Association of varicella zoster virus load in the aqueous humor with clinical manifestations of anterior uveitis in herpes zoster ophthalmicus and zoster sine herpete

S Kido,1 S Sugita,1 S Horie,1 M Miyanaga,2 K Miyata,2 N Shimizu,3 T Morio,4 M Mochizuki1

ABSTRACT

Aim: To investigate whether clinical manifestations of anterior uveitis are associated with the viral load of varicella zoster virus (VZV) in the aqueous humor in patients with herpes zoster ophthalmicus (HZO) and zoster sine herpete (ZSH).

Methods: After informed consent was given, a aliquot of aqueous humor was collected from patients with VZV anterior uveitis (n = 8). Genomic DNA of the human herpes viruses was measured in the aqueous humor by two PCR assays: a qualitative multiplex PCR and a quantitative real-time PCR.

Results: All patients had unilateral acute anterior uveitis with high intraocular pressure, mutton fat keratic precipitates with some pigmentation, and trabecular meshwork pigmentation. Multiplex PCR demonstrated VZV genomic DNA in all of the samples, but not in other human herpes virus samples (human simplex virus types 1 and 2, Epstein–Barr virus, cytomegalovirus and human herpes virus types 6, 7 and 8). Real-time PCR revealed a high copy number of VZV DNA in the aqueous humor. After the initial onset of anterior uveitis, iris atrophy and distorted pupil with paralytic mydriasis developed. The intensity of iris atrophy and pupil distortion, but not ocular hypertension, correlated with the viral load of VZV in the aqueous humor.

Conclusion: VZV viral load in the aqueous humor correlated significantly with damage to the iris (iris atrophy and pupil distortion) in patients with HZO and ZSH.

Materials and Methods

Subjects

The subjects were eight patients (three men and five women; age range 43–71 years (mean 61)) with diagnosed VZV anterior uveitis at Tokyo Medical and Dental University Hospital and Miyata Hospital between December 1999 and September 2007. The clinical diagnosis of VZV anterior uveitis was based on observation of anterior uveitis associated with either HZO or ZSH. After informed consent had been obtained, an aliquot of aqueous humor (0.1–0.2 ml) was obtained from each patient. The research followed the tenets of the Declaration of Helsinki, and the study was approved by the institutional ethics committees of Tokyo Medical and Dental University.

PCR assay

The aqueous humor samples were centrifuged at 3000 rpm for 5 min and used for the following PCR assays: multiplex PCR and real-time PCR.

Multiplex PCR was designed to qualitatively measure the genomic DNA of eight human herpes viruses: herpes simplex virus type 1 (HSV-1) and type 2 (HSV-2), VZV, Epstein–Barr virus (EBV), cytomegalovirus (CMV), human herpes virus type 6 (HHV-6), type 7 (HHV-7) and type 8 (HHV-8).

DNA was extracted from the aqueous humor samples using a DNA minikit (Qiagen, Valencia, CA, USA). Multiplex PCR was performed using LightCycler (Roche, Basle, Switzerland). The primer sequences and PCR conditions for VZV were as previously described.

Real-time PCR was performed only for HHV, the genomic DNA of which was detected by multiplex PCR. It was performed by using Amplitaq Gold and a Real-Time PCR 7500 system (ABI, Forster City, CA, USA). The primer sequences for VZV (ORF29) used in real-time PCR were designed to use Primer Express (ABI) forward, AACTTTTACATCCAGCCTCGCG; reverse, GAAAACCCAAAAGGTTCTCGAG. The probe was FAM-TGCTTTTCACGAGGGAACG-TAMRA. The following PCR conditions were used: denaturation at 95°C for 10 min, 95°C for 15 s, and 60°C for 60 s for 40 cycles.
Statistical analysis
Statistical analysis was performed using the Mann–Whitney U test. Statistical significance was set at $p < 0.05$.

RESULTS
Clinical manifestations
In four of the patients, there was an episode of skin eruption with neuralgia in the area of the first branch of the trigeminal nerve; this was clinically diagnosed as HZO (table 1). No skin eruptions were observed in the other patients, although they complained of pain near the first branch of the trigeminal nerve, leading to the diagnosis of ZSH. At various intervals after the onset of HZO or ZSH, the patients developed unilateral anterior uveitis. All patients had cells and flare in the anterior chamber, with increased flare values measured by a laser flare meter (Cowa, Tokyo, Japan). Four patients exhibited mutton-fat KPs (table 1), which had brownish pigmentation and were small or medium in size. Three patients with HZO already exhibited iris atrophy and pupil distortion on referral. High intraocular pressure (IOP) was recorded in all patients except in case 3 (table 1). A gonioscopic examination revealed wide, open angles in all patients, and higher pigmentation in the affected eye than the other eye in six of the eight patients. Ophthalmoscopic examinations revealed no significant inflammatory lesions in the retina and choroid. These clinical results led us to a diagnosis of anterior uveitis associated with HZO or ZSH, with paracentesis subsequently performed in order to carry out the PCR analysis.

After confirmation of the presence of VZV in the aqueous humor by multiplex PCR, systemic anti-VZV agents (aciclovir or valaciclovir) and aciclovir ointment were administered for at least 4 weeks together with a topical corticosteroid (eg, betamethasone) and anti-glaucoma agents (eg, timolol and latanoprost). Iris atrophy and pupil distortion developed during the clinical course (fig 1, table 2). The anterior uveitis and high IOP responded well to treatment.

PCR analysis of the aqueous humor
Qualitative PCR (multiplex PCR) detected genomic DNA of VZV but not of other human herpes viruses (HSV-1, HSV-2, EBV, CMV, HHV-6, HHV-7 and HHV-8) in the aqueous humor of all eight patients (table 1). In peripheral blood samples, however, no genomic DNA of any of the eight human herpes viruses, including VZV, was detected in any of the patients.

Quantitative PCR (real-time PCR) detected significant viral loads of VZV DNA in the aqueous humor of the eight patients ranging from $3.8 \times 10^2$ to $1.2 \times 10^7$ copies/ml (table 2). It is of note that the VZV viral load in the aqueous humor correlated with the intensity of iris atrophy and pupil distortion (table 2, fig 1). Patient 1 had the highest VZV viral load ($1.2 \times 10^7$ copies/ml) in the aqueous humor and also had the most severe iris atrophy, with multiple wide areas of segmental iris atrophy and a widely dilated pupil (fig 1A, table 2). Patients 2–5 had the second

![Table 1 Clinical findings at initial presentation in patients with varicella zoster virus (VZV) anterior uveitis](image)

### Table 1 Clinical findings at initial presentation in patients with varicella zoster virus (VZV) anterior uveitis

<table>
<thead>
<tr>
<th>Case</th>
<th>Age (years)</th>
<th>Sex</th>
<th>Eye</th>
<th>VA</th>
<th>IOP (mm Hg)</th>
<th>Mutton-fat KPs</th>
<th>Cells in AC</th>
<th>Flare in AC</th>
<th>Eruption</th>
<th>Time from onset to treatment</th>
<th>Herpes virus DNA</th>
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<tbody>
<tr>
<td>1</td>
<td>67</td>
<td>F</td>
<td>Left</td>
<td>0.3</td>
<td>38</td>
<td>+</td>
<td>3+</td>
<td>106</td>
<td>Partial</td>
<td>–</td>
<td>2 months</td>
</tr>
<tr>
<td>2</td>
<td>70</td>
<td>F</td>
<td>Left</td>
<td>0.3</td>
<td>22</td>
<td>+</td>
<td>3+</td>
<td>106</td>
<td>Partial</td>
<td>–</td>
<td>2 weeks</td>
</tr>
<tr>
<td>3</td>
<td>68</td>
<td>F</td>
<td>Right</td>
<td>0.8</td>
<td>10</td>
<td>+</td>
<td>2+</td>
<td>34</td>
<td>None</td>
<td>–</td>
<td>2 months</td>
</tr>
<tr>
<td>4</td>
<td>56</td>
<td>M</td>
<td>Left</td>
<td>1.2</td>
<td>46</td>
<td>–</td>
<td>3+</td>
<td>241</td>
<td>Partial</td>
<td>–</td>
<td>2 months</td>
</tr>
<tr>
<td>5</td>
<td>70</td>
<td>F</td>
<td>Left</td>
<td>1.2</td>
<td>25</td>
<td>–</td>
<td>1+</td>
<td>13</td>
<td>Partial</td>
<td>–</td>
<td>2 months</td>
</tr>
<tr>
<td>6</td>
<td>44</td>
<td>M</td>
<td>Right</td>
<td>0.6</td>
<td>46</td>
<td>–</td>
<td>3+</td>
<td>140</td>
<td>Partial</td>
<td>1 week</td>
<td>–</td>
</tr>
<tr>
<td>7</td>
<td>43</td>
<td>F</td>
<td>Left</td>
<td>1.2</td>
<td>28</td>
<td>–</td>
<td>2+</td>
<td>15</td>
<td>Partial</td>
<td>1 month</td>
<td>–</td>
</tr>
<tr>
<td>8</td>
<td>71</td>
<td>M</td>
<td>Right</td>
<td>0.3</td>
<td>28</td>
<td>–</td>
<td>2+</td>
<td>13</td>
<td>None</td>
<td>None</td>
<td>–</td>
</tr>
</tbody>
</table>

Aqueous humor samples from eight cases were analysed for human herpes virus DNA by multiplex PCR.

*Herpes viruses excluding VZV, ie, herpes simplex virus type 1 and type 2, Epstein–Barr virus, cytomegalovirus, human herpes virus types 6, 7 and 8.

VA, visual acuity; IOP, intraocular pressure; KPs, keratic precipitates; AC, anterior chamber.

Figure 1 Iris photographs for patients with varicella zoster virus (VZV) anterior uveitis. Slit-lamp photographs are available for seven cases as shown. The numbers indicate the copies of the VZV genomic DNA for each of the aqueous humor samples (copies/ml). Arrows point to areas of iris atrophy. Consent has been obtained for publication of this figure.
highest viral loads in the aqueous humor (~1×10^6 copies/ml), and these patients developed multiple segmental or circular iris atrophy with moderate pupil distortion, although these signs were not as marked as in patient 1 (fig 1B–E, table 2). Patients 6 and 7 had the third highest viral load (10^6–10^7 copies/ml) and exhibited circular iris atrophy with minimum pupil distortion (fig 1F,G). Patient 8 had the lowest viral load. This patient exhibited circumferential iris atrophy with minimal pupil distortion.

**DISCUSSION**

Human herpesviruses are known to be involved in many ocular pathological conditions, such as keratitis, anterior uveitis and necrotising retinitis. Early treatment with anti-viral agents, based on a rapid and accurate diagnosis of the viral infection in ocular tissues, is clinically important in order to avoid the irreversible tissue damage and visual impairment caused by viral infection. Recent advances in PCR methodology have made it possible to screen for viral infection and further quantify the intensity of the viral infection. Recent advances in PCR methodology have made it possible to screen for viral infection and further quantify the intensity of the viral infection in ocular inflammatory diseases.1–3

We next examined whether higher viral load in the aqueous humor was significantly associated with the intensity of the iris atrophy and pupil distortion. For this analysis, we separated the patients into two groups: higher viral load (patients 1–5) and lower viral load (patients 6–8). The extent of iris atrophy was significantly (p<0.05) larger in the group with higher viral load. In addition, the maximum pupil diameter in the pupil distortion was significantly (p<0.05) larger in the group with higher viral load. These results suggest that higher viral load in the aqueous humor is closely associated with the intensity of the iris atrophy and pupil distortion.

### Table 2 Virolological analysis and ocular findings after treatment of patients with varicella zoster virus (VZV) anterior uveitis

<table>
<thead>
<tr>
<th>Case</th>
<th>VZV DNA (copies/ml)</th>
<th>VA</th>
<th>IOP (mm Hg)</th>
<th>Cells in AC</th>
<th>Flare in AC</th>
<th>Iris atrophy (%)</th>
<th>Pupil</th>
<th>Maximum pupil diameter (mm)</th>
<th>Systemic treatment</th>
<th>Follow-up (months)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>1.2×10^6</td>
<td>mm</td>
<td>10</td>
<td>–</td>
<td>13</td>
<td>Wide</td>
<td>40</td>
<td>Distorted, mydriasis</td>
<td>8.1</td>
<td>ACV, ACZ</td>
</tr>
<tr>
<td>2</td>
<td>1.2×10^6</td>
<td>0.9</td>
<td>14</td>
<td>–</td>
<td>8</td>
<td>Segmental</td>
<td>4</td>
<td>Distorted, mild mydriasis</td>
<td>3.4</td>
<td>VCV, ACZ</td>
</tr>
<tr>
<td>3</td>
<td>1.0×10^6</td>
<td>0.8</td>
<td>7</td>
<td>–</td>
<td>5</td>
<td>Segmental</td>
<td>15</td>
<td>Distorted, mydriasis</td>
<td>5.0</td>
<td>VCV</td>
</tr>
<tr>
<td>4</td>
<td>9.8×10^6</td>
<td>0.9</td>
<td>10</td>
<td>–</td>
<td>73</td>
<td>Circular</td>
<td>20</td>
<td>Distorted, mydriasis</td>
<td>7.9</td>
<td>VCV</td>
</tr>
<tr>
<td>5</td>
<td>9.8×10^6</td>
<td>0.8</td>
<td>20</td>
<td>–</td>
<td>22</td>
<td>Segmental</td>
<td>29</td>
<td>Distorted, mydriasis</td>
<td>7.0</td>
<td>VCV</td>
</tr>
<tr>
<td>6</td>
<td>1.3×10^6</td>
<td>1.5</td>
<td>13</td>
<td>–</td>
<td>6</td>
<td>Circular</td>
<td>1</td>
<td>Normal</td>
<td>2.6</td>
<td>ACV</td>
</tr>
<tr>
<td>7</td>
<td>2.2×10^10</td>
<td>2</td>
<td>16</td>
<td>–</td>
<td>3</td>
<td>Circular</td>
<td>1</td>
<td>Distorted, mild mydriasis</td>
<td>4.3</td>
<td>VCV</td>
</tr>
<tr>
<td>8</td>
<td>3.8×10^6</td>
<td>1.2</td>
<td>12</td>
<td>–</td>
<td>9</td>
<td>None</td>
<td>0</td>
<td>None</td>
<td>2.6</td>
<td>ACV, ACZ</td>
</tr>
</tbody>
</table>

The copy number of the VZV genome in aqueous humor was evaluated with real-time PCR. The extent of iris atrophy was calculated using Photoshop Elements V2.0.

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COMPETING INTERESTS: None declared.

ETHICS APPROVAL: Ethics approval was obtained.

PATIENT CONSENT: Consent has been obtained for publication of fig 1.

REFERENCES


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