

## **Mechanical stress potentiates the differentiation of periodontal ligament stem cells into keratocytes**

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## **Supplementary materials and methods**

### ***Isolation and characterization of PDLSCs***

Human PDLSCs were isolated from surgically removed impacted third molars. The periodontal ligaments were cut into pieces (3-4 mm<sup>3</sup>), and enzymatically digested in 3 mg/ml collagenase type I (Worthington Biochemicals Corp., Freehold, NJ) and 4 mg/ml dispase II (Roche Diagnostics, Indianapolis, IN) for 1 hour at 37°C. Single cell suspensions were obtained by passing cells through a 70 µm strainer (BD Falcon™ Labware, Franklin Lakes, NJ). The cells were seeded on plates and cultured in growing medium, Minimum Essential Medium-α (Life technologies, Grand Island, NY) with 10% FBS (Life technologies) and 1% penicillin-streptomycin (Life technologies). The PDLSCs used in the present project have recently been characterized in another study of ours (Chen et al.; *Stem Cell Res Ther* 2017). They were found to have clonogenic property, multi-lineage differentiation potential and positively expressed MSCs surface markers (CD73, CD90 and CD105), and negative expression for CD11b, CD19, CD34, CD45 and HLA-DR.

### ***Western blot analysis***

Protein was extracted from cell samples by RIPA lysis and centrifugation. The concentration was quantified using Bio-Rad Protein Assay (Bio-Rad, Hercules, CA, USA). Loaded protein samples were ran for 1 hour at 150 V in pre-cast polyacrylamide gel (Bio-Rad), and subsequently transferred to a PVDF Blotting Membrane (GE Healthcare, Little Chalfont, Buckinghamshire, UK) for 1 hour at 100 V. After blocking in 5 % bovine serum albumin (BSA) in TBST for 1 hour, the blots were incubated with primary antibodies (Supplementary Table 3) overnight at 4 °C. After that, the blots were washed with TBST for three times, and then incubated with a HRP-linked secondary antibody (Supplementary Table 3) for 1 hour. After wash in TBST for another three times, the blots were incubated with ECL (GE Healthcare) for 5 minutes. Odyssey® Fc Dual-Mode Imaging System (LI-COR, Lincoln, NE, USA) was used to visualize the bands. And ImageJ analysis software (NIH) was used to evaluate the densitometry.

**Supplementary Table 1.** Probes used for qPCR.

Gene name	Gene symbol	Assay ID
Aldehyde dehydrogenase 3A1	<i>ALDH3A1</i>	Hs00964880_m1
CD34	<i>CD34</i>	Hs00990732_m1
Lumican	<i>LUM</i>	Hs00929860_m1
Collagen type I	<i>COL1</i>	Hs00164004_m1
Collagen type V	<i>COL5</i>	Hs00609133_m1
Alpha 2 smooth muscle actin	<i>ACTA2</i>	Hs00909449_m1
$\beta$ -actin	<i>ACTB</i>	4352667

**Supplementary Table 2.** Primers used for qPCR.

Genes	5'-3'	Primers
Glyceraldehyde 3-phosphate dehydrogenase ( <i>GAPDH</i> )	Forward	TGACGCTGGGGCTGGCATTG
	Reverse	GGCTGGTGGTCCAGGGGTCT
Scleraxis ( <i>SCX</i> )	Forward	CGAGAACACCCAGCCCAAAC
	Reverse	CTCCGAATCGCAGTCTTTCTGTC
Tenomodulin ( <i>TNMD</i> )	Forward	TGGGTGGTCCCTCAAGTGAAAGT
	Reverse	CTCGACGGCAGTAAATACAACAATA
Integrin alpha 1 ( <i>ITGA1</i> )	Forward	AAGGGGAGAACTTCGGAGTGA
	Reverse	AAATGAGCAGCATTAAACAGCAAC
Integrin alpha 2 ( <i>ITGA2</i> )	Forward	GGGAATCAGTATTACACAACGGG
	Reverse	CCACAACATCTATGAGGGAAGGG
Integrin beta 1 ( <i>ITGB1</i> )	Forward	TTCAGTGAATGGGAACAACGA
	Reverse	ATGCAAGGCCAATAAGAACAA
Non-muscle myosin II B ( <i>MYH10</i> )	Forward	GCCGCCAACAAATTAGTCCGT
	Reverse	GCGTTTAAGCTGCTTCATCCGA

**Supplementary Table 3.** Antibodies used for immunofluorescence staining and western blot.

Antibody	Company	Code
ALDH3A1	Abcam	Ab76976
CD34	Santa Cruz	sc-9095
Lumican	Santa Cruz	sc-166871
Lumican	Abcam	ab168348
Keratocan	Santa Cruz	sc-66941
Collagen type I	Abcam	ab34710
Collagen type V	Santa Cruz	sc-166155
Alpha smooth muscle actin	Abcam	ab5694
$\beta$ -Actin	Cell Signal	4967
Polyclonal Swine Anti-Rabbit Immunoglobulins/TRITC	Dako	R0156
Polyclonal Rabbit Anti-Mouse Immunoglobulins/TRITC	Dako	R0270
Anti-mouse IgG, HRP-linked Antibody	Cell Signal	7076
Anti-rabbit IgG, HRP-linked Antibody	Cell Signal	7074