Monoclonal origin of localised orbital amyloidosis detected by molecular analysis

Sylvia Pasternak, Valerie A White, Randy D Gascoyne, Stephen R Perry, Royce L C Johnson, Jack Rootman

Abstract
Aims—Primary localised orbital amyloidosis is a rare disease. The purpose of this study was to describe two cases of primary orbital amyloidosis and emphasise the value of molecular analysis of immunoglobulin gene rearrangement in identifying a monoclonal population of cells responsible for the amyloid production.

Methods—Charts and biopsy specimens of each case were reviewed. Conventional light microscopy, immunohistochemistry, and polymerase chain reaction (PCR) analysis for immunoglobulin gene rearrangement were performed in both cases.

Results—An unusual presentation of localised primary amyloidosis with bilateral and extensive enlargement of multiple extraocular muscles was seen in case 1. The presence of amyloid deposits was confirmed by biopsy in both cases. Evidence of a monoclonal population of plasma cells was shown by immunohistochemical analysis in case 2 only. The monoclonal origin of the cells responsible for the amyloid deposition was determined by PCR analysis demonstrating immunoglobulin heavy chain gene rearrangement in both cases.

Conclusions—A monoclonal population of plasma cells responsible for the amyloid deposition was present in these two cases. PCR analysis is extremely helpful in determining monoclonality, a finding that may have important therapeutic and prognostic implications.

Materials and methods
Charts were reviewed for details of the patients’ presentation and history. Haematoxylin and eosin stained slides were obtained from formalin fixed paraffin embedded tissue samples. Sections from the paraffin blocks were also examined immunohistochemically by the streptavidin–biotin peroxidase technique using commercially available polyclonal antibodies to the following antigens: IgG (1:10 000; Dako, Glostrup, Denmark), IgA (1:1000; Dako), IgM (1:2000; Dako), kappa light chains (1:4000; Dako), lambda light chains (1:8000; Dako).

For PCR analysis of case 1, DNA was extracted from fresh frozen material on an automated DNA extractor (Applied Biosystems, model 341, Foster City, CA, USA). For case 2, DNA was extracted from paraffin embedded tissue sections using proteinase K in buffer following deparaffinisation in xylene and redissolution in graded alcohols. DNA 0.5 μg from case 1 and 10 μl diluted DNA from case 2 were used in a 50 μl PCR reaction mixture. A consensus PCR strategy was employed using V region and J region primers for the Ig heavy chain locus. Thirty five rounds of amplification were used and the PCR products visualised on an 8% polyacrylamide gel. The sizes of the monoclonal bands were compared with the 123 base pair marker. Clonal VDJ IgH rearrangements appear as tight bands ranging in size from 70 to 140 base pairs. The specimens were analysed with a monoclonal B cell leukaemia/lymphoma KM3 cell line (positive control) and a polyclonal reactive lymph node (negative control).

Results
Case 1
A 44-year-old white woman developed typical symptoms of hyperthyroidism when aged 33. She was found to have elevated thyroid hormone levels and was treated with radioactive iodine. Within 1 year of radioactive iodine therapy, she presented with bilateral ocular irritation and protruding eyes. Over the next 7 years the proptosis gradually increased from an initial Hertel measurement of 21 mm to 28 mm bilaterally. She underwent bilateral orbital decompression by a medial approach, achieving only 3 mm recession on the right and 5 mm on the left. Her soft tissue changes progressed and she underwent bilateral mullerectomies, blepharoplasties, and right dacryocystorhinostomy for epiphora following the decompression.
She was referred to the University of British Columbia Orbit Clinic for second opinion 9 years into her orbital disease. Review of her previous computed tomography (CT) scans showed progressive deviation from the usual pattern of thyroid orbitopathy. The muscles were very irregularly shaped and enlarged, and both the orbital fat and tendons were also involved (Fig 1).

On ophthalmic examination her visual acuity was 20/25 bilaterally. Both upper lids showed some brawny induration and the right lacrimal gland was palpable. The conjunctiva was minimally injected on the right. There was downward displacement of the right globe by 2 mm and bilateral exophthalmos with Hertel measurements of 25 mm on the right and 23 mm on the left. A mild right lateral rectus underaction was present. The intracocular pressure was 15 mm Hg in primary position but elevated to 25 mm Hg on upgaze bilaterally. Fundus examination was unremarkable. Full clinical examination, blood tests, bone marrow biopsy, and CT scan of the thorax and abdomen were normal.

Orbital biopsy was suggested because a lymphoproliferative disorder or a sclerosing lesion were considered as possible diagnoses.

Biopsy of the orbital mass and right lateral rectus muscle showed extensive amorphous eosinophilic material that stained positively with Congo red and showed apple green birefringence under polarised light (Fig 2A). Numerous blood vessels were surrounded by collections of plasma cells, admixed with occasional lymphocytes (Fig 2B). Immunoperoxidase stains for immunoglobulin heavy and light chains did not show preferential staining. However, material submitted for PCR analysis showed a monoclonal rearrangement of the immunoglobulin heavy chain gene as evidenced by a band in the appropriate size range (Fig 3). This finding indicated the presence of a monoclonal population of plasma cells, presumably responsible for the amyloid production, and consistent with primary (AL type) amyloidosis.

The patient was treated with radiotherapy (3000 cGy) to the right orbit, which was the side most affected. The proptosis regressed and no recurrence has developed after a follow up of 22 months. A discrete mass developed in the left orbit 18 months after the first biopsy. This was biopsied and showed similar histology. The left orbit was also treated with radiotherapy and there has been no recurrence after 3 months.

Figure 1. Case 1. Axial computed tomography showing bilateral enlargement of the extraocular muscles, more prominent in the right orbit. The extraocular muscles are thickened and irregular; the muscle tendons are also involved.

Figure 2. (A) Case 1. Apple green birefringence under polarised light after staining with Congo red, confirming the amyloid nature of the deposits (×400). (B) Case 1. High power photomicrograph showing collections of plasma cells around the blood vessels (haematoxylin and eosin; ×400). (C) Case 2. Staining of the majority of the cells with antikappa antibody (haematoxylin counterstain, ×400). (D) Case 2. Staining of few cells with anti-lambda antibody (haematoxylin counterstain, ×400).
Monoclonal origin of localized orbital amyloidosis detected by molecular analysis

Monoclonal origin

Amyloidosis

Down's syndrome

Familial amyloid polyneuropathy

Dialysis cardiomyopathy

Cerebral amyloidosis

Hereditary amyloid polyneuropathy

Primary amyloidosis

Amyloidosis associated with multiple myeloma

Reactive amyloidosis

Familial Mediterranean fever

Familial amyloid polyneuropathy (type I, II)

Familial amyloidotic cardiomyopathy

Senile systemic amyloidosis

Familial amyloid polyneuropathy (type III) (Iowa)

Familial amyloid polyneuropathy (type IV) (Finnish)

Hereditary cerebral haemorrhage with amyloidosis (Icelandic)

Alzheimer's disease

Down's syndrome

Hereditary cerebral haemorrhage with amyloidosis (Dutch)

Dialysis related amyloidosis

Amyloidosis associated with medullary carcinoma of the thyroid

Table 1 Clinical amyloidosis syndromes and corresponding amyloid fibril

<table>
<thead>
<tr>
<th>Clinical syndrome</th>
<th>Fibroblastic protein</th>
<th>Origin</th>
</tr>
</thead>
<tbody>
<tr>
<td>Primary amyloidosis</td>
<td>AL</td>
<td>Monoclonal immunoglobulin light chains</td>
</tr>
<tr>
<td>Amyloidosis associated with multiple myeloma</td>
<td>AL</td>
<td>Monoclonal immunoglobulin light chains</td>
</tr>
<tr>
<td>Reactive amyloidosis (secondary amyloidosis)</td>
<td>AA</td>
<td>Serum amyloid A (SAA) = acute phase reactant</td>
</tr>
<tr>
<td>Familial Mediterranean fever</td>
<td>AA</td>
<td>Serum amyloid A (SAA) = acute phase reactant</td>
</tr>
<tr>
<td>Familial amyloid polyneuropathy (type I, II)</td>
<td>ATTR</td>
<td>Transthyretin</td>
</tr>
<tr>
<td>Familial amyloidotic cardiomyopathy</td>
<td>ATTR</td>
<td>Transthyretin</td>
</tr>
<tr>
<td>Senile systemic amyloidosis</td>
<td>ATTR</td>
<td>Transthyretin</td>
</tr>
<tr>
<td>Familial amyloid polyneuropathy (type III) (Iowa)</td>
<td>AApoAI</td>
<td>Apolipoprotein Al</td>
</tr>
<tr>
<td>Familial amyloid polyneuropathy (type IV) (Finnish)</td>
<td>Agel</td>
<td>Gelsolin</td>
</tr>
<tr>
<td>Hereditary cerebral haemorrhage with amyloidosis (Icelandic)</td>
<td>ACys</td>
<td>Ceratin C</td>
</tr>
<tr>
<td>Alzheimer's disease</td>
<td>A</td>
<td>Protein</td>
</tr>
<tr>
<td>Down's syndrome</td>
<td>A</td>
<td>Protein</td>
</tr>
<tr>
<td>Hereditary cerebral haemorrhage with amyloidosis (Dutch)</td>
<td>A</td>
<td>Protein</td>
</tr>
<tr>
<td>Dialysis related amyloidosis</td>
<td>2-microglobulin</td>
<td>2-microglobulin</td>
</tr>
<tr>
<td>Amyloidosis associated with medullary carcinoma of the thyroid</td>
<td>Precalcitonin</td>
<td>Precalcitonin</td>
</tr>
</tbody>
</table>

Stained and had the birefringent qualities of amyloid. A few patches of plasma cells and lymphocytes were present. Immunoperoxidase stains revealed preferential staining for kappa light chains (Fig 2C and 2D). PCR analysis showed a monoclonal immunoglobulin heavy chain gene rearrangement, confirming a monoclonal population of plasma cells (Fig 3).

The patient was treated with radiotherapy (3000 cGy). The left orbital lesion regressed and there is no evidence of recurrence after a follow up of 15 months.

Discussion

Amyloidosis comprises a variety of different disease processes in which extracellular deposition of abnormal protein fibrils occur. These fibrils are derived from different proteins in different forms of the disease. All amyloid fibrils are arranged in a β pleated sheet formation, which produces the characteristic apple green birefringence under polarised light, following staining with Congo red. Recently, precise identification and characterisation of the different types of fibril proteins that constitute amyloid deposits has become possible and amyloidosis can now be classified, not only based on clinicopathological features, but also on the nature of the amyloid protein (Table 1).11-16

In the orbit amyloidosis may be localised or part of a systemic disorder.17 18 Systemic amyloidosis involving the orbit may be primary or associated with multiple myeloma. It usually results from deposition of AL amyloid, which is a protein derived from monoclonal immunoglobulin light chains.19-21 An important clinical differential point with respect to the likelihood of systemic disease is involvement of the skin of the eyelids with a waxy yellow subcutaneous deposit, frequently associated with spontaneous haemorrhage, which is almost always related to a systemic process.17

Localised amyloidosis may produce a diffuse deposition, or may form nodules or tumoral amyloidosis (so called amyloidomas).22 Amyloidomas have been described in many sites including the respiratory tract,21-25 genitourinary tract,26-27 spleen,28 breast,29 soft tissues,20 brain,30-31 gastrointestinal tract,32 and heart.33

Although in most of these cases the specific type of amyloid fibril has not been definitely established, the presence of a lymphocytic and/or plasma cell infiltration,34 24 25 29 31 35 sug-
gests AL amyloid deposition. In addition, monoclonality could be identified by immuno-

histochemistry in some cases.\textsuperscript{22,23,27,7} The treatment usually consisted of surgical removal of the amyloid mass and, in the cases where follow up information is available, no systemic diseases developed.\textsuperscript{21,25,28,33} However, local recurrences were not uncommon.\textsuperscript{21,22}

Localised primary orbital amyloidosis is rare. Table 2 outlines the 23 localised orbital amyloidosis cases reported in the literature. Most of the cases involved the anterior orbit. There are five cases involving the lacrimal gland,\textsuperscript{1,9} one case with amyloidosis isolated to one extraocular muscle,\textsuperscript{4} and one case with amyloid involvement of two extraocular muscles.\textsuperscript{1} Five patients had bilateral orbital involvement.\textsuperscript{1}

Many cases of orbital amyloidosis show infiltration of plasma cells suggesting production of immunoglobulins as the basis for the amyloid deposition.\textsuperscript{1,8} However, only in four recent orbital cases was an attempt made to identify the specific type of amyloid fibril involved, by immuno-

histochemistry,\textsuperscript{4} immunoelectron microscopy,\textsuperscript{8} and extraction and isolation of the amyloid protein.\textsuperscript{9} In these cases amyloid derived from immunoglobulins was encountered, similar to what has been observed in localised amyloidosis elsewhere in the body.\textsuperscript{20,21,24,27} Although immuno-

histochemical analysis may show evidence of monoclonality in some cases, as in our case 2, it may fail in others, as in our case 1. PCR analysis definitely established the monoclonal origin of the plasma cells in our two cases. This finding further contributes to the concept of amyloid derived from immunoglobulins as the pathogenic mechanism in localised orbital amyloidosis.

Our case 1 is unique in the fact that there is bilateral orbital amyloidosis with extensive enlargement of multiple extraocular muscles which, to our knowledge, has not been previously reported. The patient’s antecedent thyroid disease raised the possibility of AA amyloid (reactive amyloid) deposition secondary to the inflammation related to thyroid orbitopathy. However, if that was the case, a monoclonal population of plasma cells should not be present. In addition, there was no histological evidence of thyroid orbitopathy on the biopsied right lateral rectus muscle.

Some features of localised orbital amyloidosis should be emphasised. Firstly, the amyloid usually extends deeply into the orbit and with an infiltrative pattern, making total surgical excision difficult. Secondly, although no systemic disease developed in the cases where follow up information is available, local recurrences were very frequent.\textsuperscript{1,4} Therefore, therapy aimed at eradicating the neoplastic plasma cells might be considered to arrest the process. We believe that it is reasonable to treat localised amyloidosis with radiotherapy or, perhaps, chemotherapy, if progression is documented and a monoclonal population of cells can be identified.

Both of our patients were treated with radiotherapy. No major side effects occurred but evaluation of the results will require a longer follow up period.


None of the authors has proprietary or financial interest in any product or company mentioned in this study.

1 Nehen JH. Primary localized orbital amyloidosis. Acta Ophthal-


2 Cohen MM, Lessell S. Amyloid tumor of the orbit. Neuro-


5 Levine M, Buckman G. Primary localized orbital amyloidosa-


### Table 2 Reported cases of localised orbital amyloidosis

<table>
<thead>
<tr>
<th>Author</th>
<th>No of cases</th>
<th>Location</th>
<th>Treatment</th>
<th>Follow up</th>
<th>Type of amyloid</th>
<th>Lympocytic/plasma cell infiltration</th>
</tr>
</thead>
<tbody>
<tr>
<td>Nehen\textsuperscript{1} (1979)</td>
<td>14 (4 bilateral)</td>
<td>Orbit (11)</td>
<td>Surgical excision</td>
<td>5 cases: recurrence</td>
<td>NA</td>
<td>Plasma cells present in 7 cases</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Lacrimal gland (3)</td>
<td></td>
<td>2 cases: no recurrence</td>
<td>NA</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>7 cases: lost to follow up</td>
<td>NA</td>
<td></td>
</tr>
<tr>
<td>Cohen\textsuperscript{2} (1979)</td>
<td>1</td>
<td>Inner quadrant of the right orbit</td>
<td>Surgical excision</td>
<td>Recurrence after 2 years</td>
<td>NA</td>
<td>Scattered plasma cells</td>
</tr>
<tr>
<td>Finlay\textsuperscript{3} (1980)</td>
<td>1</td>
<td>Left orbit</td>
<td>Surgical excision</td>
<td>Recurrence after 8 months</td>
<td>NA</td>
<td>Few plasma cells</td>
</tr>
<tr>
<td>Lucas\textsuperscript{4} (1982)</td>
<td>2</td>
<td>Right upper orbit (1 case)</td>
<td>Surgical excision</td>
<td>Recurrence after 1 month</td>
<td>AL</td>
<td>Lymphocytes and occasional plasma cells</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Left anterior lower orbit (1 case)</td>
<td>Surgical excision</td>
<td>Recurrence after 2 years</td>
<td>AL</td>
<td>Lymphocytes and plasma cells</td>
</tr>
<tr>
<td>Levine\textsuperscript{5} (1986)</td>
<td>1</td>
<td>Right lacrimal gland</td>
<td>Surgical excision</td>
<td>NA</td>
<td>NA</td>
<td>Lymphocytes and occasional plasma cells</td>
</tr>
<tr>
<td>Holmstrom\textsuperscript{6} (1987)</td>
<td>1</td>
<td>Right medial rectus muscle</td>
<td>Strabismus surgery</td>
<td>NA</td>
<td>NA</td>
<td>No information</td>
</tr>
<tr>
<td>Eris\textsuperscript{7} (1989)</td>
<td>1</td>
<td>Right horizontal rectus muscle</td>
<td>Strabismus surgery</td>
<td>Stable after 26 months</td>
<td>NA</td>
<td>No information</td>
</tr>
<tr>
<td>Conlon\textsuperscript{8} (1991)</td>
<td>1 (bilateral)</td>
<td>Lacrimal gland</td>
<td></td>
<td>No information</td>
<td>NA</td>
<td>AL</td>
</tr>
<tr>
<td>Tan\textsuperscript{9} (1994)</td>
<td>1</td>
<td>Upper outer quadrant of right orbit</td>
<td></td>
<td>No information</td>
<td>NA</td>
<td>AH</td>
</tr>
</tbody>
</table>

NA= not available.
Monoclonal origin of localized orbital amyloidosis detected by molecular analysis