Bilateral macular coloboma and pigmented paravenous retinochoroidal atrophy

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Abstract

A patient had bilateral macular coloboma with aggregations of pigment clumps located perivascularly, predominantly paravenously, and in other parts of the retina. The *Toxoplasma* IgG antibody was negative. The diagnosis of bilateral macular coloboma with pigmented paravenous retinochoroidal atrophy was made and seemed to be a developmental abnormality in origin.

Macular coloboma is thought to result from intrauterine inflammation or as an abnormality of development. Most cases are now recognised as being due to an intrauterine infection with *Toxoplasma gondii*. A developmental abnormality seems to be the cause in those patients with a hereditary or family origin,^{1,2} and those with other ocular or systemic abnormalities.³⁻⁸

We report a case of bilateral macular coloboma associated with pigmented paravenous retinochoroidal atrophy and negative *Toxoplasma* IgG antibody in which the macular lesions seem to be of a developmental origin. To our knowledge the appearance of these two congenital anomalies has not been previously reported in the English literature.

Case report

A 23-year-old woman presented with a history of progressive blurring of vision in both eyes for a duration of 8 years. There was no contributory medical or family history.

Examination of the patient revealed the best corrected visual acuity as 20/400 in both eyes. The anterior segments were normal. Ophthalmoscopy showed a bilateral excavated, non-pigmented macular coloboma about 4×4 disc diameter in size with ectatic sclera at the base. A few large choroidal vessels were visible at the base and the retinal vessels crossing the defect were attenuated. The optic discs were normal. Aggregations of pigment clumps were located perivascularly, mainly paravenously, and in some other parts of the retina. Zones of peripapillary and radial patches of chorioretinal atrophy were situated behind the pigmented muffs and mainly along the vessels (Figs 1 and 2). Fluorescein angiography showed hypofluorescence with filling of the choroidal vasculature in both coloboma areas with a rim of hyperfluorescence surrounding the margin in the later phase of the examination. There were also blocked fluorescence from the pigment clumps and the pigment epithelial window defect due to pigment epithelial atrophy (Figs 3 and 4). The Farnsworth-Munsell 100-hue test showed a mild colour vision defect. Goldmann perimetry examination revealed central scotoma with constricted peripheral fields. Electroretinography showed a reduction in b-wave amplitude. Dark adaptation curves showed monophasic and elevated final rod thresholds in both eyes. Ultrasonography showed a bilateral ectatic base with an axial length of 21.8 mm in the right eye and 22.0 mm in the left eye.

A systemic evaluation, including skull and chest x rays, complete blood count, erythrocyte sedimentation rate, antinuclear antibody,

Figure 1 Retinal photograph of right fundus.

Figure 2 Retinal photograph of left fundus.

Figure 3 Fluorescein angiography of right fundus.

Figure 4 Fluorescein angiography of left fundus.

C-reactive protein, and VDRL test was non-contributory. Serological tests for cytomegalovirus, rubella, and herpes simplex were negative. A *Toxoplasma* IgG antibody test was negative at 1 in 10 dilutions by using an indirect fluorescence test.

Discussion

The patient described herein showed a bilateral macular coloboma and perivascular, mainly paravenous, aggregations of pigment clumps.

Mann⁹ classified macular coloboma into three types, namely pigmented macular coloboma, non-pigmented macular coloboma, and macular coloboma associated with abnormal vessels. The present case did not fall clearly into any of these three types. Though the macular lesion seemed to be of the non-pigmented type paravenous pigment aggregations showed a peculiar manifestation.

Macular coloboma is known to develop from an intrauterine inflammation or as a developmental abnormality of the eye. The majority of cases are due to intrauterine *Toxoplasma* infection.¹⁰ However the infective process does not produce macular coloboma in families or in

association with other ocular or systemic abnormalities such as retinitis pigmentosa,³ Leber's amaurosis,⁴ retinal dystrophy,⁵ idiopathic infantile hypercalciuria,⁶ or a specific skeletal abnormality.^{7,8} An indirect fluorescent test for measuring serum anti-*Toxoplasma* IgG titre was negative in this patient at 1 in 10 dilution. Desmonts¹¹ emphasised the importance of calculating the ratio of the antibody titres between aqueous and serum to make the diagnosis. We did not perform anterior chamber paracentesis since we thought it was invasive and might be harmful to the patient. Though the negativity of the serum antibody test did not exclude the presence of a *Toxoplasma* infection the association with peculiar retinal changes in this case revealed that it was unlikely that the macular lesions were due to a previous *Toxoplasma* infection.

Pigmented paravenous retinochoroidal atrophy is characterised by the presence of perivascular, predominantly paravenous, aggregations of pigment clumps associated with peripapillary and radial zones of chorioretinal atrophy and is thought to be degenerative in origin.¹²⁻¹⁵ Though some parts of the retina in this patient showed chorioretinal atrophy the marked perivascular pigment aggregations and chorioretinal atrophy were similar to that of the disease.

The present case showed a bilateral macular coloboma with the perivascular existence of pigment clumps that seemed to result from a common insult affecting their developmental process and could not be explained by a single *Toxoplasma* macular lesion. The *Toxoplasma* IgG antibody test was negative in this patient. Though the aetiology of this case is still unknown the presence of a bilateral macular coloboma, pigmented paravenous retinochoroidal atrophy, and negative laboratory examinations lead us to speculate that it is a developmental abnormality in nature.

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