Early drusen formation in the normal and aging eye and their relation to age related maculopathy: a clinicopathological study

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Abstract

Aim—To describe the early formation of drusen and their relation to normal aging changes at the macula and to the development of age related maculopathy (ARM).

Method—Histopathological features of 353 eyes without histological evidence of ARM are described and correlated with the clinical appearance. In addition, 45 of these eyes were examined by transmission electron microscopy.

Results—Drusen were detected histopathologically in 177 (50%) eyes but were seen clinically in only 34% of these. Drusen were mainly small hard drusen with an occasional soft distinct drusen: no soft indistinct drusen were seen. Only those drusen deposits larger than 25–30 µm in diameter were detectable clinically. Preclinical drusen in eyes with only an occasional drusen were seen on electron microscopy as entrapment sites of coated membrane bound bodies which formed adjacent to the inner collagenous zone of Bruch’s membrane. In contrast, preclinical drusen deposits in eyes with many drusen were seen as accumulations of amorphous material which appeared hyalinised by light microscopy. A distinct feature were rows of dense hyalinised microdrusen (1–2 µm in diameter), over which larger globular hyalinised drusen formed.

Conclusion—Histological and ultrastructural examination can recognise and distinguish the earliest drusen formed as a result of normal aging from those associated with ARM. In eyes without diffuse deposits, histologically all drusen were of the hard hyalinised variety or their derivatives; no soft drusen composed of membranous debris were found. These findings support and explain those of other authors; no soft drusen composed of membranous debris were found. These findings support and explain those of other authors who do not consider the presence of a few small hard drusen to be a risk factor for the development of ARM.

Materials and methods

A bank of 616 eyes (364 patients) had been collected from 1967 to 1994. All the patients had been examined clinically by one of us (SHS) at 1–3 yearly intervals, the last examination being 2 weeks to 60 months before death (mean 13.9 months). Examination included best corrected visual acuity and direct funduscopy. Fundus photography and, from 1976, fluorescein angiography was carried out where possible using the Zeiss 30° fundus camera. Only eyes considered on clinical examination to be normal or with age related macular changes were included; excluded were eyes in which the fundus could not be visualised and those with other retinal pathologies including pathological myopia, retinal vascular diseases, moderate to severe diabetic retinopathy (some patients with mild background retinopathy were included), and end stage glaucoma.
All eyes in the bank were examined by light microscopy (LM) and over 100 were additionally examined by transmission electron microscopy (EM) by standard techniques. Serial sections 8 µm thick were cut horizontally through the macula to include the foveola, optic nerve, and cornea, so that the type, size, and number of drusen outside the macula could also be recorded. Every tenth section was stained by the picro-Mallory method with an average of 15 (range 8 to over 40) sections examined per eye. Histopathological findings on some of these eyes have been previously presented. The eyes were divided into six groups depending on the appearance of the BLD, as this reflects the stage of degeneration of the retinal pigment epithelium (RPE). Briefly, eyes with no BLD (group 1) or those with only patchy BLD (group 2) were considered normal. ARM was diagnosed when the BLD formed a continuous (diffuse) layer beneath the RPE (groups 3 and 4). The end stages of age related macular degeneration, geographic atrophy, and CNV were classed as groups 5 and 6 respectively.

This paper presents the results of the subgroup of 353 eyes of 202 patients defined as normal with no BLD (group 1) or normal aged with only patchy BLD (group 2). Forty five of these eyes were examined by EM.

Bruch’s membrane was defined as a trilaminar structure consisting of the inner and outer collagenous zones separated by the elastic layer and excluding the basement membranes of the RPE and the choriocapillaris. Light microscopic definitions of BLD, and hard and soft drusen were used, based on the picro-Mallory method and periodic acid Schiff (PAS) staining, in conjunction with the international classification for ARM. BLD was defined on LM as the material found between the plasma and basement membrane of the RPE, staining blue on the picro-Mallory method. Hard drusen (nodular drusen) have a globular hyalinised appearance, are strongly PAS positive, and stain red with the picro-Mallory method. Soft drusen (accumulation of the basal linear deposit) are faintly PAS positive and stain a pale blue with a finely granular or ground glass appearance on the picro-Mallory method. They have sloping edges and, on semithin sections, may appear optically empty. They only occur in the presence of a continuous layer of BLD at the macula. Clinically, the following drusen were defined: small hard drusen were <63 µm in diameter with sharply defined borders and early bright hyperfluorescence on fluorescein angiography; soft distinct drusen were >63 µm with solid nodular appearance and distinct edges; soft indistinct drusen were >63 µm with blurred margins, and only faint late hyperfluorescence on fluorescein angiography.

Drusen size was measured on fundus photography using the Wisconsin grid and circle templates designed for use with Zeiss 30° films. On LM size was estimated using a measuring gaticule in the ×25 and ×40 eyepiece objective. The width of one RPE cell (approximately 12–14 µm) was found to be a useful reference. On EM size was calculated from photographs using the known magnification.

Results
There were 139 eyes in group 1 and 214 in group 2 (see Table 1).

### Table 1  General features of study eyes

<table>
<thead>
<tr>
<th>Group 1</th>
<th>Group 2</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>No BLD</td>
<td>Patchy BLD</td>
</tr>
<tr>
<td>Number of eyes (patients)</td>
<td>139 (80)</td>
<td>214 (122)</td>
</tr>
<tr>
<td>Females (%)</td>
<td>8 (10%)</td>
<td>9 (7%)</td>
</tr>
<tr>
<td>Average age (years)</td>
<td>64.7</td>
<td>73.8</td>
</tr>
<tr>
<td>Age range (years)</td>
<td>43–83</td>
<td>60–96</td>
</tr>
<tr>
<td>Examined EM (number of eyes)</td>
<td>18</td>
<td>27</td>
</tr>
</tbody>
</table>

BLD = basal laminar deposit; EM = electron microscopy.

Figure 1  Basal lamina deposit over drusen of mixed composition. (A) Earliest BLD from a group 2 eye consisting of irregular nodules of fibrillar material (arrow) continuous with the RPE basement membrane; there is no membranous debris. (B) In comparison, later BLD seen here in an eye from group 4 consists of fibrillar, amorphous, and long spacing collagen material with a palisading or banded appearance. Membranous debris (arrow) is interspersed between the strands of BLD and within the drusen. (A) A 56 year old man with normal fudinal appearance, vision 6/6. (B) A 68 year old man with soft indistinct (membranous) drusen and a few small hard drusen, vision 6/6. (A) ×21 600 (B) ×11 400.
staining with the picro-Mallory method, was seen in both groups. In the younger eyes (from 40 years old) this appeared focally, becoming continuous throughout the posterior pole in older eyes. In group 2 eyes the Bruch's membrane changes were more advanced with hyalinisation which reached the base of chorioidal capillaries, the capillaries becoming widely spaced between hyalinised pillars of Bruch's membrane. Patchy basophilia corresponding to calcification was seen in eyes older than 65 years.

Electron microscopy showed that this thickening and hyalinisation of Bruch's membrane was due to the accumulation, predominantly in the outer collagenous zone, of coated membrane bound bodies (CMBB), and of long spacing collagen. These bodies were either intact or ruptured, releasing fragments of degraded membrane, 70 nm coated vesicle-like bodies, dense granular material, and electron lucent droplets. There was an accompanying increase in native collagen within both the inner and outer collagenous zones.

**BASAL LAMINAR DEPOSIT (BLD)**

Group 1 (normal) and group 2 (aging) eyes were distinguished histologically by the presence of BLD. Group 1 eyes had no BLD and the RPE was regular. Group 2 eyes had small patches of BLD and the RPE showed an increased amount of lipofuscin. The BLD was located primarily over the apex of small hyalinised drusen (see Figs 8C and 9), best seen on EM, and also over thickened and hyalinised segments of Bruch's membrane. On LM the BLD stained blue with the picro-Mallory method. On EM the earliest BLD was seen as irregular nodules continuous with the original RPE basement membrane and composed of the predominantly fibrillar form of BLD (Fig 1A) as described by Loeffer and Lee. At a late stage, BLD is composed of an accumulation of fibrillar, amorphous, and components resembling long spacing collagen (Fig 1B).

A few sparse coiled membranes were also seen in 23 of the 27 group 2 eyes examined by EM, representing the earliest pathological sign of membranous debris. The membranes were only seen in areas where BLD was present and first appeared admixed within the BLD (Fig 1B). These coiled membranes had the bilayered structure of lipids but were much smaller and lacked the membrane coating found on the CMBB of Bruch's membrane described above (see Table 2). They could also be distinguished from the electron lucent droplets which have no limiting membrane, and were found in most eyes aged over 55 years, predominantly beneath the RPE basement but also percolating into the collagenous zones and within the CMBB (Fig 2).

**CHOROID**

The chorioidal capillaries occupied approximately 70–80% of a given length beneath the fovea in group 1 eyes with an occasional wide spacing between adjacent chorioidal capillaries being occupied by fibrous tissue. In group 2 eyes, capillaries still occupied around 70% of a given length but were occasionally more widely spaced between hyalinised pillars of Bruch's membrane. A possible macrophage was seen in only two eyes, one beneath and one within a regressing small hard drusen.

**DRUSEN**

Drusen were detected histopathologically on the sections examined in 177 eyes (50%) but were seen clinically in only 34% of these. Only those drusen deposits larger than 30 µm in size, around the diameter of two RPE cells, were detectable on funduscopy, occasionally 25 µm on fluorescein angiography; deposits smaller than this are therefore described as preclinical drusen. The drusen detected clinically were all of the small hard variety and in 90% of eyes were few in number (less than 10). Six eyes, all from patients aged over 70 years, had in addition a few soft distinct drusen. No eye had soft indistinct drusen. Nine eyes of five patients had

<table>
<thead>
<tr>
<th>Deposit</th>
<th>Site</th>
<th>Ultrastructural features on EM</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bruch's membrane change</td>
<td>predominantly OCZ</td>
<td>coated membrane bound bodies intact or ruptured as above</td>
</tr>
<tr>
<td>Entrapment sites</td>
<td>rest on ICZ</td>
<td>coated membrane bound bodies</td>
</tr>
<tr>
<td></td>
<td>raise RPE bm</td>
<td>coated membrane bound bodies</td>
</tr>
<tr>
<td>Membranous debris</td>
<td>three sites:</td>
<td>coated membrane bound bodies</td>
</tr>
<tr>
<td></td>
<td>(1) internal to RPE bm</td>
<td>coated membrane bound bodies</td>
</tr>
<tr>
<td></td>
<td>(2) within RPE bm</td>
<td>coated membrane bound bodies</td>
</tr>
<tr>
<td></td>
<td>(3) external to RPE bm</td>
<td>bilayered structure of lipids</td>
</tr>
<tr>
<td></td>
<td>only in presence of BLD</td>
<td>not coated</td>
</tr>
</tbody>
</table>

Table 2 Comparison of material seen in Bruch's membrane, entrapment sites, and membranous debris

EM = transmission electron microscopy; OCZ = outer collagenous zone of Bruch's membrane; ICZ = inner collagenous zone of Bruch's membrane; RPE bm = basement membrane of the retinal pigment epithelium; BLD = basal laminar deposit.
large (>20) numbers of drusen including two eyes from one patient, aged 60, in whom the whole posterior pole of both eyes was studded with masses of small hard drusen. Although drusen in all stages of evolution were seen in eyes with low and moderate numbers of drusen, the sequence of events in the development of drusen was most clearly established from the nine eyes with significant numbers of drusen. The drusen in groups 1 and 2 were similar.

Table 3  Clinicopathological correlation of drusen deposits in 353 eyes with normal aging

<table>
<thead>
<tr>
<th>Histopathology of drusen deposits</th>
<th>Group 1</th>
<th>Group 2</th>
<th>Clinical appearance</th>
</tr>
</thead>
<tbody>
<tr>
<td>None — 80 (58%)</td>
<td>96 (45%)</td>
<td>normal, no drusen</td>
<td></td>
</tr>
<tr>
<td>&lt;30 µm only (subclinical)</td>
<td>29</td>
<td>53</td>
<td>normal, no drusen</td>
</tr>
<tr>
<td>1/2–1 RPE cell high</td>
<td>5</td>
<td>28</td>
<td>normal, no drusen</td>
</tr>
<tr>
<td>At least one 30–63 µm</td>
<td>15</td>
<td>24</td>
<td>occasional small hard drusen</td>
</tr>
<tr>
<td>few 1–2 per section moderate &lt;10 per section</td>
<td>5</td>
<td>—</td>
<td>scattered small hard drusen</td>
</tr>
<tr>
<td>At least one &gt;63 µm</td>
<td>3</td>
<td>6</td>
<td>occasional large hard drusen or drusen cluster</td>
</tr>
<tr>
<td>few 1–2 per section moderate &lt;10 per section</td>
<td>2</td>
<td>5</td>
<td>scattered small hard drusen</td>
</tr>
<tr>
<td>significant &gt;10 per section</td>
<td>2</td>
<td>—</td>
<td>occasional large hard drusen (hard cluster derived drusen)</td>
</tr>
<tr>
<td>Total with drusen deposits</td>
<td>59 (42%)</td>
<td>118 (55%)</td>
<td>masses of small hard drusen +hard or soft distinct drusen</td>
</tr>
</tbody>
</table>

*Grading: the number of drusen deposits seen on light microscopic serial sections was graded as few (0–2 per section), moderate (<10 per section), or significant (>10 per section).

Table 4  Histopathological and ultrastructural features of preclinical drusen deposits

<table>
<thead>
<tr>
<th>Deposit type</th>
<th>LM appearance</th>
<th>EM appearance</th>
<th>Size</th>
<th>Clinical correlation</th>
</tr>
</thead>
<tbody>
<tr>
<td>Entrapment sites</td>
<td>smallest not detectable, hyalinised</td>
<td>smallest only seen on EM, intact and ruptured CMBB</td>
<td>up to ½ RPE cell high</td>
<td>not found in eyes with many drusen</td>
</tr>
<tr>
<td>Amorphous</td>
<td>all hyalinised stain red on P-M</td>
<td>all amorphous electron dense</td>
<td>up to 10 µm</td>
<td>may be found in all eyes</td>
</tr>
<tr>
<td>(i) iridescent plaques</td>
<td>all amorphous electron dense</td>
<td>all amorphous electron dense</td>
<td>all sizes may reach clinical size &gt;30 µm</td>
<td>found in eyes with many drusen</td>
</tr>
<tr>
<td>(ii) globular drusen root-like extensions</td>
<td>all amorphous electron dense</td>
<td>all amorphous electron dense</td>
<td>rows up to 10 RPE cells in length</td>
<td>not found in eyes with many drusen</td>
</tr>
<tr>
<td>(iii) microdrusen over which small globular drusen may develop</td>
<td>all amorphous electron dense</td>
<td>all amorphous electron dense</td>
<td>maximum: 30 µm diameter up to 1 RPE cell high</td>
<td></td>
</tr>
</tbody>
</table>

Mixed | hyalinised heterogeneous stain on P-M | entrapment site material with amorphous material | not found in eyes with many drusen |

LM = light microscopy; EM = transmission electron microscopy; CMBB = coated membrane bound bodies; RPE = retinal pigment epithelium; P-M = Picro-Mallory stain.
are thus described together. The clinical correlation of the various histopathological findings are given in Tables 3 and 4.

PRECLINICAL DRUSEN FORMATION (<25 µm DIAMETER)

On light microscopy, the earliest evidence of drusen formation took several forms: either

tiny plaques in Bruch’s membrane, small hyalinised globular deposits, or microdrusen occurring singly or in a row.

The first drusen related change to appear on LM were plaques or focal patches of thickened hyalinised Bruch’s membrane around one RPE cell in diameter. These stained red with the picro-Mallory method, and had an iridescent appearance, noted best on racking the microscope up and down, which allowed them to be distinguished from the normal patchy hyalinisation which is seen from the fifth decade.

Drusen were first seen as tiny deposits on the inner surface of an iridescent plaque (Fig 3) Globular in shape, usually single, sometimes multiple, and often beneath a single RPE cell, they had a dense hyalinised appearance on LM staining either red, blue, or both with the picro-Mallory method. In many cases, the hyalinisation extended into the outer collagenous zone, indenting a choroidal capillary or extending down an intercapillary pillar, giving the appearance of “roots” beneath the drusen (Fig 3B). A range of sizes was seen. When small, <1 RPE cell in diameter (6–14 µm) and up to half the height of an RPE cell (6 µm), they displaced the base of the overlying RPE cell but were too small to elevate the

Figure 4 Preclinical small globular drusen and microdrusen. Microdrusen each around 1–2 µm in size forming a row (between arrows) of around 125–150 µm. Open arrow marks a cluster of two small preclinical drusen. A 74 year old man with significant numbers of small hard drusen and drusen clusters, vision 6/6. A 0.5 µm plastic section methylene blue, ×420.

Figure 5 Preclinical drusen composed of entrapment sites, (A) and (B) only detectable on EM. (A) Intact (left) and ruptured (right) coated membrane bound bodies (CMBB) situated between the inner collagenous zone (icz) and basement membrane of the RPE (arrow). The ruptured CMBB have released their contents into the underlying collagenous zones. (B) Amorphous material that appears to be derived from the rupture or breakdown of CMBB. Arrow points to fragments of CMBB. (C) Entrapment site composed predominantly of vesicular material which is also present in the collagenous zones. Arrow points to a patch of BLD over the drusen. (D) Entrapment site composed of ruptured CMBB. Note the electron dense amorphous material in the outer collagenous zone giving the appearance of a root-like extension beneath the drusen similar to Figure 1 (B). (A), (B), (C) A 60 year old man with normal fundus appearance, vision 6/6. (D) A 43 year old man with normal fundus appearance, vision 6/6. (A) ×8880, (B) ×9700, (C) ×8000, (D) ×6250.
apical cell border (Fig 3A left). However, the RPE cells overlying larger drusen of height equal to or more than one cell were displaced and attenuated (Fig 3B and 3C). They gradually enlarge, eventually reaching the size of a clinical drusen (>30 µm).

A distinct finding were small (1–2 µm diameter) rounded elevations of a dense hyalinised appearance staining red with the picro-Mallory method which were termed microdrusen (Figs 4 and 9). In eyes with many drusen they were a universal feature, commonly forming rows up to 10 RPE cells in length, and in places forming a continuous layer. On EM the microdrusen were composed of extremely dense amorphous material. Although more electron dense than the surrounding Bruch’s membrane, the structure of the underlying collagenous zones was still evident. Small globular drusen showed a tendency to cluster and frequently these clusters formed over a row of microdrusen which appeared to pass beneath them.

While on LM all these early deposits had a hyalinised appearance, they could be distinguished on EM into two forms. Some were composed on EM of amorphous electron dense material similar to the microdrusen. But another form, which we have termed entrapment sites was composed of intact and ruptured (Fig 5A right and left) CMBB similar to those that accumulate within the outer collagenous zone of the aging Bruch’s membrane.14 15 These CMBB contained coated vesicle-like bodies, dense granules (Fig 5A), amorphous material (Fig 5B), and electron lucent droplets (Fig 5C). In some entrapment sites, the CMBB had ruptured, appearing to release their vesicular contents into the underlying collagenous zones (Fig 5A, 5C). Rarely, there was some association with electron dense plaques or root-like extensions (Fig 5D), as seen more commonly beneath small hyalinised drusen (Fig 3B). Entrapment sites were seen in eyes with only occasional drusen: they were not a feature of the six eyes (three patients) with large numbers of drusen which were examined by EM: only in one of these eyes was a single preclinical drusen with entrapment sites found, all other preclinical drusen deposits detected were of the amorphous type. Entrapment sites appear “trapped” between an elevated RPE basement membrane and collagen fibrils of the inner collagenous zone (Fig 5). Although the smallest entrapment sites (Fig 5A and B) were only detectable on EM, larger ones (Fig 6) were also seen on LM appearing as small hyalinised deposits. Pure entrapment sites were less than half the height of an RPE cell and were subclinical. Some larger drusen deposits, generally up to one RPE cell in height and 30 µm in diameter with the largest seen being a three drusen cluster of 36 µm height and 325 µm diameter, were of mixed composition on EM (Fig 7), and appear to be formed by the addition of amorphous material and entrapment site material which lie in different regions within the drusen. These mixed drusen stained in a heterogeneous fashion with the picro-Mallory method, the denser amorphous material staining red and the more dispersed entrapment material staining blue (Fig 3C). Early preclinical drusen deposits seen on LM were therefore distinguishable on EM into either those consisting of amorphous material only, entrapment sites, or entrapment sites enlarged by amorphous material.

**CLINICAL DRUSEN (>25 µM IN SIZE)**

Drusen larger than two RPE cells in diameter or 25–30 µm were clinically identifiable (Fig 8). Only a few clinical drusen in each eye were composed of mixed entrapment site material and amorphous material. On histopathology, most clinical drusen had a hyalinised appearance and by EM, an amorphous electron dense structure (Fig 9). They showed a tendency to cluster and there was a continuous gradation of size, the majority being 25–63 µm in size, suggesting that they were being formed continuously. They were seen clinically as small hard drusen, yellowish with sharp borders, and with
bright early hyperfluorescence on fluorescein angiography.

Drusen appeared to enlarge by incorporation of several neighbouring deposits. Most drusen greater than 63 µm in size had a lobulated appearance on LM suggesting that they were formed by enlargement and fusion of a cluster of drusen. Between single or clusters of clinical drusen were smaller subclinical drusen formed in relation to rows of microdrusen. The largest hyalinised drusen seen was 250 µm diameter.

On LM the hyalinised contents of most drusen over 63 µm became paler staining less intensely, beginning from the base (Fig 10A). On EM, the drusen showed a greater or lesser degree of dispersion of the contents leaving a rim of apparently firmer amorphous material at the apex (Fig 10B). Once this rim was lost, the drusen lose their hard margins and spread out. Dispersal of the amorphous contents appeared to have resulted from a process resembling flocculation, the amorphous material condensing into particles surrounded by electron lucent spaces (Fig 10B).

**FATE OF CLINICAL DRUSEN**

Degenerative changes were seen in drusen over >25 µm in diameter in the form of outpouchings from the smooth globular surface of the drusen (Fig 11A). The incorporation of RPE pigment granules was seen in drusen from three eyes (Fig 11A and B). EM showed that this was due to an attenuated and degenerate overlying RPE, the contents of which were herniating through a break in the RPE basement membrane into the drusen (Fig 11C). Many drusen in group 2 eyes had a thin layer of early form BLD over the surface on EM.

Regression of drusen occurred in two ways. In some drusen, the dispersal process continued with extreme loss of material which gave rise to a moth eaten or washed out appearance. More commonly, and in older patients, the drusen contents became coarsely granular (Fig 12A): the youngest patient in whom a granular drusen was detected was 68 years old. As the drusen contents broke down, the basement membrane of the overlying RPE which had been stretched over the globular deposits, became infolded and collapsed onto Bruch’s membrane (Figs 12B, C).

Drusen regression was accompanied by degenerative changes in the RPE with cell fallout and attenuation alternating with enlargement and hyperpigmentation of remaining cells (Fig 12A). Macrophage invasion was seen in one regressing drusen.

**Discussion**

In our study, we identified two methods of drusen formation. In the first, the initial event appeared to be the entrapment of coated membrane bound bodies between the basement
membrane of the RPE and the collagen fibrils of the inner collagenous zone. These entrapment sites, identified on EM, may enlarge by the addition of amorphous material to form clinically visible small hard drusen. Although of similar composition to the CMBB that accumulate in the inner and outer collagenous zones in normal aging, the entrapment does not appear to be due to excessive deposition since Bruch’s membrane was only partially filled with age related deposits. Rather, it is likely that the CMBB, thought to be released from the basal plasma membrane of the RPE, are unable to enter the ICZ owing to the tight meshwork of collagen fibrils beneath them. We therefore prefer to call them entrapment sites rather than shedding sites, a term used by Feeney-Burns et al. We found no evidence that these deposits were formed by the process of evagination and pinching off of cytoplasm from the base of the RPE and in no section did we observe this material passing through the basement membrane of the RPE.

The membrane bound material found in entrapment sites should be distinguished from the membranous debris that accumulates on both surfaces of the basement membrane of the RPE in eyes with ARM; as basal mounds on the internal surface; and as a continuous layer on the external surface, called basal linear deposit by Green and Enger, which is the electron microscopic diagnostic hallmark of ARM (Table 2)—a continuous layer of BLD being the light microscopic hallmark of ARM. In contrast with the material found in entrapment sites, these coiled membranes are
smaller and have the bilayered structure of lipids and are not coated. Although the largest drusen composed only of entrapment site material was subclinical (less than the height of one RPE cell), nevertheless they can enlarge to clinical size (>25–30 µm) through the addition of amorphous material. Only small numbers of these mixed drusen of clinical size were present in an eye, with even fewer detected in eyes with significant numbers of drusen. Entrapment sites may be seen in eyes of all ages and have also been detected in primate eyes, and are probably therefore part of normal aging metabolism rather than a pathogenic process.

In the second method of drusen formation, the earliest deposits were plaques and microdrusen of the hyalinised amorphous type resting as focal excrescences on the inner surface of a thickened Bruch’s membrane, often with root-like extensions into both collagenous zones. Larger globular drusen develop over these changes. Entrapment site material was never found in the microdrusen nor in the overlying globular drusen suggesting that a different and pathological process is involved in this method of drusen formation. Larger drusen, up to 250 µm, appeared to be formed by the fusion of clusters of smaller deposits. In eyes with many drusen, the sequence of events appears to be as follows: (1) a row of microdrusen, composed of dense amorphous material, forms on the inner collagenous zone; (2) a overlying cluster of preclinical drusen develops; (3) one or more of these preclinical drusen enlarge to form the clusters seen clinically. A single drusen is formed by the incorporation of neighbouring smaller drusen; (4) as they enlarge the drusen contents disperse on LM, giving a flocculent appearance on EM. The dispersal commences at the base leaving a rim of denser amorphous material at the apex. This may be responsible for the finding by Mullins et al. that only the cores of some hard drusen contain specific lectin binding glycoconjugates; (5) when the RPE over the drusen degenerates, the drusen regresses, the contents become coarsely granular on LM and the drusen may collapse. This sequence was best detected by study of the nine eyes with significant numbers of drusen in which, importantly, no entrapment sites were seen. In comparison with entrapment sites, therefore, we believe that these second type of early drusen represents precursors to pathological accumulations.

It is possible that the microdrusen play a pathogenic role in the later development of soft drusen. One soft drusen type, previously termed serogranular drusen, clinically has a yellowish inspissated appearance, and fluoresces only late or not at all on fluorescein angiography. They commonly calcify as they regress and are often seen surrounding areas of geographic atrophy. On histopathology they have a granular structure with a row of microdrusen at their base. The presence of microdrusen and continuity with hyalinised drusen suggest that they may result from broken down clusters of hard drusen. The presence of microdrusen may also account for the marked increase in drusen numbers that can develop over time in previously normal appearing retina between existing drusen. Another postulated role for microdrusen may be in the formation of large confluent soft drusen or drusenoid detachments when occurring on a background of small hard drusen. One assumed pathogenic mechanism for these soft drusen is that they are formed by the addition of serous fluid which accumulates when fluid egress by the RPE pumping mechanism is delayed across Bruch’s membrane rendered hydrophobic by an increase of lipids. In older eyes with ARM, this barrier might be reinforced by the lipid membranous debris that
accompanies the BLD. In eyes without BLD, it is possible that this hydrophobic barrier is formed by the rows of dense microdrusen.

It is stressed that this is not a quantitative paper addressing the incidence of drusen. The incidence of drusen detected clinically in our series is lower than that noted by recent epidemiological studies. This can be explained by differences in examination methods. These studies were based on drusen detection using 30° stereoscopic photographs which can be expected to detect features two or three times as often as funduscopy. Fundus photography could not be performed in all eyes in our series however, owing both to the early date of many examinations (eyes were collected from 1967) and to the medical problems, most notably dementia and stroke, in many patients.

An important observation is that all the drusen detected in this group of 353 eyes without the diffuse deposits that mark ARM were of the small hard hyalinised variety or of their derivatives and that no soft indistinct drusen were seen. Our finding that only drusen larger than 30 μm were detectable on funduscopy or fluorescein angiography confirms clinical definitions of minimum apparent drusen. Our study also clarifies the nature of the drusen in patients with massed drusen, the so called starry sky appearance. They have been called basal laminar drusen and attributed to a nodular thickening of the RPE basement membrane. The two eyes in our series with this appearance had large numbers of hard hyalinised drusen.

Lipoidal degeneration of the RPE was found in only one eye with early BLD and preclinical hyalinised drusen. The changes found in Bruch’s membrane, including electron lucent droplets and the development of BLD, confirm the findings of other workers.

We observed two differing patterns of drusen regression and disappearance. In younger eyes, the hyalinised drusen contents gradually faded, giving the drusen a moth eaten appearance. In contrast, the hyalinised amorphous contents in older patients became coarsely granular. Drusen regression was associated with degenerative changes of the overlying RPE cells. Therefore, while the clinical prognostic implications of drusen disappearance are unknown, at least on the histopathological level drusen disappearance does not indicate a return to normality. Outpouchings were observed projecting from the surface of hyalinised or degenerating drusen which were confirmed on serial sections to be continuous with the parent drusen. This would account for the so called intracellular RPE drusen noted by previous observers.

Conclusion

In eyes without diffuse deposits of BLD or membranous debris, all drusen histologically are of the hyalinised variety or of their derivatives. Ultrastructural examination would appear to identify two methods of drusen formation. The first are entrapment sites which may enlarge by the addition of amorphous material, and are seen mainly in eyes with only a few drusen. By contrast, in eyes with many drusen, early drusen form through a pathologica laccumulation of amorphous material only.

This finding of two methods of early drusen formation helps to clarify the question of the clinical significance of small hard drusen. The identification of entrapment sites, probably a normal and universal aging process, supports and helps to explain the findings of other authors who, from epidemiological and histopathological studies, consider the presence of a few small hard drusen to be normal and not to be a risk factor for the development of ARM. In contrast, in eyes with large numbers of small hard drusen, it is likely that the drusen represent pathological accumulations. These patients with masses of hard drusen would be poorly classified as ARM, although both Bressler et al and Klein et al found that eyes with large areas of small hard drusen have an increased risk of developing soft indistinct drusen, a clinical marker of early ARM. Since these eyes, especially with the onset of pigment changes, are at risk of complications such as geographic atrophy and, more rarely, choroidal neovascularisation they could be considered to have a hard drusen maculopathy.

8 Sarks SH. Changes in the region of the choriocapillaris in ageing and degeneration. 23rd Conc Ophthalmol, Kyoto 1979:228–38.