Autologous limbal transplantation in patients with unilateral corneal stem cell deficiency

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Abstract

Aim—To describe a surgical technique for autologous limbal stem cell transplantation and the outcome of a series of patients with unilateral stem cell deficiency.

Methods—A report of six consecutive patients who underwent autologous limbal stem cell transplantation is presented. The primary diagnosis included alkali burn (n=3), conjunctival intraepithelial neoplasia (CIN) (n=1), recurrent pterygium (n=1), and contact lens induced keratopathy (n=1). The autologous transplanted tissue consisted of peripheral cornea, limbus, and conjunctiva obtained from the contra-lateral eye. Three of the above patients underwent penetrating keratoplasty in association with autolimbal transplantation. A significant modification to established techniques was the close monitoring of conjunctival epithelial migration in the immediate postoperative period. If conjunctival epithelium threatened to migrate on to the corneal surface, it was mechanically removed at the slit lamp and prevented from crossing the limbus. This was required in three patients.

Results—The mean follow up was 18.8 months. The outcome was satisfactory in all cases: a stable corneal surface was restored and there was a substantial improvement in vision and symptoms. One patient had a primary failure of the corneal allograft associated with glaucoma, and 6 months later developed a retinal detachment. No complications were noted in the donor eye with the exception of one patient who developed filamentary keratitis along the edge of the donor site.

Conclusion—Autologous limbal transplantation with corneal, limbal, and conjunctival carriers was found to be useful for ocular surface reconstruction, over a mid-term follow up, in patients with unilateral stem cell deficiency. Close monitoring of the migration of conjunctival epithelium in the immediate postoperative period, and preventing it from crossing the limbus, ensured that the corneal surface was re-epithelialised exclusively from epithelial cells derived from the transplanted limbal tissue. This approach should improve the success of this procedure.

Clinical and laboratory evidence indicates that corneal stem cells are located at the limbus. 1–3 Limbal deficiency, or loss of corneal stem cells, is associated with conjunctivalisation of the corneal surface, recurrent and persistent epithelial defects, chronic inflammation, scarring, and ulceration of the cornea. The management of these ocular surface disorders continues to pose a challenge. Early methods of reconstruction of the ocular surface included conjunctival transplantation and keratoepithelioplasty. Conjunctival transplantation was based on the theory of transdifferentiation of conjunctival epithelium into cornea-like epithelium. 4–8 However, we now know that conjunctival epithelial transdifferentiation (that is, a morphological, biochemical, and physiological transformation of conjunctival epithelium into corneal epithelium) does not occur, 9–10 and conjunctival transplantation to reconstruct the corneal surface in patients with corneal stem cell deficiency has been abandoned. Keratoepithelioplasty was proposed by Thoft 11 as another alternative to reconstruct the ocular surface in patients with corneal stem cell deficiency. In this technique lenticules of peripheral corneal epithelium with superficial stroma were grafted. Subsequently, the same author modified the technique to include limbal tissue, acknowledging the importance of stem cell transplantation in these conditions, for a successful long term outcome. The use of cultured limbal epithelium is, at present, still under investigation. 12 Limbal transplantation, as proposed by Kenyon and Tseng, 13 is probably the best current option for ocular surface reconstruction in patients with total corneal stem cell deficiency.

This report describes a modified surgical technique for autologous limbal transplantation and the outcome in a series of patients with unilateral corneal stem cell deficiency.

Methods

Six consecutive patients with unilateral corneal stem cell deficiency who underwent autologous limbal transplantation were included. Demographics and clinical characteristics are summarised in Table 1. Five patients had long standing symptoms and signs characteristic of chronic and diffuse corneal stem cell deficiency including decreased vision, pain, photophobia, discomfort, corneal vascularisation and scarring, chronic redness, and recurrent episodes of epithelial defects. One patient (case 6) had a recurrent pterygium with restriction of ocular motility. The pterygium had been excised four times over a period of 6 years. A conjunctival autotransplant was used to cover
the sclera at the time of the fourth excision, performed a year before presentation. Despite this the pterygium had recurred and encroached on the visual axis causing reduced vision and diplopia.

The surgical technique was similar in all cases. The donor tissue consisted of corneal-limbal-conjunctival explant(s) that were harvested from the contralateral normal eye, except in case 6, where it was harvested from the superior limbus of the same eye. In cases 1–4, two explants, corresponding to 2 clock hours (11–1 o'clock and 5–7 o'clock) and consisting of 2 mm of peripheral cornea, limbus, and 3 mm of bulbar conjunctiva, were harvested. In case 5, only one explant from the superior limbus of the contralateral eye and in case 6 only one explant, corresponding in circumferential length to 3 clock hours, were harvested. In case 6, the pterygium was excised from the limbus.

The recipient eye was prepared by incising the conjunctiva, at the limbus, adjacent to the area with the most severe corneal surface abnormalities, or at the superior and inferior limbus. The abnormal corneal epithelium and the superficial fibrovascular scar tissue were stripped off by blunt dissection. The calibrated diamond knife was set to 100 µm (an allowance of 50 µm was made for the lack of epithelium) and a bed, corresponding in dimensions to the donor explants, was fashioned at the recipient site(s). The donor tissue was then sutured onto the recipient eye with two interrupted 10-0 nylon sutures at the corneal margin and two along the scleral edge of the explant (Fig 1a). The conjunctiva of the recipient eye was then approximated to the donor conjunctiva with interrupted 8-0 Vicryl sutures, taking a bite into episclera. When a penetrating keratoplasty was also required, this was performed after the limbal explants were first sutured into place (Fig 1b).

Two patients (cases 3 and 5) underwent penetrating keratoplasty (PK) at the time of limbal transplantation, and case 4 underwent PK 4 months after the original procedure. In case 6, mitomycin C (0.04%, 1 minute) was applied intraoperatively with a Weck-cel sponge to the subconjunctival tissue after excision of the recurrent pterygium and fibrous tissue. This was irrigated thoroughly before suturing the limbal explant.

Postoperative treatment consisted of preservative-free topical chloramphenicol and prednisolone 0.5% eye drops, four times daily for the first 2 weeks. Steroids were then tapered rapidly. Autologous serum drops were used in the immediate postoperative period, as previously described. Frequent preservative-free artificial tears were also used.

Table 1  Demographics and clinical data of patients undergoing autologous limbal graft

<table>
<thead>
<tr>
<th>Patient No, eye age (years), sex</th>
<th>Eye</th>
<th>Primary diagnosis</th>
<th>Previous surgeries</th>
<th>VA before ALT</th>
<th>VA associated</th>
<th>Months of follow up</th>
<th>VA after ALT</th>
<th>Complications</th>
<th>Success</th>
</tr>
</thead>
<tbody>
<tr>
<td>1, 39, F roll</td>
<td>R</td>
<td>CIN</td>
<td>Excisional biopsy (alcohol, cryotherapy)</td>
<td>6/36</td>
<td>—</td>
<td>19</td>
<td>6/9</td>
<td>Recurrence of CIN (controlled with MMC)</td>
<td>Yes</td>
</tr>
<tr>
<td>2, 30, M roll</td>
<td>R</td>
<td>Chemical burn</td>
<td>FS, ECCE + AV, AC-IOL, PK (×2), PK</td>
<td>1/60</td>
<td>—</td>
<td>31</td>
<td>6/36</td>
<td>—</td>
<td>Yes</td>
</tr>
<tr>
<td>3, 79, M roll</td>
<td>R</td>
<td>Aphakia, CL wear</td>
<td>HM PK</td>
<td>18</td>
<td>HM*</td>
<td>—</td>
<td>6/36</td>
<td>Glaucoma, PK failure*</td>
<td>No*</td>
</tr>
<tr>
<td>4, 24, M roll</td>
<td>L</td>
<td>Chemical burn</td>
<td>Mucosal graft</td>
<td>16</td>
<td>6/36</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>Yes</td>
</tr>
<tr>
<td>5, 58, F roll</td>
<td>L</td>
<td>Chemical burn</td>
<td>PK</td>
<td>15</td>
<td>6/36</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>Yes</td>
</tr>
<tr>
<td>6, 50, M recurrent pterygium</td>
<td>R</td>
<td>Pterygium excision (×4)</td>
<td>Conjunctival autokeratoplasty graft (×1)</td>
<td>14</td>
<td>6/9</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>Yes</td>
</tr>
</tbody>
</table>

VA = visual acuity; ALT = auto-limbal transplantation; CIN = carcinoma in situ; MMC = mitomycin C; CL = contact lens; FS = filtration surgery; ECCE = extracapsular cataract extraction; AV = anterior vitrectomy; AC-IOL = anterior chamber intraocular lens implantation; PK = penetrating keratoplasty; HM = hand movements.

*VA loss and failure after retinal detachment.

Figure 1  (a) Diagram illustrating the placement of auto-limbal explants at the 6 and 12 o'clock positions of recipient eye. (b) Auto-limbal transplant with corneal graft. The graft is usually 7 mm in size as the peripheral 2 mm, both above and below, is covered by the explants.
All patients were followed daily or twice daily until corneal epithelialisation was complete. If conjunctival epithelium was seen to progress towards the limbus or encroach on to the cornea, it was mechanically debrided under topical anaesthesia, at the slit lamp as previously described, to ensure that corneal re-epithelialisation occurred with corneal epithelial cells derived from the transplanted explants. This was required in three patients (Fig 2a–f).

Results

In this series of patients median follow up time was 18.8 months (Table 1). No intraoperative complications occurred. Postoperatively, the limbal grafts started epithelial outgrowths within the first 2 days (Fig 2a, b) and the whole corneal surface was completely epithelialised within 2 weeks, in all cases. There was no infection, limbal graft failure, or slippage of tissue. The epithelium was stable, without recurrence of epithelial defects, transparent, and smooth. There was no corneal neovascularisation. Improvement of vision and symptoms after surgery was substantial in all cases (see Table 1 and Fig 3a–e). Case 3 had primary failure of the corneal allograft and glaucoma. Four weeks later he underwent repeat PK and diode laser cyclophotocoagulation was applied to the areas not covered by the limbal tissue. The outcome was satisfactory until 6 months later, when a retinal detachment was noted. Although the corneal surface remained healthy during follow up, his visual function deteriorated to hand movements.

In the donor eyes there were no intraoperative complications, refractive changes, chronic inflammation, persistent epithelial defects, or corneal neovascularisation. One of the patients (case 3) developed filamentary keratitis in the donor eye, which was controlled with intense topical lubrication.

Case 1, who had conjunctival intraepithelial neoplasia, had a recurrence of the lesion...
following auto-limbal transplantation. This had extended on to the cornea for approximately 4 mm at the time it was first seen. She was treated with mitomycin C drops and the abnormal epithelium disappeared, leaving the eye symptom free with a visual acuity of 6/9. Topical mitomycin C therapy did not affect the viability of the limbal grafts during the 6 month follow up period after completion of treatment. Histological examination (available for two cases) of the fibrovascular membrane with overlying epithelium, excised from the corneal surface at the time of surgery, showed numerous intraepithelial goblet cells and basal intraepithelial lymphocytes. Preoperative impression cytology was performed for only one patient and this too showed multiple goblet cells. Pathological examination thus provided further evidence of the conjunctival origin of the “corneal epithelium”.

Discussion

A variety of techniques of limbal transplantation have been reported. All these procedures provide a new source of epithelium for a diseased ocular surface and the removal of the host’s altered corneal epithelium and pannus. From the donor tissue, transient amplifying cells are generated which migrate onto the denuded corneal surface of the host. The donor tissue can be obtained from the fellow eye (autograft) in cases of unilateral disease. Cadaveric whole globe or corneoscleral rim or a living relative (allograft) can be used when both eyes are affected. Limbal transplantation procedures also vary depending on the carrier tissue used for the transfer of the limbal stem cells. Carrier tissue is needed in limbal transplantation because it is not possible to transfer limbal stem cells alone. Either conjunctiva (conjunctival limbal graft) or corneal/limbal stroma (keratolimbal graft) have been used as carrier tissue for limbal stem cells.

This study describes the outcome of six patients who underwent a modified surgical technique of autologous limbal transplantation for a variety of ocular surface diseases. Superficial corneal stroma, perilemmal sclera, and conjunctiva were used as carriers for limbal cells and providers of an adequate microenvironment for their survival and replication. All cases achieved rapid surface healing with restoration of a smooth, stable, and optically improved surface, resulting in improved visual acuity. A key factor contributing to the success of this study is the early recognition of conjunctival epithelial migration towards the limbus and the prevention of such migration onto the cornea. To achieve this end it is imperative to monitor closely all patients following transplantation, until corneal re-
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