eyes responded after 3–4 days. The improvement in the corneal infection involved all three parameters measured, area of keratitis, depth of lesion, and degree of vascularisation.

In milder cases the plasma was effective with reversal of the lesion, whereas in severe cases the plasma effectively brought the infection under control, and the final result was a firm scar (Table 3). It would be advantageous to initiate therapy at the earliest sign of clinical infection, but in this experimental analysis we wished to determine the plasma’s efficacy against established infections.

The purely physical action (lavage) of instillation of solutions on the corneal surface has been shown to be beneficial by removing necrotic tissue and the exoenzymes produced by pseudomonas. Therefore saline was administered in the same manner to the control eyes. It would thus seem that the dual action of anti-LPS (bactericidal and antitoxic) is effective in pseudomonas keratitis. Significantly, no eyes in the anti-LPS treated group perforated. Bohigan et al. have shown that, in contrast to the ‘melting’ character of pseudomonas keratitis in man, the condition in rabbits follows an ulcerative, necrotic course, such that the addition of collagenase inhibitors to the normal regimen had no significant effect on the clinical course.

We believe that the addition of anti-LPS could significantly change the corneal stromal reaction. In another study, with carbenicillin and gentamicin, it was found that the keratitis continued to worsen for up to seven days before improvement was noted. In a different experimental model anti-LPS has already been observed to be more effective therapeutically than gentamicin. In our study with anti-LPS, improvement commenced within three days. Experiments in progress show that anti-LPS is also effective in similar infections in guinea-pigs.

The dosage regimen used was arbitrary (40 drops/min for 5 minutes 3 times daily for 8 days). We have yet to determine whether this may be modified or simplified. Treatment by the subconjunctival route will also be investigated, as preliminary results indicate that this method is tolerated by the normal rabbit eye.

A possible complication of anti-LPS treatment could be a systemic reaction to foreign equine antigens. However, this has not occurred in other experimental models where anti-LPS has been used, or in veterinary clinical practice.

The addition of topical corticosteroid after eight days resulted in an additional dramatic decrease in superficial vascularisation and a reduction in the area of scarring. The serum treatment does not therefore

---

Fig. 1A  Rabbit eye infected with Pseudomonas aeruginosa. Note extensive opacification and infection.

Fig. 1B  Same eye after eight days of anti-LPS therapy. The extent of keratitis and scarring is much reduced with control of infection.
contraindicate subsequent use of steroids, and indeed this seems to be beneficial.

We believe that the use of anti-LPS in the treatment of pseudomonas keratitis in rabbits shows favourable results, with none of the disadvantages of conventional antibiotic therapy, and that it has a potential application in the therapy, and perhaps prophylaxis, of the condition in man.

This work was supported by grants from the Anglo-American Corporation and the South African Medical Research Council.

References

Topical immunotherapy for pseudomonas keratitis in rabbits: use of antilipopolysaccharide plasma.

N. H. Welsh, A. J. Rauch and S. L. Gaffin

doi: 10.1136/bjo.68.11.828

Updated information and services can be found at:
http://bjo.bmj.com/content/68/11/828

**Email alerting service**

Receive free email alerts when new articles cite this article.
Sign up in the box at the top right corner of the online article.

**Notes**

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/