Quantitative assessment of macular thickness in normal subjects and patients with diabetic retinopathy by scanning retinal thickness analyser

Yusuke Oshima, Kazuyuki Emi, Shigeki Yamanishi, Masanobu Motokura

Abstract

Aims—To evaluate the scanning retinal thickness analyser (RTA), a novel non-invasive imaging instrument, in diagnosing and quantitatively characterising diabetic macular oedema, and to investigate the relation between central macula thickness measured by RTA and other clinical examinations.

Methods—Central macular thickness was measured using the RTA in 40 normal subjects and 60 patients with diabetic retinopathy. The reproducibility of the retinal thickness measurements was evaluated by calculating the mean of the inter- and intra-session variations. Central macular thickness was correlated with the results of visual acuity measurements, biomicroscopy, and fluorescein angiography.

Results—Intra- and inter-session reproducibility of the RTA in normal subjects was plus or minus 5.2% (16 µm) and plus or minus 6.1% (19 µm), respectively. The mean central macular thickness was 182 (SD 16) µm in normal subjects, 283 (116) µm in diabetic eyes without clinically significant macular oedema (CSMO), and 564 (168) µm in diabetic eyes with CSMO. Central macular thickness was significantly greater (p<0.001) in eyes with diabetic retinopathy than in normal subjects, even when macular thickening did not meet the standard for CSMO (p=0.019) measured by biomicroscopy. Although greater fluorescein leakage at the macula results in greater central macular thickness, only eyes with diffuse leakage had statistically significant macular thickening compared with normal subjects (p=0.022). Central macular thickness measured with the RTA was significantly correlated with the logarithmic converted visual acuity (r²=0.76) in diabetic eyes.

Conclusion—Scanning RTA, which has good reproducibility, might be useful to quantitatively detect and monitor macular thickening in diabetic retinopathy. Central macular thickness was highly correlated with logarithmic converted visual acuity in diabetic macular oedema.

Diabetic macular oedema, a common cause of central visual loss in patients with diabetic retinopathy, usually results from the breakdown of the blood-retinal barrier, which leads to abnormal fluid accumulation in the retinal layer and increased retinal thickness. Although early detection and diagnosis are important to successfully treat diabetic macular oedema, the clinical detection and evaluation methods currently used are limited to slit lamp biomicroscopy, stereoscopic fundus photography, and fluorescein angiography, which provide only a subjective evaluation of retinal thickness. Although fluorescein leakage from the perifoveal capillary network indicates the breakdown of the blood-retinal barrier at the macula, angiographic findings are neither a reliable nor a quantitative indicator of the degree of fluid accumulation and retinal thickening. Subjective observations using slit lamp biomicroscopy and/or fundus stereoscopy cannot quantify the subtle changes of macular thickening.

Recently, several new techniques to measure quantitatively the retinal thickness have been explored. The scanning retinal thickness analyser (RTA), a novel instrument developed for non-invasive, multiple optical cross-sectional retinal visualisation that is based on the principle of laser biomicroscopy, provides objective and quantitative measurements of retinal thickness. The distance between the images of the anterior and posterior retinal intersections is calculated by an analysis algorithm to produce accurate retinal thickness mapping. Zeimer and associates and Asrani and colleagues in their early experience with the RTA, reported its potential for clinical use. Landau and associates reported their in vivo measurements of retinal thickness in healthy volunteers. At present, however, few studies of quantitative retinal thickness measurements have been reported in patients with diabetic retinopathy.

In this prospective study, we used a commercially available, rapid scanning RTA with a newly developed analysis algorithm to examine the macular thickness in age and race matched groups of normal subjects and patients with diabetic retinopathy. We evaluated the clinical reproducibility of this new instrument and investigated its potential as an objective test to diagnose and quantitatively measure the central macular thickness in diabetic retinopathy.
Macular thickness measurement of diabetic macular oedema by RTA

LogMAR = logarithm of the minimum angle of resolution.
Goldmann contact lens, fundus photography, lamp biomicroscopy with a 90 dioptre lens or fundus examination was performed by slit and retinal thickness measurement. A detailed fluorescein angiography (patient group only); clinical examination; fundus photography; assessments: corneal refractive power, refractive error, Axial length (mm) 24.21 (0.67) 24.55 (0.83) Range −1.75 to +1.50 −1.50 to +2.25 LogMAR (SD) 0.08 (0.52) 3.78 (3.61) *Refractive error is shown in spherical equivalent. LogMAR = logarithm of the minimum angle of resolution.

Forty age matched individuals (22 men and 18 women) volunteered to participate as normal control subjects. The corrected visual acuities of this group ranged from 20/25 to 20/20. With the exception of the corrected visual acuities, all variables (age, sex, refractive error, and race) were well matched in both groups. Normal subjects were excluded if they had a history of diabetic mellitus or other ocular disease except refractive error. The demographic data of the study participants are shown in Table 1.

In the normal subjects, one eye of each volunteer was randomly selected for retinal thickness measurement. In patients with diabetic retinopathy, the study eye also was randomly selected to assure the independence of data analysis when both eyes met the criteria. Before the study, a full explanation of the procedure was given to all participants and informed consent was obtained in all cases.

**Table 1** Demographic data for normal subjects and patients

<table>
<thead>
<tr>
<th>Group-size (no)</th>
<th>Normal subjects</th>
<th>Patients with diabetic retinopathy</th>
</tr>
</thead>
<tbody>
<tr>
<td>Female/male</td>
<td>22/18</td>
<td>33/27</td>
</tr>
<tr>
<td>Age (years)</td>
<td>53.2 (15.8)</td>
<td>55.7 (10.1)</td>
</tr>
<tr>
<td>Range</td>
<td>25 to 69</td>
<td>24 to 73</td>
</tr>
<tr>
<td>Refractive error (D)*</td>
<td>-0.73 (1.78)</td>
<td>-0.05 (1.34)</td>
</tr>
<tr>
<td>Range</td>
<td>-1.75 to +1.50</td>
<td>-1.50 to +2.25</td>
</tr>
<tr>
<td>Axial length (mm)</td>
<td>22.41 (0.67)</td>
<td>24.55 (0.83)</td>
</tr>
<tr>
<td>Range</td>
<td>22.87 to 25.54</td>
<td>23.17 to 25.87</td>
</tr>
<tr>
<td>LogMAR (SD)</td>
<td>0.08 (0.52)</td>
<td>3.78 (3.61)</td>
</tr>
</tbody>
</table>

*Refractive error is shown in spherical equivalent.

LogMAR = logarithm of the minimum angle of resolution.

The examination included the following assessments: corneal refractive power, refractive error, and best corrected visual acuity; a clinical examination; fundus photography; fluorescein angiography (patient group only); and retinal thickness measurement. A detailed fundus examination was performed by slit lamp biomicroscopy with a 90 dioptre lens or Goldmann contact lens, fundus photography, fluorescein angiography using a 35 degree field fundus camera, and retinal thickness measurement using a RTA was performed in all patients with diabetic retinopathy.

The clinical diagnosis of diabetic retinopathy was based on the results of a detailed slit lamp biomicroscopic examination and fluorescein angiography. The severity of diabetic retinopathy was classified by means of a fundus examination and photography according to the criteria of the Early Treatment Diabetic Retinopathy Study (ETDRS)\(^{15}\)—that is, non-proliferative diabetic retinopathy (levels 20–53) or proliferative diabetic retinopathy (level 61 or above). To evaluate the differences between the study groups, the presence of clinically significant macular oedema (CSMO) was determined, and the fluorescein angiographic findings were graded before the macular thickness measurements were undertaken. CSMO was diagnosed when one or more of the following were detected: retinal thickening involving the centre of the macula or within 500 µm of the centre; hard exudate or exudates at or within 500 µm of the centre of the macula, when associated with thickening of the adjacent retina; and a zone or zones of retinal thickening one disc area or larger, with any part of the retinal thickening within 1 disc diameter of the centre of the macula.\(^{21}\) To evaluate the effect of the breakdown of the blood-retinal barrier on the perifoveal capillary network, the degree of leakage on the fluorescein angiograms at the macula was graded using a modified ETDRS protocol (Table 2).

Placing a transparent, size matched, plastic mask with concentric rings over the early phase angiograms, the innermost ring was centred over the foveal avascular zone to grade the leakage from the perifoveal capillary network. Each diabetic patient was clinically examined by one of two retinal specialists (KE, MM) who were masked to the results of the retinal thickness assessment. The angiographic findings were graded by one of us (SY), who did not contribute to the clinical grading portion and was masked to the results of the retinal thickness assessment.

**Scanning Retinal Thickness Analyser**

The scanning RTA (Telia Technology Ltd, Mevaseret Zion, Israel) is based on the principle of slit lamp biomicroscopy, as previously described.\(^{16–18}\) A green helium neon (534 nm) laser slit of approximately 400 µW was projected on the retina; the image was scanned by a slit beam and the light passing through the slit was detected by a photodetector and translated into a time dependent electronic signal. The time delay measured between the two maximum values of photodetector output, emanating from the vitreoretinal interface and

<table>
<thead>
<tr>
<th>Level</th>
<th>Definition</th>
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<tbody>
<tr>
<td>None</td>
<td>No dye leakage from perifoveal capillary network</td>
</tr>
<tr>
<td>Questionable</td>
<td>Questionable or &lt;25% dye leakage</td>
</tr>
<tr>
<td>Focal</td>
<td>&gt;25% but &lt;66% dye leakage</td>
</tr>
<tr>
<td>Diffuse</td>
<td>&gt;66% dye leakage</td>
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the chorioretinal interface, was proportional to the retinal thickness at the measured location. In the prototype scanning RTA used in the present study, the newest software based on a paradigm analysis algorithm (Version 6.30, Talia Technology Ltd) was developed to analyse the scan images automatically based on this principle.

For the retinal thickness measurement, the pupils were dilated to a minimum of 6 mm with a mixture of 0.5% phenylephrine hydrochloride and 0.5% tropicamide (Mydrin-P, Santen Pharmaceuticals, Osaka, Japan). The subjects were shown a target consisting of a pattern of numbers, and an infrared laser light emitting diode was used for monitoring the fixation (Fig 1A). After activation of a foot pedal, the laser slit scanned a retinal area $2 \times 2$ mm$^2$. Ten slit laser images (Fig 1B) covering this area were obtained within 200 ms. After performing this procedure nine times, an area $6 \times 6$ mm$^2$ covering the posterior pole was scanned. In each slit image, 10 points were analysed by an analysis algorithm (Version 6.30, Talia Technology Ltd). At each measured point, the algorithm considered the peaks on both sides of the densitometric reading curve (Fig 1C) to represent the reflections of the outermost and innermost retinal layers. The distance between the two peaks is proportional to the retinal thickness. A relative retinal thickness can be obtained by the following formula.

\[
\text{Relative Retinal Thickness} = \frac{\text{Distance between peaks}}{\text{Slit Image Width}}
\]
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\[ T = D \times M/Sb \]

where \( T \) = relative retinal thickness, \( D \) = the width of the two peaks calculated from the scan profile, \( M \) = magnification that changes depending on the refractive error of the individual, and \( Sb \) = stereo base.

Thus, the 10 slit images of each scan can be automatically analysed to generate a topographic map in numeric form—that is, 100 points \((10 \times 10)\) of relative retinal thickness values covering a retinal area \(2 \times 2 \text{mm}^2\) (Fig 1D). Given the calibration factor based on measurement of the corneal refractive power (dioptres) and refractive error (dioptres) of the study eye, the relative retinal thickness values were automatically converted to absolute micrometre units by the thickness correction software (Talia Technology Ltd).

Before measuring the macular thickness in the diabetic eyes, the intra- and intersession reproducibility of the RTA was evaluated in normal subjects by a single examiner (YO). To determine the intrasession reproducibility, three scans from the centre frame including the macula were evaluated in a single session in each of the normal subjects. One hundred points \((10 \times 10)\) were evaluated from each scan, and the coefficient of variance (standard deviation/mean) of the retinal thickness values in all three scans from each subject was calculated for each of the 100 points. To assess the intersession reproducibility, two sessions were carried out at 1 or 2 week intervals; three scans centred on the macula obtained from two different visits were evaluated for each subject. One hundred points \((10 \times 10)\) were evaluated from each scan, and the coefficient of variance (standard deviation/mean) of the retinal thickness values in all scans of each subject was calculated for each of the 100 points. The intrasession reproducibility also was assessed in the same way in the group of patients with diabetes.

CENTRAL MACULAR THICKNESS MEASUREMENT

Because an area of \(2 \times 2 \text{mm}^2\) obtained from a routine fovea centred scan using the RTA is too large to cover the foveal region in EDTRS type regions, which anatomically corresponds to approximately 500 \(\mu\text{m}\) in diameter, the average of 100 points \((10 \times 10)\) of the retinal thickness measured from the fovea centred scan (covering an area \(2 \times 2 \text{mm}^2\)) cannot directly represent the central macular thickness. In the central macular thickness assessment, a small area of nine points \((3 \times 3)\) including the fovea, corresponding to an area of \(600 \times 600 \mu\text{m}^2\), was chosen from a fovea centred scan and defined as the central macula (Fig 1D). Thus, the central macular thickness, which corresponds approximately to the retinal thickness at the foveal region in EDTRS type regions, was calculated as the mean value of the \(3 \times 3\) points in the frame acquisition mode obtained from a fovea centred scan image.

STATISTICAL ANALYSIS

Measurement of the central macular thickness was correlated with the visual acuity using linear regression. Visual acuity was measured in each eye using a Snellen visual acuity chart, and then converted to the logarithm of the minimum angle of resolution (logMAR). In this notation, a score of 0 corresponds to a visual acuity of 20/20 and each doubling of the visual angle increases the score by three units (that is, 20/40 = 3, 20/50 = 4, and 20/100 = 10).

To evaluate the differences between the study groups, the unpaired Mann-Whitney U test was used as the non-parametric equivalent of the \(t\) test for two independent samples. Spearman’s correlation coefficient was calculated to determine the statistical correlation between any two non-normally distributed variables. A \(p\) value smaller than or equal to 0.05 was considered significant.

Results

REPRODUCIBILITY AND CENTRAL MACULAR THICKNESS IN NORMAL SUBJECTS

The mean retinal thickness obtained from the \(10 \times 10\) measurements of the 40 normal eyes was \(312 (26) \mu\text{m}\). The mean reproducibility of the scans obtained at the same examination session was 5.2%, which corresponded to approximately plus or minus 16 \(\mu\text{m}\). The reproducibility of the points obtained from two examinations was plus or minus 6.1%, which corresponds to approximately plus or minus 19 \(\mu\text{m}\).

The macular thickness at the centre \((3 \times 3\) central points) in the 40 normal eyes is presented as a function of age (Fig 2). The average macular thickness was 182 (16) \(\mu\text{m}\). Linear regression of the macular thickness showed no statistically significant correlation between the macular thickness and age (Spearman’s correlation coefficient, \(r=0.12\); \(p=0.99\)). The macular thickness was also statistically independent of sex, and no difference was found between left and right eyes.

REPRODUCIBILITY AND CENTRAL MACULAR THICKENING IN DIABETIC RETINOPATHY

The mean retinal thickness obtained from the 60 eyes with diabetic retinopathy was \(428 (112) \mu\text{m}\). The mean reproducibility of the scans obtained at the same examination session was plus or minus 6.3%, which corresponded
to approximately plus or minus 27 µm. There was no statistically significant difference (p=0.24) in the intrasession reproducibility between the normal group and the patient group.

Macular thickening in diabetic retinopathy was well delineated on the optical section slit image of the scanning RTA (Figs 3 and 4). The mean central macular thickness of the 60 eyes with diabetic retinopathy was 446 (208) µm (range 156–906 µm), which was significantly (p<0.001) greater than that of the normal subjects.

**RELATION BETWEEN RTA, FLUORESCIN ANGIOGRAPHY, AND BIOMICROSCOPY**

Of the 60 eyes in the retinopathy group, stereo biomicroscopic examination revealed that 36 eyes (60%) had CSMO. The visual acuities in eyes with CSMO and in eyes without CSMO varied, respectively, and ranged from 20/400 to 20/40 and 20/60 to 20/20. Fluorescein angiography demonstrated that 21 of 60 eyes (35%) had diffuse leakage, and 18 eyes (30%) had focal leakage at the macular lesions. The results of the central macular thickness measurements using the scanning RTA are summarised in Table 3.

The relation between the distribution of central macular thickness and clinical evaluation with slit lamp biomicroscopy is shown in Figure 5. Although each subset had a wide range of retinal thickness values, the macular thickness was significantly (p=0.008) greater in

![Figure 3](image1)

(A) Fundus photograph obtained from the right eye of a patient with localised diabetic macular oedema. Optical sectioning was performed within the area bound by the square. The black arrowhead indicates the fixation target for the central scan, presumably corresponding to the centre of the fovea. (B) A composite of optical section images shows the presence of intraretinal cysts (arrowheads) at the macula that cause elevation of the vitreoretinal interface and loss of a normal foveal depression. The chorioretinal interface (vertical black arrows) is partly obscured because of intraretinal fluid accumulation, but is visualised in the superior and inferior retinal area (vertical white arrows).

![Figure 4](image2)

(A) Fundus photograph obtained from the left eye of a patient with diffuse diabetic macular oedema. The square delineates the scanned region demonstrated in the next optical section image. The black arrowhead indicates the fixation target for the central scan, presumably corresponding to the centre of the fovea. (B) Multiple cross sectional scanning image shows the relative elevation of the interretinal surface with gross foveal distortion. The vitreoretinal interface at the centre of the fovea (white arrows) showed an absent foveal depression.
Eyes with non-CSMO

14

18

10

21

16

CSMO = clinically significant macular oedema.

†Central macular thickness was measured using the scanning retinal thickness analyser.

unpaired Mann-Whitney U test.

*Central macular thickness compared with normal subjects. Statistical analysis was done using the

The positions of the medians (solid line within the boxes) illustrate the increasing macular thickness in eyes with increasing degrees of fluorescein leakage.

Figure 7 The relation between central macular thickness and visual acuity in eyes with diabetic retinopathy. The visual acuity is reported as the logarithm of the minimum angle of resolution (logMAR). Linear regression demonstrates a good correlation (n=60; r²=0.76; p<0.001) between the macular thickness and visual acuity. The equation for the linear correlation was based on the following formula: macular thickness (µm) = 195.8 + 48.7 × logMAR (r²=0.76).

Discussion

In the present study, the results indicate that for quantifying retinal thickness the scanning RTA is highly reproducible and has low intra- and intersession variability. Under standard conditions, the standard deviation of plus or minus 26 µm indicates that the scanning RTA detects retinal thickening of more than approximately 50 µm. The intersession reproducibility of plus or minus 5.2% and the intersession reproducibility of plus or minus 6.1%,
which correspond to approximately 16 and 19 µm, respectively, suggest that the scanning RTA can clinically screen changes in retinal thickening. The intra-session reproducibility in the patient group (plus or minus 6.3%) indicates that the scanning RTA also could detect the retinal thickness in eyes with diabetic retinopathy enrolled in the present study.

The mean central macular thickness in normal eyes measured in the present study was 182 µm, which is comparable to the results reported in two previous studies.18 19 The absolute values of macular thickness quantified by the scanning RTA are still uncertain because a comparison between the in vivo retinal thickness measurements and histological analysis has not been conducted. However, for the present, central macular thicknesses ranging from 150 to 214 µm (within twice the standard deviation of the average normal value) can be considered to be within the normal variations of the RTA measurement.

Data from the normal subjects in the present study showed slight variability in macular thickness. However, no statistically significant relation between macular thickness and aging was found. Figure 2 shows that the central macular thickness is independent of patient age. This result differs from the results reported by Landau and associates,19 in which the fovea thickness increased with age. Additional studies with more subjects are required to resolve this discrepancy.

A cross sectional study reported that macular oedema in various retinal disorders has no direct association with visual acuity.20 However, a previously published study evaluated macular thickening using only qualitative methods such as slit lamp biomicroscopic examination, fundus photography, or fluorescein angiography. With a quantitative measurement of the retinal thickness, foveal thickening was strongly associated with decreased visual acuity.21 In the present study, the central macular thickness measured using the RTA was found to be linearly correlated with visual acuity expressed on a logMAR scale (r²=0.76), which agrees with the results of studies that measured the macular thickness in retinal diseases using optical coherence tomography.22 23

Routine slit lamp biomicroscopy can provide information that is useful in the clinical diagnosis of macular oedema. However, the actual macular thickening, particularly in the early stage of macular oedema, is hard to estimate using slit lamp biomicroscopy. Shahidi and associates reported that clinical examination using slit lamp biomicroscopy permits the detection of retinal thickening only when it is more than 60% of the normal retinal thickness.24 In the present study, 24 eyes (40%) with various degrees of diabetic retinopathy were clinically diagnosed without CSMO. However, the macular thickness was significantly greater (p=0.019) in these eyes than in normal control subjects. When CSMO was found, the central macular thickness markedly increased to more than 350 µm based on our results. Slit lamp biomicroscopy cannot identify mild or localised macular thickening. Because biomicroscopy grading is based on a distinction of the local differences rather than comparisons with absolute normal values, it also is limited in detecting diffuse oedema as a total elevation of the macula. By quantitatively measuring the actual thickness, scanning RTA can more reliably detect changes in central macular thickness before CSMO develops.

Fluorescein angiography is traditionally used to assess retinal vascular permeability in patients with diabetic retinopathy. The degree of fluorescein leakage at the macula was generally considered to be associated with retinal thickening. However, our results show that this qualitative method cannot reliably detect changes in macular thickening. The wide variations in thickness values in each subgroup led us to propose that the degree of fluorescein leakage is not directly correlated with retinal thickening. Fluorescein leakage, which physiologically indicates the breakdown of the blood-retinal barrier, is not always accompanied by appreciable fluid accumulation in the retina.5 In a subclinical stage, slight dye leakage on fluorescein angiography could appear without retinal thickening because the balance between the vascular permeability and the ability of the retinal pigment epithelium to reabsorb fluid has been maintained. As retinopathy progresses, this balance is disturbed, resulting in fluid accumulation and macular oedema.8 Thus, fluorescein angiography is useful for evaluating the severity of the dysfunction in the blood-retinal barrier, but it does not reliably quantify the degree of fluid accumulation in the retina. The scanning RTA is superior to these conventional methods for detecting macular thickening because of multiple cross sectional imaging and the direct measurement of retinal thickness.

In our clinical experience with scanning RTA, one of the chief advantages of this tool over other quantitative instrument is its short acquisition time. Within the 200 ms required for scanning, patients have no discomfort caused by prolonged light exposure, the need for steady ocular fixation, or lack of blinking. Furthermore, multiple cross sectional retinal imaging, which generates three dimensional reconstruction of the retina, may contribute to a better understanding of retinal elevation. However, we are also aware of sources of error attributable to the instrument that can explain the variability observed in the present study. The most common reasons for inadequate image quality are the irregular reflections from hard exudates accumulated in the retina and the interference of the media opacities, such as severe cataract or vitreous opacity, with the scanning of the laser. Massive intraretinal exudation can prevent the relatively short wave length of the green helium neon (534 nm) from obtaining an accurate reflection from the chorioretinal interface, resulting in unclear laser slit images. Therefore, we still believe that this prototype RTA is limited in its ability to measure the central macular thickness, particularly in patients with extensive hard exudates that accumulated at the fovea, and thus
we excluded them from the present study. However, because this type of exudation is rarely found in the early stage of diabetic retinopathy, most types and various degrees of diabetic retinopathy could be effectively evaluated with this novel instrument as we showed in the present study.

In conclusion, this study suggests that the scanning RTA, which provides multiple cross sectional visualisation of the retina, is useful to measure quantitatively macular thickening in diabetic retinopathy. Although we do not recommend that scanning RTA replace any other conventional diagnostic tool used to detect macular oedema because of its limitations, we believe that scanning RTA may contribute to early, accurate diagnosis of macular thickening in diabetic retinopathy. However, further randomised studies with large populations are required to evaluate this tool for longitudinal monitoring of the disease prognosis and treatment efficacy.

The authors have no proprietary interests in any of the materials used in this study. Presented in part at the Annual Meeting of the Association for Research in Vision and Ophthalmology, Fort Lauderdale, Florida, May 1997.

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*Br J Ophthalmol* 1999 83: 54-61
doi: 10.1136/bjo.83.1.54