Expression of gelatinase B in trachomatous conjunctivitis

Ahmed M Abu El-Asrar, Karel Geboes, Soliman A Al-Kharashi, Abdulrahman A Al-Mosallam, Luc Missotten, Liesbet Paemen, Ghislain Opdenakker

Abstract

Background/aims—Gelatinase B is a matrix metalloproteinase involved in extracellular matrix (ECM) breakdown often associated with scarring and other pathological disorders. It was investigated whether gelatinase B is involved in the pathogenesis of ECM degradation associated with trachomatous conjunctivitis.

Methods— Conjunctival biopsy specimens obtained from six patients with active trachoma, six patients with active vernal keratoconjunctivitis (VKC), and seven control subjects were studied. Immunohistochemical techniques and a specific monoclonal antibody against human gelatinase B were used, and a monoclonal antibody against macrophage CD68 to identify mononuclear cells with gelatinase B immunoreactivity. In addition, quantitative zymography was used to compare the activity of gelatinase B in conjunctival biopsy specimens from seven patients with active trachoma and seven control subjects.

Results—Gelatinase B was detected by immunohistochemistry only in polymorphonuclear cells located in the vascular lumens in three normal conjunctival biopsy specimens. In all trachoma specimens and in five VKC specimens, gelatinase B was localised in monocyte/macrophage cells, positive for the CD68 marker, and in polymorphonuclear cells. The majority of the latter cell type was located in intravascular spaces. Compared with VKC specimens, trachoma specimens showed significantly more immunoreactive gelatinase B monocytic/macrophage cells (52.3 (21.9) vs 8.2 (6.4); p < 0.001) and polymorphonuclear cells (23.2 (14.2) vs 6.3 (5.4); p = 0.013). Activated macrophages with giant cell morphology clearly stained with the gelatinase B specific monoclonal antibody were observed in trachoma specimens. Zymography revealed that gelatinase B levels in trachoma specimens were significantly higher than the levels found in normal conjunctiva (1739.6 (1078.3) vs 609.3 (395.9) scanning units; p = 0.0127).

Conclusions— The increased activity of gelatinase B and numbers of inflammatory cells containing gelatinase B in trachoma specimens suggest that this enzyme plays a part in the pathogenesis of conjunctival scarring in trachoma.

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test this hypothesis, we examined conjunctival specimens obtained from patients with active trachoma using immunohistochemistry and gelatin zymography. The findings in trachoma were compared with the findings in the conjunctiva from normal individuals, and from patients with vernal keratoconjunctivitis (VKC) which is a non-scarring, chronic allergic inflammation.

**Patients and methods**

**IMMUNOHISTOCHEMISTRY**

Six patients with severe active trachoma ranging in age from 5 to 17 years (mean age 7.7 years) were included in the study. Six patients with severe active VKC ranging in age from 7 to 15 years (mean age 10.5 years) were studied.

VKC patients had the palpebral form of the disease characterised by the presence of giant polygonal flat topped cobblestone papillae affecting the upper palpebral conjunctiva. A 2×2 mm upper palpebral conjunctival biopsy specimen was obtained from each patient after informed consent. None of the patients was on topical therapy. In addition, seven upper palpebral conjunctival biopsy specimens were obtained from patients of the same age group who underwent cataract extraction or strabismus surgery without obvious inflammation and these served as controls.

The conjunctival biopsy specimens were immediately fixed for 2–3 hours in B5 fixative composed of: (A) 90 ml distilled water, 6 g mercuric chloride, 2.074 g sodium acetate; and (B) 37% formaldehyde solution, pH 5.7. Immediately before use 9 ml of A and 1 ml of B were mixed. The specimens were then embedded in paraffin. After trypsinisation for 7 minutes at 37°C using a mixture of 0.1% trypsin (Sigma-Aldrich, Bornem, Belgium) and 0.1% CaCl2 at pH 7.8 and blocking of endogenous peroxidase activity, the sections were stained with a three step avidin-biotin-peroxidase labelled complex, tertiary antibodies consisted of biotin conjugated with horseradish peroxidase-conjugated biotin complex (ABC) peroxidase-conjugated biotin complex (ABC) and 0.1% CaCl2 at pH 7.8 and blocking of endogenous peroxidase activity. The sections were stained with CD68 monoclonal antibody was used in the first step and 0.1% gelatin (Sigma Chemical Company, St Louis, MO, USA). The gels were then stained in Coomassie brilliant blue R-250 and destained in methanol/acetic acid. The sites of gelatinase activity appeared as unstained bands on a blue background.

**GELATIN ZYMOGRAPHY**

Seven children with asymptomatic mild to moderate active trachoma ranging in age from 7 to 16 years (mean age 9.4 years) were included in the study. A 2×2 mm upper palpebral conjunctival biopsy specimen was obtained from each patient after informed consent. None of the patients was on topical therapy. In addition, seven upper palpebral conjunctival biopsy specimens were obtained from patients in a similar age group undergoing strabismus surgery without obvious inflammation and served as controls.

The conjunctival biopsy specimens were immediately snap frozen in Tissue-Tek optimum cutting temperature (OCT) compound (Miles Laboratories, IN, USA) and maintained at −80°C until use. For gelatin zymography, frozen tissues were thawed and transferred into 50 µl of phosphate buffered saline, pH 7.4, supplemented with 1% Triton X-100. This preparation was sonicated at 0°C for 3 minutes and protein concentrations were determined on a fraction. For each specimen an equivalent amount of 100 µg protein was analysed as described by Masure et al. The localisation of gelatinolytic enzymes and their molecular masses was derived by including protein sizing standards on each gel and on the basis of the known migration of gelatinase B variants (monomer, dimer, and heterodimer with neutrophil gelatinase B). The reaction product was developed by the addition of diaminobenzidine and hydrogen peroxide resulting in brown immunoreactive sites. The slides were then counterstained with Harris haematoxylin. Finally, the sections were rinsed with distilled water and coverslipped with glycerol. Control slides were treated in an identical manner, except that an irrelevant IgG mouse monoclonal antibody was used in the first step or the primary antibody was omitted.

Cells were counted in five representative microscopic fields. We used an eye piece calibrated grid with 25× magnification. With this magnification and calibration, we counted the cells present in an area of 0.155 × 0.155 mm.

**Results**

### Immunohistochemistry

**Immunoreactivity**

The immunohistochemical staining of the conjunctival epithelium and stromal cells was assessed in all patients. The expression of CD68 was not observed in the conjunctival epithelium. CD68 reactivity was observed in the stromal cells, which included inflammatory cells such as eosinophils and neutrophils. The intensity of the staining varied among individual patients. In general, the staining was more intense in patients with active trachoma than in patients with VKC. Inactive trachoma showed a lower intensity of CD68 staining compared with active trachoma.

**Immunoreactivity for REGA-2D9**

The monoclonal antibody REGA-2D9 was raised against a gelatinase B specific REGA-2D9 monoclonal antibody (1:100). The mouse monoclonal antibody REGA-2D9 was raised against natural human neutrophil gelatinase B. This implies that the antigen preparation was devoid of gelatinase A. The REGA-2D9 is an IgG1 subtype with a dissociation constant devoid of gelatinase A. The REGA-2D9 is an IgG1 subtype with a dissociation constant.
activities were expressed as arbitrary laboratory scanning units. Densitometry was with a PDI densitometric scanner and the raw data were processed with the Pharmacia Biotech software programs LabScan Utility (Version 2.00) and Image Master ID (Version 2.0). The linear range was between 200 and 7000 scanning units. Quantitation of gelatinase activity by zymography proved to be a sensitive non-isotopic detection method with a sensitivity in the picogram range.\textsuperscript{16}

All data are presented as mean (SD). The Mann–Whitney U test was used to analyse the statistical significance of differences between mean values. The differences were considered significant if p value is <0.05.

**Results**

**IMMUNOLOCALISATION OF GELATINASE B**

There was no staining in the negative control slides. In normal conjunctiva, gelatinase B was detected only in polymorphonuclear cells located in the vascular lumens in three specimens (Fig 1(1)). All trachoma specimens showed immunoreactivity with the monoclonal antibody against macrophages CD68 (Fig 1(2)) and the monoclonal antibody against gelatinase B in the epithelial and stromal inflammatory infiltrate (Fig 1(3)). Gelatinase B was restricted to polymorphonuclear cells and monocytes/macrophages that showed cytoplasmic fine granular staining. The latter cells coexpressed the CD68 marker in serial sections confirming localisation of gelatinase B in monocytes/macrophages. Figure 2(4a) is stained for gelatinase B, and Figure 2(4b) for macrophages, and it is apparent that nearly all the macrophages of this specimen are positive for gelatinase B. In the stroma most of the positively stained polymorphonuclear cells were still located in the intravascular spaces. In the centre of the lymphoid follicles, the

![Figure 1](http://bjophoto.com/)

**Figure 1** (1) Immunohistochemical staining for gelatinase B of conjunctiva from a normal control subject showing no immunoreactivity (three step avidin-peroxidase conjugated biotin complex;×300). (2) Trachoma. Immunohistochemical staining for the macrophage marker CD68 showing immunoreactivity in the epithelial and stromal inflammatory infiltrate (arrows) (three step avidin-peroxidase conjugated biotin complex;×300). (3) Trachoma. Immunohistochemical staining for gelatinase B showing immunoreactivity in the epithelial (arrows) and stromal (arrowheads) inflammatory infiltrate (three step avidin-peroxidase conjugated biotin complex;×300).
monocytes/macrophages with gelatinase B immunoreactivity were larger and some were multinucleated giant cells (Fig 2(4)). The morphology of these cells varied from round to dendritic (Fig 2(5)).

In five VKC specimens gelatinase B immunoreactivity was localised in few polymorphonuclear cells and monocytes/macrophages present in the upper stromal inflammatory infiltrate. The majority of polymorphonuclear cells were located in the intravascular spaces. Monocytes/macrophages in VKC specimens were smaller than in trachoma specimens and showed weaker immunoreactivity (Fig 2(6)). The cell counts are presented in Tables 1 and 2. The numbers of gelatinase B positive monocyte/macrophage cells and polymorphonuclear cells in trachoma specimens were significantly higher than in VKC specimens.

**ZYMOGRAPHY**

Constitutive gelatinase A was detected in six of seven control specimens, and in six of seven trachoma specimens (Fig 3). The difference between the two groups was not statistically significant (control specimens, 210.5 (114.1) scanning units; trachoma specimens, 292.0 (187.0) scanning units, \( p = 0.2623 \)).

Inducible gelatinase B was detected in all conjunctival biopsy specimens (Fig 3). Gelatinase B values in trachoma specimens were significantly higher than the values found in control specimens (control specimens, 609.3 (395.9) scanning units; trachoma specimens, 883.6 (546.7) scanning units, \( p < 0.0001 \)).
Expression of gelatinase B in trachomatous conjunctivitis

To eliminate the variability inherent in the scanning procedure and possible differences in staining intensity between the zymograms, the ratio of gelatinase B to gelatinase A (B:A) was used to quantify the presence of gelatinase B. The mean B:A ratio in trachoma specimens was significantly higher than it was in control specimens (control specimens, 2.9 (1.9); trachoma specimens, 7.8 (5.2); p = 0.025).

Discussion

Chronic inflammatory conditions resulting in the restructuring of connective tissue are often characterised by a local accumulation of macrophages. These cells orchestrate connective tissue destruction either directly, by the secretion of elevated levels of matrix metalloproteinases, or indirectly, by the production of proinflammatory cytokines, including interleukin-1 (IL-1) and tumour necrosis factor-α (TNF-α). In turn, these cytokines have been shown to induce matrix metalloproteinase gene expression in the resident cell population. There is evidence that macrophages are activated during C trachomatis infection. In a previous immunohistochemical study we demonstrated the presence of increased numbers of macrophages in the conjunctival biopsy specimens from patients with active trachoma. These macrophages showed cytoplasmic expression of IL-1α, IL-1β, TNF-α, and platelet derived growth factor.

In the present study, gelatinase A levels did not vary among conjunctival biopsy specimens from control subjects, and those from subjects with active trachoma. Human gelatinase A occurs constitutively in body fluids (serum, synovial fluid, cerebrospinal fluid) and cell culture supernatants. It is suggested that gelatinase A appears to be involved in some basal extracellular matrix turnover events, whereas gelatinase B appears to be involved in more acutely regulated events. In normal human and mouse macrophages and human macrophage cell lines, gelatinase A is constitutively produced in small, but detectable, quantities and this production remains at the same level after macrophage stimulation. In contrast, unstimulated macrophages do not produce gelatinase B, but secrete large amounts of this enzyme after appropriate stimulation by, for example, cytokines and endotoxin. Zymography indicated an increasing activity of gelatinase B in trachoma specimens. Using immunohistochemistry, we have demonstrated that gelatinase B was specifically localised in macrophages and polymorphonuclear cells present in the inflammatory infiltrate in

Table 1 Number of gelatinase B positive cells in trachoma, vernal keratoconjunctivitis (VKC), and control specimens (cells counted in an area of 0.155 x 0.155 mm)

<table>
<thead>
<tr>
<th>Patient No</th>
<th>Monocyte/macrophage cells</th>
<th>Polymorphonuclear cells</th>
</tr>
</thead>
<tbody>
<tr>
<td>Trachoma</td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>19</td>
<td>5</td>
</tr>
<tr>
<td>2</td>
<td>26</td>
<td>4</td>
</tr>
<tr>
<td>3</td>
<td>34</td>
<td>2</td>
</tr>
<tr>
<td>4</td>
<td>43</td>
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<td>6</td>
<td>64</td>
<td>1</td>
</tr>
<tr>
<td>VKC</td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>11</td>
<td>2</td>
</tr>
<tr>
<td>2</td>
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<tr>
<td>4</td>
<td></td>
<td>1</td>
</tr>
<tr>
<td>5</td>
<td></td>
<td>0</td>
</tr>
<tr>
<td>6</td>
<td></td>
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</tr>
<tr>
<td>Control</td>
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</tr>
<tr>
<td>1</td>
<td>10</td>
<td>0</td>
</tr>
<tr>
<td>2</td>
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<td>8</td>
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<tr>
<td>6</td>
<td></td>
<td>2</td>
</tr>
</tbody>
</table>

1739.6 (1078.3) scanning units; p = 0.0127.

The zymography results are presented in Table 3.

To eliminate the variability inherent in the scanning procedure and possible differences in staining intensity between the zymograms, the ratio of gelatinase B to gelatinase A (B:A) was used to quantify the presence of gelatinase B. The mean B:A ratio in trachoma specimens was significantly higher than it was in control

Table 2 Number of gelatinase B positive cells in trachoma and vernal keratoconjunctivitis (VKC) specimens (mean (SD))

<table>
<thead>
<tr>
<th>Cell type</th>
<th>Trachoma (n=6)</th>
<th>VKC (n=6)</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Monocyte/macrophage cells</td>
<td>52.3 (21.9)</td>
<td>8.2 (6.4)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Polymorphonuclear cells</td>
<td>23.2 (14.2)</td>
<td>6.3 (5.4)</td>
<td>0.013</td>
</tr>
</tbody>
</table>

Table 3 Gelatinases A and B levels in trachoma and control specimens (scanning units)

<table>
<thead>
<tr>
<th>Patient No</th>
<th>Gelatinase A</th>
<th>Gelatinase B</th>
</tr>
</thead>
<tbody>
<tr>
<td>Trachoma</td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>205</td>
<td>1097</td>
</tr>
<tr>
<td>2</td>
<td>217</td>
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<td>3</td>
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<td>2499</td>
</tr>
<tr>
<td>4</td>
<td>256</td>
<td>1648</td>
</tr>
<tr>
<td>5</td>
<td>ND</td>
<td>572</td>
</tr>
<tr>
<td>6</td>
<td>661</td>
<td>3778</td>
</tr>
<tr>
<td>7</td>
<td>276</td>
<td>1121</td>
</tr>
<tr>
<td>Control</td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>ND</td>
<td>629</td>
</tr>
<tr>
<td>2</td>
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<td>848</td>
</tr>
<tr>
<td>3</td>
<td>179</td>
<td>1110</td>
</tr>
<tr>
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<td>238</td>
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<tr>
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<td>371</td>
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<tr>
<td>6</td>
<td>171</td>
<td>166</td>
</tr>
<tr>
<td>7</td>
<td>64</td>
<td>137</td>
</tr>
</tbody>
</table>

ND = not detected.

specimens (control specimens, 2.9 (1.9); trachoma specimens, 7.8 (5.2); p = 0.025).

Figure 3 Gelatin zymography of conjunctival biopsy specimens from normal subjects (A), and from patients with active trachoma (B). The zymographies of these samples show the presence of both gelatinase A (72 kDa) and gelatinase B (92 kDa).
trachoma specimens. Macrophages, monocytes, and neutrophils cells are the main producers of gelatinase B.22 Most of the stromal polymorphonuclear cells were in the intravascular spaces indicating that macrophages were primarily responsible for the production of gelatinase B. Some macrophages were organised into giant cells that stained positively with the anti-gelatinase B monoclonal antibody. Macrophages synthesise and secrete matrix metalloproteinases which are capable of degrading all macromolecular constituents of the extracellular matrix.3 These enzymes appear to function as a proteolytic cascade of which gelatinase B is a downstream element and whose activation is initiated after that of stromelysin.4 The intracellular localisation of gelatinase B in macrophages can be interpreted as evidence of active synthesis of this enzyme because macrophages do not store this metalloproteinase.5 In general, the expression of gelatinase B by macrophages in trachoma agrees with immunolocalisation studies in other chronic inflammatory diseases such as interstitial lung diseases,1 emphysema,1 pulmonary fibrosis,6 aneurysmal and occlusive aortic diseases,7 rheumatoid arthritis,8 coronary atherosclerotic lesions,9 temporal arteritis10 and chronic synovitis.11 Chlamydiae are obligate intracellular Gram negative bacteria which cause persistent intracytoplasmic infection of eukaryotic cells. The mechanism by which C trachomatis induces gelatinase B expression by macrophages has not been determined. The effects of C trachomatis on the synthesis and release of this enzyme may be related to the production of cytokines. C trachomatis is an effective inducer of IL-1 and TNF-α production by monocytes.34 Whole spleen cells produced TNF-α in vitro in response to murine C trachomatis in a mouse model of pneumonia caused by murine C trachomatis.37 IL-1α and IL-1β mRNA and bioactivity are induced in murine lungs in response to chlamydial infection.38 In agreement with these studies we have demonstrated upregulated production of IL-1α, IL-1β, and TNF-α in the conjunctiva from patients with active trachoma.10 TNF-α, IL-1β, and TNF-α selectively upregulate gelatinase B by macrophages.16 As in all Gram negative bacteria, the outermost monolayer of C trachomatis is composed of lipopolysaccharide (LPS), a complex glycolipid essential for bacterial survival.9 LPS is a potent inducer of the biosynthesis and secretion of gelatinase B in macrophages.16 17 40–42 Recently, Ingalls and associates42 have shown that the LPS from Neisseria gonorrhoeae or from Salmonella minnesota. According to these investigators, this relatively weak induction of an acute immune response may partially explain the asymptomatic nature of chlamydial infection.

Gelatinase B degrades denatured collagens (gelatin), collagen types IV, V, VII, and X, elastin and fibronectin.42 43 Therefore, expression of gelatinase B in trachoma may contribute to progressive breakdown of conjunctiva by degrading minor constituents of the extracellular matrix. The degradation of fibrillar collagen types I, II, and III is initiated by collagenases and completed by gelatinase B. The degradation of collagen types IV and V associated with basement membranes permits the extravasation of macrophages into conjunctival tissue. It has been demonstrated that in vitro migration of T lymphocytes across a basement membrane equivalent is mediated by gelatinase B.39 Macrophage gelatinase B might enhance fibroblast migration into conjunctival epithelium by creating breaks in epithelial basement membrane. Since collagen fragments may be chemotactic for monocytes, degradation of collagen by macrophages could further promote monocyte recruitment.10 In addition, chronic release of specific collagen fragments in the conjunctiva by gelatinase B might contribute to selection of autoreactive T cells and generation of autoimmunity.12

In conclusion, the findings reported in this study provide evidence that active synthesis of gelatinase B by macrophages might be involved in matrix degradation and promotion of conjunctival scarring in trachoma. These findings are potentially important from a fundamental standpoint because they suggest a pathogenic role for gelatinase B in the development of conjunctival scarring. From a practical standpoint, these findings raise the possibility that inhibitors of metalloproteinase activity, and specifically of gelatinase B, might provide a novel form of therapy for prevention of conjunctival scarring in trachoma.

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