Topically administered timolol and dorzolamide reduce intraocular pressure and protect retinal ganglion cells in a rat experimental glaucoma model

M Seki, T Tanaka, H Matsuda, T Togano, K Hashimoto, J Ueda, T Fukuchi, H Abe

Aims: This study sought to elucidate the effects of timolol and dorzolamide on intraocular pressure (IOP) and retinal ganglion cell (RGC) death in an experimental model of glaucoma in rat.

Methods: Mild elevation of IOP was induced in rats by intracameral injection of India ink and subsequent laser trabecular photocoagulation. IOP was measured before the surgical procedures and weekly thereafter. Timolol (0.5%), timololXE (0.5%), dorzolamide (1%), and artificial tears (vehicle) were topically applied daily. Retinal sections were prepared for histology to determine RGC number.

Results: Timolol, timololXE, and dorzolamide induced a significant reduction in IOP (p < 0.05) and counteracted the reduction in RGC number that occurred in vehicle treated glaucomatous eyes (p < 0.05). The coefficient of correlation between RGC number and IOP was significant in the dorzolamide treated group (r = −0.908, p < 0.005), but not in other groups (p > 0.05).

Conclusions: Both timolol formulation and dorzolamide reduced IOP and protected RGCs in a rat model of experimental glaucoma. It cannot be ruled out that timolol might protect RGCs by additional mechanisms other than simply lowering of IOP.

G laucoma is a disorder featuring irreversible retinal ganglion cell (RGC) death which can be correlated with elevated intraocular pressure (IOP); however, the mechanism of RGC death in glaucoma is not fully understood.1 At present, the only well established treatment of glaucoma involves lowering the IOP.2 Nevertheless, randomised controlled clinical trials have demonstrated that even if medical and surgical treatments successfully lower the IOP, visual disturbances continue to progress in some patients.3,4 Thus, new methods of treating glaucoma by directly preventing RGC death are being investigated. Such neuroprotective strategies to protect RGCs in vitro or animal models of neural damage induced by optic nerve transection, intraocular injection of glutamate, and transient ischaemic injury of the retina. In order to assess effectively the therapeutic value of such agents in glaucoma, however, they must be tested in animal models of glaucoma and then in randomised controlled clinical trials involving patients with glaucomatous optic neuropathy.5 Therefore, we investigated effects of timolol, a β adrenoceptor antagonist, and dorzolamide, a carbonic anhydrase inhibitor, upon IOP and RGC number using an experimental glaucoma model to assess their neuroprotective effects.

MATERIALS AND METHODS
Animal model of glaucoma
All experimental procedures involving animals were performed in accordance with both the ARVO statement for the use of animals in ophthalmic and vision research and our institutional guidelines for care and use of laboratory animals. Adult male Wistar rats (Japan SLC, Hamamatsu, Japan), 9 weeks of age (250–300 g), were housed in standard lighting conditions (12 hour light and 12 hour dark cycle). Chronic elevation of IOP was induced in one eye of each animal by intracameral injection of India ink and subsequent laser trabecular photocoagulation 1 week after the injection.7 In each animal, the opposite eye was left unoperated. IOP was measured at the end of the light phase (17:00–18:00) with a Tono-Pen XL tonometer (BioRad, Santa Ana, CA, USA) with topical anaesthesia with a drop of 2% lidocaine applied to the eyes according to a previously described method8 once before the surgical induction of glaucoma and weekly thereafter. Rats with an IOP greater than 22 mm Hg were classified as glaucomatous. The experimental schedule is summarised in figure 1A.

Drug treatment
Topical administration of antiglaucoma drugs was started 1 week after the laser photocoagulation and lasted for 3 weeks. Medications used in this study were timolol maleate (0.5%, Timolol; Merck, Whitehouse Station, NJ, USA, n = 6), timolol maleate gel forming solution (0.5%, TimololXE; Merck, n = 7), and dorzolamide hydrochloride (1%, Trusopt; Merck, n = 7). Ten μl of eye drop was applied twice, once, and three times daily, respectively for each drug, following clinical protocols in Japan. Artificial tears (Mytear; Senju Pharmaceutical, Osaka, Japan, n = 8) were used as vehicle (three times daily), and unoperated eyes from the vehicle treated animals served as controls (n = 7).

Histopathology
Animals were deeply anaesthetised by chloral hydrate and sacrificed by decapitation. Posterior eye cups were fixed with 4% paraformaldehyde, dehydrated, and embedded in parafﬁn. Radial sections (4 μm) were stained with Mayer’s haematoxylin solution (Wako, Osaka, Japan) and Eosin Alchohol Solution (Wako). Specimens were observed at a distance of 1.5 mm from the centre of the optic nerve head under an AxioPhoto2 microscope (Carl Zeiss, Jena, Germany), and photomicrographs were captured using a digital camera (AxioCam; Carl Zeiss). Cells in the ganglion

Abbreviations: BDNF, brain derived neurotrophic factor; IOP, intraocular pressure; RGC, retinal ganglion cell
Neuroprotection by timolol and dorzolamide

The effect of topically applied antiglaucoma drugs on IOP in the rat glaucoma model

Time course of mean IOP level in each group is shown in figure 1B. Treatment with timolol, timolol XE, and dorzolamide all resulted in a decrease in IOP (p < 0.01, paired t test). However, IOP was also reduced after 3 weeks even in the vehicle treated group (p < 0.05).

Before drug treatment, the IOP of glaucomatous eyes was not significantly different between experimental groups (p = 0.92, one way ANOVA) (fig 1C). Both timolol and timolol XE significantly lowered IOP (p < 0.001 in comparison with the vehicle treated group, one way ANOVA followed by Fisher’s PLSD multiple comparison test). Dorzolamide also reduced IOP after 1 and 2 weeks of treatment (p < 0.05). The effect on IOP level of timolol and timolol XE was significantly greater than that of dorzolamide, indicating that they are more effective in lowering IOP in this animal model (p < 0.05 in comparison with the dorzolamide treated group, one way ANOVA followed by Fisher’s PLSD multiple comparison test).

The therapeutic effect of antiglaucoma drugs on RGCs in the rat glaucoma model

In order to analyse the therapeutic efficacy of antiglaucoma drugs on RGC degeneration in this animal model, radial sections of the retina were prepared (fig 2), and RGC number was counted (table 1). In glaucomatous eyes, RGC number was markedly decreased (control v vehicle, p < 0.005, one way ANOVA followed by Fisher’s PLSD multiple comparison test). Treatment with antiglaucoma drugs efficiently prevented to varying extents this decrease in RGC number (p < 0.05 in comparison with the vehicle treated eyes). Although timolol and dorzolamide treated groups did show reduced RGC number when compared to control eyes (p < 0.05), no statistically significant difference was observed between control and timolol XE treated eyes (p = 0.09).

Correlation between IOP and RGC number in the rat glaucoma model

A weak negative correlation was present between IOP and RGC count when Pearson’s correlation coefficient analysis was made with all glaucomatous eyes (r = -0.377, p < 0.05) (fig 3). Although a highly significant negative relation was
found between IOP and RGC number in the dorzolamide treated group \( (r = -0.908, p < 0.005) \), no such statistically significant correlation was found in either of the other two treatment groups \( (p > 0.05) \).

**DISCUSSION**

In the present study, we assessed the effects of timolol and dorzolamide upon IOP and RGCs in a rat model of experimental glaucoma,\(^7\) whose reproducibility was demonstrated by others.\(^8\) Our present study showed that both timolol and dorzolamide reduced IOP (fig 1) and protected RGCs (fig 2 and table 1). Further correlation analysis revealed that IOP was correlated with RGC damage in dorzolamide treated, but not in timolol and timolol XE treated eyes. Thus, dorzolamide might protect RGCs from degeneration mainly in an IOP dependent manner under the conditions used in this study. In contrast with our study, however, dorzolamide was not effective in preventing RGC death in DBA/2J mice that had spontaneously developed elevated IOP.\(^9\) Although lower IOP preserves visual field of patients with primary open angle glaucoma,\(^12\) some studies failed to demonstrate correlation between IOP and visual field sensitivity in timolol treated glaucomatous individuals.\(^13\) \(^14\) Similarly, we did not see any sign of such correlation in the animal model of glaucoma in the present study, leaving open the mechanism by which timolol protected RGCs. There have been some previous reports of the potential neuroprotective effects of \( \beta \) adrenergic antagonists as assessed both in vitro\(^9\) and in an in vivo animal model of retinal insult induced by transient ischaemia.\(^15\) Thus, it cannot be ruled out that timolol might protect RGCs by additional mechanisms other than lowering IOP in the present study.

One possibility for how topically applied timolol rescues RGCs involves brain derived neurotrophic factor (BDNF), which is a strong neuroprotectant of RGCs.\(^16\) \(^17\) Topically applied betaxolol, a \( \beta \) adrenergic antagonist, was shown to stimulate BDNF mRNA expression in the retina.\(^18\) Therefore, upregulation of BDNF expression upon topical administration of timolol might protect RGCs. BDNF is one of the most attractive candidate neuroprotective drugs for glaucoma; its therapeutic potential has been demonstrated in a rat glaucoma model.\(^19\) In that study, BDNF was injected directly into the eyeball, but frequent injections into patients' vitreous space are not practical. We previously demonstrated that BDNF is expressed endogenously in the rat retina.\(^20\) The discovery raises a more practical therapeutic approach: induction of endogenous BDNF by external administration of agents with a smaller molecular weight, such as \( \beta \) adrenergic antagonists, may be useful to treat glaucoma.

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**Table 1** RGC count in radial section (cells/mm)

<table>
<thead>
<tr>
<th></th>
<th>RGC count (cells/mm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>36.1 (3.1)†</td>
</tr>
<tr>
<td>Vehicle</td>
<td>26.4 (3.0)†</td>
</tr>
<tr>
<td>Timolol</td>
<td>30.2 (3.5)††</td>
</tr>
<tr>
<td>Timolol XE</td>
<td>33.0 (4.1)††</td>
</tr>
<tr>
<td>Dorzolamide</td>
<td>31.6 (2.8)††</td>
</tr>
</tbody>
</table>

Results are expressed as means (SD) for control \( (n=7) \) and vehicle \( (n=8) \), timolol \( (n=6) \), timolol XE \( (n=7) \), and dorzolamide treated \( (n=7) \) eyes. Statistical comparisons were made by one way ANOVA followed by Fisher's PLSD multiple comparison test: \( p < 0.005 \) v control, \( p < 0.05 \) v control, \( p < 0.005 \) v vehicle, \( p < 0.05 \) v vehicle.

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**Figure 2** To determine RGC number in retinas of experimental animals, cells in the ganglion cell layer were counted in radial sections. GCL, ganglion cell layer; IPL, inner plexiform layer; INL, inner nuclear layer; OPL, outer plexiform layer; ONL, outer nuclear layer. Scale bar = 50 \( \mu \text{m} \).

**Figure 3** Pearson's correlation coefficient analysis was used to investigate the correlation between IOP and RGC number. IOP and RGC number were weakly negatively correlated for all glaucomatous eyes \( (r = -0.377, p < 0.05) \).
REFERENCES
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