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Editorials

The "thin man" phenomenon: imperfect filling in of visual space

Normal people do not perceive an "empty" region in the portion of their visual field corresponding to the physiological blind spot. Instead, the visual system perceptually fills in the blind spot with information surrounding it. Similar types of perceptual filling in also occur in patients with pathological monocular and binocular homonymous scotomas.^{1,2} Furthermore, stabilising the image of a spot on the retina leads to perceptual filling in of the spot with the surrounding background.³ Perceptual filling in of stabilised images can also be demonstrated for colour, brightness, and texture.⁴⁻¹⁰ Safran *et al* have previously shown that a spatial distortion of images occurs in the field surrounding a scotoma.¹¹ They have now shown that the filling in phenomenon itself is not precise (see p 137, this issue). They report two patients with right inferior homonymous paracentral scotomas resulting from ischaemic brain insults in whom field defects were markedly smaller when tested with an Amsler grid than with a tangent (Bjerrum) screen. In addition, after both patients fixated another person's face or neck for 5-10 seconds, the other person's left shoulder (corresponding to the region of the field defect) appeared narrower than the right shoulder. Safran *et al* call this perceptual alteration "the thin man" phenomenon. Some patients with this alteration are aware of it; others are not, at least not until they are specifically questioned about it.

Filling in of a pathological homonymous scotoma is apparently related to expansion of receptive fields in the visual cortex,^{12,13} with the responsible neural mechanisms localised in retinotopic visual areas.³ The time required for this filling in to occur depends in part on the size, shape, and location of the scotoma. It appears to reflect the time required for figure ground segregation to fail rather than a slow spread of a surface feature from one region of the visual field into another,³ and it may be related to horizontal connections between pyramidal neurons in the extrastriate cortex.¹⁴

The findings reported here by Safran *et al* raise numerous questions with regard to the "thin man" phenomenon. Does it occur with homonymous field defects on both sides

of visual space or only with right sided defects? Does it occur only for homonymous paracentral scotomas or for more peripheral scotomas as well? Does the size of the defect influence the severity of the phenomenon and in what way? We look forward to further work by Safran and his colleagues as well as others for the answers to these and other questions. In the meantime, we agree with Safran *et al* that patients who complain of blurred vision, particularly after a known or suspected stroke, should be carefully tested for an homonymous paracentral scotoma, the clue to which may be the patient's spontaneous or queried observation of apparent asymmetry of a symmetric complex image in a particular region of visual space during the clinical examination.

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- 1 Bender MB, Teuber LH. Phenomena of fluctuation, extinction and completion in visual perception. *Arch Neurol Psychiatry* 1946;55:627-58.
- 2 Sergent J. An investigation into perceptual completion in blind areas of the visual field. *Brain* 1988;111:347-73.
- 3 De Weerd P, Desimone R, Ungerleider LG. Perceptual filling-in: a parametric study. *Vis Res* 1998;38:2721-34.
- 4 Gerrits HJM, de Haan B, Vendrik AJH. Experiments with retinal stabilized images. Relations between the observations and neural data. *Vis Res* 1966;6:427-40.
- 5 Riggs LA, Ratliff F, Cornsweet JC, *et al*. The disappearance of steadily fixated visual test objects. *J Opt Soc Am* 1953;43:495-501.
- 6 Troxler D. Über das Verschwinden gegebener Gegenstände innerhalb unseres Gesichtskreises. In: Himly K, Schmidt JA, Jena JA, eds. *Ophthalmologische Bibliothek II* 1804:1-119.
- 7 Yarbus AL. *Eye movements and vision*. New York: Plenum Press, 1967.
- 8 Ramachandran VS, Gregory RL. Perceptual filling-in of artificially induced scotomas in human vision. *Nature* 1991;350:699-702.
- 9 Ramachandran VS, Gregory RL, Aiken W. Perceptual fading of visual texture borders. *Vis Res* 1993;33:717-21.
- 10 Spillmann L, Kurtenbach A. Dynamic noise backgrounds facilitate target fading. *Vis Res* 1992;32:1941-6.
- 11 Safran AB, Duret F, Mermoud C, *et al*. Altered perception of distances with homonymous paracentral scotomas. *Vis Res* 1996;36:S214.
- 12 De Weerd P, Gattass R, Desimone R, *et al*. Responses of cells in monkey visual cortex during perceptual filling-in of an artificial scotoma. *Nature* 1995;377:731-4.
- 13 Pettet MW, Gilbert CD. Dynamic changes in receptive field size in cat primary visual cortex. *Proc Natl Acad Sci USA* 1992;89:8366-70.
- 14 Levitt JB, Yoshioka T, Lund JS. Intrinsic cortical connections in macaque visual area V2: evidence for interaction between different visual streams. *J Comp Neurol* 1994;342:551-70.

The healing optic nerve in glaucoma: transforming growth factor β and optic nerve head remodelling

Despite the many advances in the therapy and diagnosis of primary open angle glaucoma, and the recognition of intraocular pressure as the major modifiable risk factor,^{1,2} the pathogenesis of the disease still remains unclear. The most pathognomonic feature of glaucoma is optic disc cupping and the primary site of glaucomatous optic nerve damage appears to be at the optic nerve head.³ However, the exact mechanisms by which this damage occurs have not been elucidated. Changes in lamina cribrosa morphology and nerve fibre bundle pore morphometry^{4,5} have been documented in this disease, in association with alterations in the surrounding extracellular matrix.^{6–11} The tortuous course of the individual nerve fibres may also play a role.¹² In this issue of the *BJO* (p 209), Pena *et al* have found that the production of a growth factor, transforming growth factor β (TGF- β 2) is considerably increased in the optic nerve heads of patients with open angle glaucoma but not in normal individuals. This finding is important because of its implications for the pathogenesis of glaucomatous damage. Like all interesting research findings, it raises further intriguing questions. Is TGF- β 2 stimulated in response to nerve axon loss or changes in extracellular environment such as intraocular pressure induced compression, stretching, and shearing forces? Is TGF- β 2 production neuroprotective or do its effects on extracellular matrix compromise nerve fibre function and integrity?

The main structural component of the optic nerve head is the lamina cribrosa through which pass all the optic nerve fibres exiting the eye. It is continuous with the sclera, and consists of stacks of fenestrated connective tissue plates (cribriform plates), each pore allowing passage of nerve fibre axons.¹³ It has long been accepted that the lamina cribrosa is the weakest part of the sclera. If the intraocular pressure (IOP) is raised for a prolonged period, as in cases of chronic glaucoma, the lamina cribrosa bows outwards producing a “cupped” optic disc.¹⁴ This bowing can also be induced in the short term at higher levels of IOP. These structural changes are very similar even in patients with so called normal tension glaucoma, suggesting that physical forces at the nerve head do play a role in cupping, whatever the level of IOP. Histological analysis has demonstrated regional differences in lamina cribrosa structure with the superior and inferior areas being weakest and hence most susceptible to damage from raised IOP, with collapse and prominent posterior bowing in advanced glaucomatous disease.¹⁵ Differences in primary structure and cellular responses to “stress” including production of growth factors may explain in part the individual variation in the pattern of optic nerve damage seen with similar levels of IOP.

TGF- β is a multifunctional growth factor found throughout the body, and implicated in the processes of scarring.^{16–19} In the eye, TGF- β 2 appears to be the predominant isoform.^{20–23} Pena *et al* suggest the TGF- β 2 in glaucomatous eyes is produced by astrocytes in the lamina cribrosa, which as a consequence take up a “reactive” phenotype, seen characteristically in various neurological disorders—Alzheimer’s disease,²⁴ multiple sclerosis,²⁵ and after neuronal injury. This phenotype is implicated in the development of glial scar formation,²⁶ and in its remodelling. Although no gliosis is seen in glaucomatous optic neuropathy, they postulate that TGF- β may activate astrocytes to stimulate extracellular matrix remodelling of the

lamina cribrosa. TGF- β is a prominent component of the healing response to damage in many parts of the body, and the optic nerve head may be no exception.

The cellular responses at the optic nerve head to changes in IOP are not yet known. However, the effects of different types of forces on a variety of cell types has been studied, most extensively in endothelium, where shear stress has been shown to produce vasodilatation, a process mediated by nitrous oxide and probably a change in cell-cell interactions via expression of integrins.²⁷ Centrifugal tension has been shown to increase expression of growth factor receptors in fibroblasts, via stimulation of β 1 integrin expression,²⁸ and dermal fibroblasts alter production of matrix metalloproteinases (MMP) in response to different tensile loads.²⁹ It is therefore conceivable that changes in the IOP produce cellular responses in the lamina cribrosa, altering gene expression and the synthesis or degradation of extracellular components, and ultimately the support structure of the nerve fibres.

The idea that the optic nerve head may respond to dynamic physical force changes in its environment via alterations in TGF- β activity is very important. Historically, TGF- β was believed to stimulate scarring by inhibiting MMP production and stimulating tissue inhibitors of MMPs (TIMPs).³⁰ Work in our laboratory suggests, however, that the cellular effects of TGF- β depend on interactions with the surrounding extracellular matrix³¹—TGF- β inducing different MMP and TIMP profiles in different extracellular environments. Cell-matrix interactions are mediated via integrin receptors,³² and TGF- β is known to directly affect integrin expression, which in turn determines MMP production.³³ Hence, TGF- β is a potent modulator of extracellular remodelling.

Pena *et al* also comment on TGF- β having a neuroprotective effect in glaucoma. A number of possible mechanisms of neuroprotection have been reviewed by Flanders *et al* in the context of neurodegenerative diseases,³⁴ including antioxidant properties of TGF- β , its enhancement of mitochondrial potential, its maintenance of neuronal calcium homeostasis, and finally its inhibitory effects on apoptosis. Again, although these areas are currently being explored, the significance of these processes in glaucomatous optic neuropathy is not yet established.

As we approach the next millennium, this is an exciting period in glaucoma research. Technological advances in imaging have made it possible to visualise the lamina cribrosa pores *in vivo*.^{35,36} If we can now correlate *in vivo* features with changes at a cellular and molecular level, the processes occurring within the lamina cribrosa may provide us with an alternative tool not only to diagnose and monitor glaucoma but also to treat this fascinating but complex group of diseases.

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- 1 Sommer A. Glaucoma: facts and figures (Doyle lecture). *Eye* 1996;10:295–301.
- 2 Quigley HA. Number of people with glaucoma worldwide [see comments]. *Br J Ophthalmol* 1996;80:389–93.

- 3 Quigley HA, Hohman RM, Addicks EM, *et al.* Morphological changes in the lamina cribrosa correlated with neural loss in open-angle glaucoma. *Am J Ophthalmol* 1983;**95**:673–91.
- 4 Fontana L, Bhandari A, Fitzke FW, *et al.* In vivo morphometry of the lamina cribrosa and its relation to visual field loss in glaucoma. *Curr Eye Res* 1998; **17**:363–9.
- 5 Jonas JB, Mardin CY, Schottzer Schrehaur U, *et al.* Morphometry of the human lamina cribrosa surface. *Invest Ophthalmol Vis Sci* 1991;**32**:401–9.
- 6 Hernandez MR, Pena JDO. The optic nerve head in glaucomatous optic neuropathy. *Arch Ophthalmol* 1997;**115**:389–95.
- 7 Hernandez MR, Andrzejewska WM, Neufeld AH. Changes in the extracellular matrix of the human optic nerve in primary open-angle glaucoma. *Am J Ophthalmol* 1990;**109**:180–8.
- 8 Morrison JC, Dorman Pease ME, Dunkelberger GR, *et al.* Optic nerve head extracellular matrix in primary optic atrophy and experimental glaucoma. *Arch Ophthalmol* 1990;**108**:1020–4.
- 9 Quigley HA, Brown A, Dorman Pease ME. Alterations in elastin of the optic nerve head in human and experimental glaucoma. *Br J Ophthalmol* 1991;**75**:552–7.
- 10 Johnson EC, Morrison JC, Farrell S, *et al.* The effect of chronically elevated intraocular pressure on the rat optic nerve head extracellular matrix. *Exp Eye Res* 1996;**62**:663–74.
- 11 Fukuchi T, Sawaguchi S, Hara H, *et al.* Extracellular matrix changes at the optic nerve lamina cribrosa in monkey eyes with experimentally chronic glaucoma. *Graefes Arch Clin Exp Ophthalmol* 1992;**230**:421–7.
- 12 Morgan JE, Jeffery G, Foss AJE. Axon deviation in the human lamina cribrosa. *Br J Ophthalmol* 1998;**82**:680–3.
- 13 Anderson DR. Ultrastructure of human and monkey lamina cribrosa and optic nerve head. *Arch Ophthalmol* 1969;**82**:800–14.
- 14 Gray H. *Gray's anatomy*. 36th ed. Edinburgh: Churchill Livingstone, 1980.
- 15 Quigley HA, Addicks EM. Regional differences in the structure of the lamina cribrosa and their relation to glaucomatous optic nerve damage. *Arch Ophthalmol* 1981;**99**:137–43.
- 16 Ashcroft GS, Dodsworth J, van Boxtel E, *et al.* Estrogen accelerates cutaneous wound healing associated with an increase in TGF-beta1 levels [see comments]. *Nat Med* 1997;**3**:1209–15.
- 17 Shah M, Foreman DM, Ferguson MW. Neutralisation of TGF-beta 1 and TGF-beta 2 or exogenous addition of TGF-beta 3 to cutaneous rat wounds reduces scarring. *J Cell Sci* 1995;**108**(Pt 3):985–1002.
- 18 Levine JH, Moses HL, Gold LI, *et al.* Spatial and temporal patterns of immunoreactive transforming growth factor-beta-1, -beta-2 and -beta-3 during excisional wound repair. *Am J Pathol* 1993;**143**:368–80.
- 19 Merwin JR, Roberts A, Kondaliah P, *et al.* Vascular cell responses to TGF-beta3 mimic those of TGF-beta1 in vitro. *Growth Factors* 1991;**5**.
- 20 Lutty GA, Merges C, Threlkeld AB, *et al.* Heterogeneity in localization of isoforms of TGF-beta in human retina, vitreous and choroid. *Invest Ophthalmol Vis Sci* 1993;**34**:477–87.
- 21 Pasquale LR, Dorman Pease ME, Lutty GA, *et al.* Immunolocalisation of TGF-beta1, TGF-beta2 and TGF-beta3 in the anterior segment of the human eye. *Invest Ophthalmol Vis Sci* 1993;**34**:23–30.
- 22 Connor TB, Roberts AB, Sporn MB, *et al.* Correlation of fibrosis and transforming growth factor-beta type 2 levels in the eye. *J Clin Invest* 1989;**83**:1661–6.
- 23 Hales AM, Chamberlain CG, McAvoy JW. Cataract induction in lenses cultured with transforming growth factor-beta. *Invest Ophthalmol Vis Sci* 1995;**36**:1709–13.
- 24 Flanders KC, Lippa CF, Smith TW, *et al.* Altered expression of transforming growth factor-beta in Alzheimer's disease. *Neurology* 1995;**45**:1561–9.
- 25 Peress NS, Perillo E, Seidman RJ. Glial transforming growth factor (TGF)-beta isotypes in multiple sclerosis: differential glial expression of TGF-beta1, 2 and 3 isotypes in multiple sclerosis. *J Neuroimmunol* 1996;**71**:115–23.
- 26 Ridet JL, Malhotra SK, Privat A, *et al.* Reactive astrocytes: cellular and molecular cues to biological function. *Trends Neurosci* 1997;**20**:570–7.
- 27 Muller JM, Chilian WM, Davis MJ. Integrin signaling transduces shear stress dependent vasodilation of coronary arterioles. *Circulation Res* 1997;**80**:320–6.
- 28 Sundberg C, Rubin K. Stimulation of beta1 integrins on fibroblasts induces PDGF independent tyrosine phosphorylation of PDGF beta-receptors. *J Cell Biology* 1992;**119**:741–52.
- 29 Eastwood M, Porter R, Khan U, *et al.* Quantitative analysis of collagen gel contractile forces generated by dermal fibroblasts and the relationship to cell morphology. *J Cell Physiol* 1996;**166**:33–42.
- 30 Roberts AB, Sporn MB, Assoian RK, *et al.* Transforming growth factor beta: rapid induction of fibrosis and angiogenesis in vivo and stimulation of collagen formation in vitro. *Proc Natl Acad Sci USA* 1986; **83**:4167–71.
- 31 Cordeiro MF, Ocleston NL, Constable PH, *et al.* Effect of TGF-beta 1, 2 and 3 on human ocular fibroblast mediated collagen lattice contraction and matrix metalloproteinase production in lattices and monolayers. *Invest Ophthalmol Vis Sci* 1996;**37**(suppl):S1118.
- 32 Heino J, Ignatz RA, MEH, *et al.* Regulation of cell adhesion receptors by transforming growth factor-beta: Concomitant regulation of integrins that share a common beta1 subunit. *J Biol Chem* 1989;**264**:380–8.
- 33 Riikonen T, Westermarck J, Koivisto L, *et al.* Integrin-alpha2-beta1 is a positive regulator of collagenase (MMP-1) and collagen alpha1(I) gene expression. *J Biol Chem* 1995;**270**:13548–52.
- 34 Flanders KC, Ren RF, Lippa CF. Transforming growth factor-beta in neurodegenerative disease. *Progress Neurobiol* 1997;**54**:71–85.
- 35 Bhandari A, Fontana L, Fitzke FW, *et al.* Quantitative analysis of the lamina cribrosa in vivo using a scanning laser ophthalmoscope. *Curr Eye Res* 1997; **16**:1–8.
- 36 Fitzke FW. Colour imaging using a scanning laser ophthalmoscope. *Br J Ophthalmol* 1998;**82**:337–8.