Pseudoexfoliative disease: histochemical evidence of an affinity with zonular fibres

A. GARNER AND R. A. ALEXANDER
From the Department of Pathology, Institute of Ophthalmology, University of London

SUMMARY The histochemical staining properties of the abnormal material deposited in the anterior segment of the eye in the pseudoexfoliation syndrome and the zonular ligaments of the lens are essentially the same. Both present the staining characteristics of oxtalatan, the microfibrillar component of elastic tissue. Reasons are advanced for regarding the pseudoexfoliative material as a product of the ciliary and lenticular epithelium.

Despite having been a recognised entity for over 60 years the essential nature of pseudoexfoliative disease in the anterior segment of the eye is still obscure. This lack of understanding is reflected in the less than satisfactory names that have been given to it. Though it was initially regarded as an exfoliation of the lens capsule, comparison with the true exfoliation occurring as an occupational hazard in workers exposed to intense radiant heat, such as glass-blowers, led Dvorak-Theobald to question this assumption and introduce the term pseudoexfoliation. Indeed the first histological reports concerning the 'exfoliated' material by Busacca had already given cause to doubt a lens capsule origin, and Gifford suggested a derivation from the zonular ligaments of the lens. It was hoped that the application of electron microscopy would supplement the limited information obtained from conventional light microscopy, and it has done so but without resolving the essential nature of the pseudoexfoliated material. Possibilities which currently command some support include amyloid formation, basement membrane exfoliation, and zonule degeneration, but, as Dark and Streiten, observe, none has unequivocal substantiation.

The purpose of the present communication is to describe some histochemical findings which point to an affinity between the pseudoexfoliative material and the essential protein component of the lens zonules.

Material and methods
Tissue from eight patients aged between 75 and 88 years, with pseudoexfoliative disease diagnosed on the basis of histological examination at the Institute of Ophthalmology between 1980 and 1982, was subjected to a number of histochemical procedures concerned principally with the detection of elastic and related fibres. The specimens had previously been fixed in formol-saline and embedded in paraffin wax. Sections cut at 5 µm were stained by the following techniques:

Verhoeff's iron-haematoxylin to demonstrate mature elastic tissue.
Aldehyde-fuchsin to demonstrate elastic fibres.
Aldehyde-fuchsin after exposure to an oxidising agent (Caroat, the active principle of which is potassium peroxymonosulphate, supplied by Degussa Ltd., Cheadle Hulme, Cheshire) to identify the microfibrillar component of elastic, i.e., oxtalatan.

Chrome haematoxylin modified to substitute Caroat as the oxidising agent in place of potassium permanganate advocated in Gomori's original method.
Gomori's trichrome stain.
Combined alcian blue period acid Schiff sequence.
Congo red for amyloid.
Thioflavine T.
Haematoxylin and cosin.

The complete anterior segment was present in five cases. In two lensectomy had previously been performed on account of cataract formation and in one the lens alone was available.

Results
The staining responses, recorded separately for the pseudoexfoliative material, the zonules and the lens capsules, are detailed in Table 1.
Table 1  Histochemical staining responses for pseudoexfoliative material, zonules, and lens capsule

<table>
<thead>
<tr>
<th>Tissue</th>
<th>Case no.</th>
<th>Verhoeff elastic</th>
<th>AF</th>
<th>Ox/AF</th>
<th>Chrome haem.</th>
<th>Gomori trichrome</th>
<th>AB/PAS</th>
<th>Congo red</th>
<th>Thioflavine</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pseudo-exfoliative material</td>
<td>1</td>
<td>-</td>
<td>±</td>
<td>+</td>
<td>+</td>
<td>G</td>
<td>AB&lt;PAS</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>G</td>
<td>AB&lt;PAS</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>3</td>
<td>-</td>
<td>±</td>
<td>++</td>
<td>+</td>
<td>G</td>
<td>AB&gt;PAS</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>4</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>G</td>
<td>AB</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>5</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>G</td>
<td>AB&lt;PAS</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>6</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>G</td>
<td>AB&lt;PAS</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>7</td>
<td>-</td>
<td>±</td>
<td>++</td>
<td>+</td>
<td>G</td>
<td>AB&lt;PAS</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>8</td>
<td>-</td>
<td>±</td>
<td>++</td>
<td>+</td>
<td>G</td>
<td>AB&lt;PAS</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Zonule</td>
<td>1</td>
<td>-</td>
<td>-</td>
<td>++</td>
<td>+</td>
<td>G</td>
<td>PAS</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>-</td>
<td>-</td>
<td>++</td>
<td>+</td>
<td>G</td>
<td>PAS</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>3</td>
<td>-</td>
<td>±</td>
<td>++</td>
<td>+</td>
<td>G</td>
<td>AB&lt;PAS</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>4</td>
<td>-</td>
<td>0</td>
<td>++</td>
<td>+</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>5</td>
<td>-</td>
<td>-</td>
<td>++</td>
<td>G</td>
<td>G</td>
<td>AB&lt;PAS</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>6</td>
<td>-</td>
<td>-</td>
<td>++</td>
<td>G</td>
<td>G</td>
<td>AB&lt;PAS</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>7</td>
<td>-</td>
<td>-</td>
<td>++</td>
<td>G</td>
<td>G</td>
<td>PAS</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>8</td>
<td>-</td>
<td>-</td>
<td>++</td>
<td>G</td>
<td>G</td>
<td>PAS</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Lens capsule</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>3</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>G</td>
<td>AB&lt;PAS</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>4</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>G</td>
<td>PAS</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>5</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>±</td>
<td>G</td>
<td>PAS</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>6</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>G</td>
<td>PAS</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>7</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>G</td>
<td>PAS</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>8</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>G</td>
<td>PAS</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>

AF=aldehyde-fuchsin. Ox=oxidised section. AB/PAS=combined alcian blue/periodic acid Schiff. G=green. 0=not performed. *Focal staining.

PSEUDOEXFOLIATIVE MATERIAL.

Pseudoexfoliative material was present by definition in all eight cases, though the amount and distribution varied somewhat. Always cellular and eosinophilic, the essential morphology was of tufts or clumps or delicate fibrils aligned perpendicularly to the surface to which they were attached, giving a characteristic 'carpet tuft' or 'hoar frost' appearance (Fig. 1).

The material did not react with Verhoeff's iron haematoxylin, but in four cases (cases 1, 3, 7, and 8) it gave a weak reaction to aldehyde-fuchsin in non-oxidised sections. Prior oxidation by exposure to potassium peroxymonosulphate, however, resulted in the pseudoexfoliative material in every case reacting with aldehyde-fuchsin with marked enhancement in the cases which had otherwise given a weak response (Figs. 2–5). Chrome haematoxylin staining gave results similar to those with aldehyde-fuchsin on oxidised tissue, though generally less intense (Fig. 6). The Gomori trichrome stain produced a green

Fig. 1  Part of a ciliary process covered by tufts of pseudoexfoliative material. (Haematoxylin and eosin, ×245).

Fig. 2  The tufts of pseudoexfoliative material on the ciliary processes are stained for elastic after oxidation of the section. (Oxidation, aldehyde-fuchsin, ×100).
colouration. In alcian blue/periodic acid Schiff stained sections the pseudoexfoliative material appeared to have two components, namely, a magenta-stained core with a coating of an alcianophilic substance. In two instances, however, where the deposits were relatively sparse and small (cases 3 and 4), the periodic acid Schiff positive component was inconspicuous. Congo red staining was essentially negative for amyloid in that the faint pinkish colouration frequently observed did not exhibit green dichroism by polarising microscopy. Thioflavine T stained sections examined by blue and by ultraviolet light showed weak fluorescence under the filters appropriate to amyloid detection.

**ZONULAR FIBRES**

The staining responses of the zonules were virtually identical to those recorded for the pseudoexfoliative

---

**Fig. 3** Parallel section to that depicted in Fig. 2 shows no response to the elastic tissue stain in the absence of prior oxidation. (Aldehyde-fuchsin, ×100).

**Fig. 4** Deposits on the anterior surface of the lens capsule stain for elastic after oxidation although the capsule is unstained. (Oxidation, aldehyde-fuchsin, ×170).

**Fig. 5** Adjacent section showing negative staining for mature elastic tissue. (Aldehyde-fuchsin, ×170).
Pseudoexfoliative disease: histochemical evidence of an affinity with zonular fibres

material. A weak reaction to aldehyde-fuchsin in the absence of preoxidation was observed in two cases (cases 3 and 8), but uniformly intense reactions were seen after oxidation in all cases. Except in case 3, in which a mixed response to alcian blue and periodic acid Schiff was seen, the zonular fibres did not take up the alcian blue stain. The reactions with amyloid stains were identical to those noted in respect of the pseudoexfoliative material.

LENS CAPSULE
Of the six cases in which lens tissue was present one (case 3) showed staining of the posterior part of the capsule with aldehyde-fuchsin that was not increased by preoxidation. Another, case 8, gave a positive reaction to aldehyde fuchsin in the equatorial region after oxidation. In all other instances the response to this stain, with or without oxidation, was negative (Figs. 4 and 5). Save for one instance (case 5), where weak response was obtained, the chrome haematoxylin method gave uniformly negative results. The capsules stained well with periodic acid Schiff but were not responsive to alcian blue and were stained green by Gomori's trichrome method. Negative reactions were obtained with Verhoeff's iron haematoxylin, and the responses to Congo red and thioflavine T corresponded to those given by the pseudoexfoliative material.

IRIS AND CILIARY BODY
Deposits of pseudoexfoliative material were commonly found covering the pigment epithelium of the iris and sometimes formed a layer on its anterior surface (Figs. 7 and 8). Material staining with aldehyde-fuchsin after oxidation was also seen within the iridal stroma, particularly around blood vessels.
(Fig. 7). A spotty deposition of similarly staining material was prominent on the basal aspect of the non-pigmented epithelium of the ciliary processes (Fig. 9).

Discussion

The notion that pseudoexfoliative material might be similar in composition to the zonular fibres of the lens was first advanced by Gifford, but evidence to support this view has been slow to appear. Ultrastructural resemblance has been noted, both zonular fibrils and the fibrils of pseudoexfoliative material consisting of microfibrils with a diameter of 8–10 nm and a cross-banding at 50 nm intervals. Even so there are subtle differences between them relating to the presence of smaller subunits in the pseudoexfoliative material and the need for special fixatives to demonstrate the banding of the zonular fibrils. That morphological similarity does not necessarily imply identity is further emphasised by the observation that the ultrastructure of the pseudoexfoliative material has also been compared with that of amyloid.

Other studies relating to the nature of the zonular fibrils which are more helpful in the context of pseudoexfoliative material composition are those showing biochemical, histochemical, and immunological comparability between the zonules and the tubular microfibrillar component of elastic tissue. These findings underscore the ultrastructural similarity between the two categories of fibril first noted by Raviola.

In view of the close similarity between zonular fibrils and the microfibrillar component of elastic tissue, commonly referred to as oxytalan when occurring in isolation from the amorphous elastin component, and some morphological affinity between the zonules and the pseudoexfoliative material, it seemed to us appropriate to examine the possibility that oxytalan is the essential constituent of the pseudoexfoliative fibrils. The results of our study, based on the histochemical characteristics of oxytalan, appear to validate this hypothesis.

Oxytalan is defined in histochemical terms as a tissue component that will react with certain stains for elastic fibres provided the tissue has first been exposed to an oxidising agent but not otherwise. This requirement is met by the deposits identified on morphological grounds as pseudoexfoliative material in all eight cases studied in whatever anatomical location. It is interesting to note also that the affinity

Fig. 8 Comparable region of the iris showing negative staining for mature elastic tissue. (Aldehyde-fuchsin, ×170).

Fig. 9 Higher magnification of part of a ciliary process shown in Fig. 2 and for comparison with Fig. 1. As a result of the oxidative stage in the staining process some of the melanin in the pigmented epithelium has been bleached serving to enhance the punctate staining (arrows) in relation to the basal aspect of the non-pigmented epithelium. (Oxidation, aldehyde-fuchsin, ×245).
of the abnormal material for chrome haematoxylin noted by Dark and Streeten11 and confirmed in the present study involves oxidation prior to staining.

The sole difference of note in our study between the two fibrils was that the pseudoexfoliative material was coated with an alcanophilic layer that was not seen in respect of the zonules. This could mean that the fibrils in pseudoexfoliative disease are 'contaminated' with glycosaminoglycan, possibly to form a proteoglycan gel as proposed by Davanger.19 Alternatively the apparent absence of glycosaminoglycan in the zonules might be a fixation artefact, as Dark and Streeten11 indicate, but, even so, as these authors also comment, it points to subtle differences between the zonules and pseudoexfoliative deposits.

We could not detect any difference in fluorescence of thiouline T stained sections between lens capsule, zonules, and pseudoexfoliative fibres, which is in contradiction to the preferential staining of the abnormal deposits reported by Dark and Streeten.11 The weak fluorescence we observed in each of these structures together with the absence of dichroism in Congo red stained sections argues against an amyloid composition for either zonules or pseudoexfoliative material.

Does knowing the nature of the pseudoexfoliative material enable us to comment on its source? With this question in mind it is relevant to observe that oxytalan as defined by both histochemistry and ultrastructure occurs in proximity to a variety of tissues. Originally it was described in the loose connective tissue surrounding tooth sockets, in tendons, ligaments, vascular adventitia, and dermis,20 and in the context of the eye it has been recognised in the cornea,21,22 iris stroma,23 Bruch’s membrane, and sclera24 as well as the zonules.11,16 In some instances oxytalan fibres are a transient feature of the developing eye in the course of maturation to true elastic tissue, but in others, such as the cornea, its presence in adult life is always a manifestation of disease. Although the circumstances in which ocular oxytalan is seen vary widely, a feature common to several of the non-ocular conditions in which it develops is fluctuating mechanical stress. Thus it is prevalent in tooth sockets, tendons, and ligaments, while in the cornea it develops in situations such as post-traumatic scarring and keratoconus, where there is anatomical distortion.24 Furthermore, it is conceivable that the oxytalan manifested as pseudoexfoliative material could be the product of either lens or ciliary processes epithelium, these also being subject to variable mechanical stresses in accordance with the demands of accommodation. On the basis that the ciliary epithelium is the source of the oxytalan tissue of the normal zonular fibrils formed in utero it is not difficult to imagine that this capacity might be reawakened in later life if the appropriate stimulus was provided.

The activation of a latent capacity on the part of the equatorial lens epithelium would explain the subcapsular accumulation of pseudoexfoliative oxytalan.

The finding of particulate material with the staining properties of oxytalan in relation to the basement membrane of the non-pigmented ciliary epithelium is in conformity with ultrastructural observations10–24 and possibly lends some support to an epithelial origin, given that fibril assembly is essentially an extracellular event. Where oxytalan appears to be a product of cells which normally form a basement membrane, such as the corneal epithelium in keratoconus25 and the corneal endothelium in Fuchs’s dystrophy,22 the fibrils are deposited within or adjacent to the membrane. To this extent the evidence that pseudoexfoliative material is basically a pre-elastic moiety does not altogether contradict the suggestion first advanced by Trantas25 and most recently espoused by Eagle et al.10 that the pseudoexfoliation syndrome is a basement membrane disorder.

A true exfoliation of the zonules as proposed by Gifford seems unlikely, because on the one hand no clinical or histological evidence of zonule disintegration has been forthcoming, and on the other such an origin would not account for the observed presence of the abnormal deposits beneath the lens capsule and close to the basement membrane of the ciliary epithelium.

Until a consensus seems probable, it is perhaps advisable, as Dark and Streeten11 suggest, to retain the term pseudoexfoliative disease. But, if our findings are accepted, some such term as oxytalanosis of the aqueous might find approval.

References


Pseudoexfoliative disease: histochemical evidence of an affinity with zonular fibres.
A Garner and R A Alexander

doi: 10.1136/bjo.68.8.574

Updated information and services can be found at:
http://bjo.bmj.com/content/68/8/574

Email alerting service
Receive free email alerts when new articles cite this article.
Sign up in the box at the top right corner of the online article.

Notes

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/