Chlamydial immunofluorescence serology in anterior uveitis

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SUMMARY  Chlamydial immunofluorescence (IF) serology was determined in 118 patients with acute anterior uveitis (AAU), half of whom had anti-chlamydia antibodies. Men and women were equally affected. The chlamydial IF antibody titre was generally 1/16-64, but a few higher values were also measured. The age distribution, ocular pathology, and duration of AAU were identical in the chlamydial seropositive and seronegative patient groups. In the seropositive group there were 3 men with Reiter’s syndrome. The prevalence of anti-chlamydia antibodies (50%) was higher than in other patients studied in the same laboratory. Further population studies are in progress to see whether our AAU patients represent an accumulation of chlamydial-positive persons.

Infections due to Chlamydia trachomatis serotypes D to K are sexually transmitted and are increasingly common among persons aged 20-40 years. In the same age group the frequency of acute anterior uveitis (AAU) is greatest.1

Specific micro-organisms, such as klebsiella, yersinia, and salmonella, are thought to be involved in the development of AAU.2-5 Chlamydia has also been linked to AAU, which may occasionally develop either during the acute course of a chlamydial conjunctivitis6-8 or months after a conjunctival infection.6,7 Intraocular inflammation may present as the ocular component of Reiter’s syndrome, of which chlamydia may be one principle aetiological factor.7,9,10

This study reports the serological findings, as measured by an indirect immunofluorescence test (IF), and conjunctival isolations of C. trachomatis in patients with AAU. Part of the study has been previously published.11

Patients and methods

The subjects of the study were 118 patients with AAU treated at the Outpatient Clinic of the Department of Ophthalmology, Turku University Central Hospital, from January 1978 to December 1979. Patients who had AAU due to ocular trauma or concomitant conjunctival or corneal diseases were excluded from the study. Only one patient had a history of genital chlamydial infection.

In biomicroscopic ocular examination the quantity and the quality of keratic precipitates, aqueous flare and cells, and vitreous inflammatory signs were recorded. Applanation tonometry and ophthalmoscopy were performed in all cases.

The following laboratory tests were carried out for each patient: erythrocyte sedimentation rate (ESR), peripheral blood leucocyte count, the latex and Waaler-Rose test for rheumatoid factor, antistreptolysin O, antistaphylokins, antiyersinia antibody and antichlamydia antibody determinations, conjunctival swabbing from both eyes for chlamydia isolation, and x-ray examination of the chest, sinuses, and lumbosacral spine. In some cases serological tests for toxoplasma, rubella, Mycoplasma pneumoniae, and antinuclear antibodies, and the isolation of gonorrhoeac from the urethra were carried out. The hospital records were reviewed with attention to the possible recurrences of AAU and associated diseases.

Antichlamydia antibody determinations were carried out by immunofluorescence test. The technique has been described previously.12 Briefly, irradiated McCoy cells on a slide were infected with a 0-5 ml of C. trachomatis (L2) inoculum to produce approximately 5000 inclusions. The slide was divided with nail polish into 32 squares, on which serial dilutions of serum were pipetted. After incubation
and washing a fluorescein isothiocyanate (FITC) conjugated pretitrated working dilution (usually 1/40) of rabbit antihuman IgG (Dako, Denmark) was added and again incubated in a moist chamber. After washing and drying the slide a cover glass was mounted with Depex and standard FITC microscopy was performed. The serum titre was recorded as the highest dilution that showed clear inclusion body fluorescence. The dilution of 1/16 was considered to be reactivity of the test.

The isolation technique for C. trachomatis has been described in detail previously. Briefly, irradiated McCoy cells grown on a cover glass in flat-bottomed tubes were infected by centrifugation (15,000 g for 1 hour at 35°C). After incubation for 72 hours in CO₂ at 35°C the cells were stained with iodine. Chlamydial inclusions were identified by light microscopy.

Results

The isolation of C. trachomatis from bilateral conjunctival swabings was negative in all patients. Out of the 118 AAU patients 59 (50%) had antibody titres ≥ 1/16 against chlamydia.

The age distribution in the seropositive and seronegative AAU patient groups was identical (Fig. 1). The mean age of chlamydial seropositive patients was 36 years (range 5-73) and that of chlamydial seronegative AAU patients 39 years (range 16-77 years). Both sexes were equally distributed among seropositive and seronegative groups. There were 31 (52.5%) men and 28 (47.5%) women in the seropositive and 35 (59.3%) and 24 (40.7%) in seronegative groups respectively.

The chlamydial IF antibody titres in the 59 chlamydial seropositive AAU patients were generally 1/16-64, but higher titres were also measured. The distribution of antichlamydia antibody titres was identical in both sexes and in different age groups.

**Clinical pictures and features of AAU**

In the 59 chlamydial seropositive and seronegative AAU patients the ocular inflammation was recurrent in 17 and 14 patients respectively. The relapse was in the same eye in 8 seropositive and in 2 seronegative patients. During the follow-up period AAU was bilateral in 4 and 5 seropositive and seronegative patients, respectively.

The ocular pathology in patients who had antichlamydia antibodies did not differ from that in the seronegative cases of AAU. There was no relationship between the intensity of the uveitis and the height of the antichlamydia antibodies. Keratic precipitates (never mutton-fat) flare and cells in the anterior chamber were seen to the same extent in both groups. One chlamydial seropositive patient had hypopyon and one hyphaema. Fibrin in the anterior chamber was present at the beginning of AAU in 6 seropositive and in 13 seronegative cases. Posterior synchiae were noticed during the disease in 18 seropositive and 22 seronegative patients, and the pupil remained irregular in 7 and 10 cases respectively.

In the anterior vitreous there were cells in 17 seropositive and 7 seronegative cases. In both groups there were 3 eyes with macular oedema, and one seropositive patient had papillary oedema. In 5 eyes in both groups the intraocular pressure was ≥ 21 mmHg and in 12 eyes in the seropositive AAU group and in 10 eyes in the seronegatives ≤ 8 mmHg. The duration of the uveitis was almost same in both groups. The mean duration was 6-6 weeks in the seropositive AAU and 5-7 weeks in the seronegative AAU group.

**General patient status**

The positive laboratory and x-ray findings other than chlamydial are summarised in Table 1. X-ray examination confirmed 5 cases in the chlamydial

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Table 1  Number of positive laboratory and x-ray examinations in the chlamydial seronegative and seropositive AAU patient groups

<table>
<thead>
<tr>
<th>Laboratory test and x-ray examinations</th>
<th>Serum IgG IFAT titre</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Number of patients</td>
</tr>
<tr>
<td></td>
<td>≤ 1/16</td>
</tr>
<tr>
<td></td>
<td>≥ 1/16</td>
</tr>
<tr>
<td>---</td>
<td></td>
</tr>
<tr>
<td>ESR ≥ 20</td>
<td>17</td>
</tr>
<tr>
<td>Leucocytes ≥ 10x10⁶/l</td>
<td>5</td>
</tr>
<tr>
<td>Latex + + +</td>
<td>1</td>
</tr>
<tr>
<td>Waaler-Rose ≥ 64</td>
<td>2</td>
</tr>
<tr>
<td>AST titre ≥ 350</td>
<td>11</td>
</tr>
<tr>
<td>Antiyersinia antibodies</td>
<td>4</td>
</tr>
<tr>
<td>X-ray: chest</td>
<td>1</td>
</tr>
<tr>
<td>sinuses</td>
<td>4</td>
</tr>
<tr>
<td>lumbosacral spine</td>
<td>8</td>
</tr>
</tbody>
</table>

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Fig. 1  Age distribution of chlamydial seropositive (titre ≥1/16) and seronegative (titre ≤ 1/8) AAU patients.
seropositive and 6 cases in the chlamydial seronegative AAU group with sacroiliitis and 2 cases in both groups with other abnormalities in the lumbo-sacral spine. All the positive x-ray findings in sinuses and chest were classified as inflammatory by the roentgenologist. In both groups there was one patient with psoriasis vulgaris. One patient with anti-chlamydia antibodies had erythema nodosum 3 months before the onset of AAU.

In the seropositive group there were 3 men with Reiter’s syndrome. They all had fresh arthritis during the AAU and had had symptoms of urethritis, prostatitis, and epididymitis—namely, discharge with and without dysuria—for about 6 months. Two of them had had gonorrhoea and one of them keratitis twice. During the urethritis the patients were treated with systemic tetracycline and/or erythromycin. There was no urethral chlamydial isolation. During the AAU all diagnostic tests except that for chlamydial blood serology were negative. Two of them had antichlamydia antibody titres of 1/512 and the third 1/256. Before the onset of ocular inflammation from one patient with Reiter’s syndrome an anti-chlamydia antibody titre of 1/512 was measured from the synovial fluid of the knee.

**OCULAR TREATMENT**

All AAU cases were treated with topically applied cycloplegics and corticosteroids. In 6 of the sero-positive patients and 5 of the seronegative the AAU was so severe that systemic corticosteroid treatment was given.

**SYSTEMIC TREATMENT**

Five patients with high antichlamydia antibodies were treated with systemic tetracycline. Three of them had Reiter’s syndrome. In the chlamydial seropositive and seronegative groups one patient had systemic tetracycline therapy for acute yersiniosis. The indication for systemic tetracycline therapy in 3 seronegative patients was inflammatory infiltrates in the chest and sinuses and cementoperiostitis. In both groups 2 patients who had elevated AST titres were treated with systemic penicillin. One child with antichlamydia antibodies and with sinusitis was treated with systemic erythromycin and the one with pulmonary infiltrates with amoxicillin. All the patients with an inflammatory process recovered well.

**Discussion**

In our study AAU observed in patients with anti-chlamydia antibodies differed neither clinically nor epidemiologically from AAU observed in patients with no antichlamydia antibodies. In both patient groups other positive laboratory tests and positive x-ray examinations for an aetiological diagnosis of AAU were equally represented. Patients with anti-chlamydia antibodies included 3 men with Reiter’s syndrome, which was the only clinical difference observed between the chlamydial positive and negative patient groups.

AAU patients with and without antichlamydia antibodies had in 6 and in 4 cases, respectively, concomitantly raised levels of anti-yersinia antibodies in their sera. The appearance of antibodies against both yersinia and chlamydia indicates double infection in these patients.4

In our AAU patient material the prevalence of antichlamydia antibodies was high (50%). The prevalence of antichlamydial antibodies in adults (London blood donors) has been reported to be 17% in women and 26% in men.4 Among pregnant Swedish women a serological study of blood samples has shown every second woman to have antichlamydia antibodies with a titre ≥ 1/32.15 In our patients, in Finnish army recruits,16 and in Finnish women attending maternal care units (P. Terho, personal communication) the percentage of chlamydial positive persons has been notably smaller, 28% and 20% respectively. The antibody determinations in our studies was made by the same laboratory in all cases. Further population studies are in progress to determine whether our AAU material represents an accumulation of chlamydial positive persons.

In experimental animals and in man chlamydial organisms have been an aetiological factor for intraocular inflammation. In rabbits intracameral inoculation of chlamydia isolated from synovial membrane of a patient with Reiter’s syndrome induced iritis.17 The same isolate injected into the knees of rabbits induced chronic arthritis, which was later complicated by iritis.18

Human chlamydial conjunctivitis has been complicated by AAU.6-8 In some cases the uveitis developed in the course of conjunctival reinfection7 or several months after the conjunctival infection had subsided.7 In Reiter’s syndrome chlamydia was isolated from the patients’ conjunctiva, and the patients had significantly elevated antichlamydia antibodies in their sera.9-10 In our material all the chlamydia isolation attempts from the conjunctiva were negative. This excludes the possibility of conjunctival reinfection by chlamydia as a trigger mechanism for AAU. Patients with conjunctival or corneal diseases were excluded from our series; none of the AAU cases was directly caused by chlamydial conjunctivitis.

In recent years there has been growing evidence for the possible role of Gram-negative organisms in the generation of AAU in genetically predisposed persons.14 This role is connected with structural
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similarities and cross-reactivity of bacteria with tissue antigens of the eye. Chlamydia have structural similarities with Gram-negative bacteria, and they might also share common immunobiological functions.

If chlamydia can be proved to have immunological properties similar to those of, for example klebsiella, they might be one alternative in the pathogenesis of postinfectious complications leading, by some unknown trigger mechanism, to inflammation in the eye. Further experimental and population studies are needed to evaluate this hypothesis.

References