Vitreous penetration of levofloxacin in the uninflamed phakic human eye

E N Herbert, I A Pearce, J McGalliard, D Wong, C Groenewald

SCIENTIFIC CORRESPONDENCE

Aims: To assess the vitreous penetration of oral levofloxacin (a new fluoroquinolone antibiotic with improved Gram positive activity) in uninflamed phakic eyes.

Methods: 15 patients for macula hole surgery were recruited to the study. 10 received a single 500 mg dose of levofloxacin by mouth preoperatively. Five acted as controls. Serum and undiluted vitreous samples were obtained at surgery and analysed by HPLC.

Results: Levofloxacin was detectable 2.5 hours after administration in the vitreous. A peak concentration of 1.6 µg/ml (or mg/l) was measured between 2.5 and 4 hours post-dose.

Conclusion: Oral levofloxacin reaches the vitreous rapidly in the uninflamed phakic eye. Levels did not reach MIC90 for the commonest infecting organisms. Nevertheless, levofloxacin would be expected to be active against a higher proportion of infecting organisms than either ciprofloxacin or ofloxacin.

The blood-ocular barrier effectively excludes many toxic substances from the eye, but also limits entry of many therapeutic agents including antibiotics. The fluoroquinolone antibiotics have been shown to produce therapeutic levels in both aqueous and vitreous after systemic administration. Levofloxacin has a unique mechanism of action, inhibiting bacterial DNA gyrase, leading to cell death. The concentration required for killing susceptible organisms is little different from that required for inhibition of growth.

Unfortunately, their spectrum of antimicrobial activity does not coincide with the organisms frequently found in endophthalmitis. Following cataract surgery 94% of culture positive infections are caused by Gram positive organisms, including 70% coagulase negative staphylococci. The activity of fluoroquinolones against Gram positive organisms has limited their usefulness in other specialties—for example, in the treatment of pneumonia. This has led to the development of fluoroquinolones with an improved bactericidal spectrum. Levofloxacin (the active S-isomer of ofloxacin) is the first of these to be licensed in the United Kingdom. Its minimum inhibitory concentrations are approximately half those for racemic ofloxacin.

We aimed to evaluate how quickly levofloxacin reached therapeutic levels in the vitreous after oral administration. Previous studies assessing vitreous penetration of antibiotics have included heterogeneous patient groups with a wide variety of underlying conditions that would be expected to severely compromise the blood-ocular barrier—for example, proliferative diabetic retinopathy, trauma, retinal detachment, dropped nucleus. We selected patients having vitrectomy for idiopathic macula hole only, so that penetration was studied in as near as possible the normal uninflamed eye.

METHODS

The study was approved by the Liverpool research and ethics committee and informed consent obtained from all participants. Patients undergoing vitrectomy to treat macula hole were recruited between June and December 2000. Exclusion criteria were any previous eye surgery, diabetic retinopathy, epilepsy, sensitivity to quinolones, renal impairment, pregnancy, and age less than 18 years. Ten patients received a single 500 mg tablet of levofloxacin a variable time before surgery. Five patients acted as controls receiving no levofloxacin. A serum sample was obtained immediately before commencement of surgery, and an undiluted vitreous sample obtained with the ocutome before commencing the infusion as previously described. Specimens were anonymised and stored at -70°C until analysed by high performance liquid chromatography (HPLC). Assays were performed in a masked fashion at the Bristol Centre for Antimicrobial Research and Evaluation. Each sample was tested once. The system was flushed between each test to eliminate the risk of carryover. Positive controls made to a known concentration were used to confirm the accuracy of the assay. Samples were deemed negative at a level of 0.1 µg/ml (or mg/l) or less. Repeat sampling of vitreous from a single patient is not possible, therefore single samples from each patient were combined to construct a model of the pharmacokinetic profile of levofloxacin.

RESULTS

Fifteen patients were recruited. Of the 10 receiving levofloxacin seven were female, three male. Mean age was 65 years (range 46–76) and mean weight was 70 kg (range 57–100 kg). All were phakic. Seven had surgery under local anaesthesia with no preoperative fasting: three had a general anaesthetic with 6 hours’ fasting. Vitreous samples were collected, on average, 20 minutes (range 15–40 minutes) after the serum samples.

Levels of levofloxacin achieved are shown in Figure 1. Levofloxacin was detectable in the serum of all treated patients, the earliest being taken only 55 minutes after administration (range 3.0–6.9 µg/ml). The range of vitreous levels was 0.0–1.6 µg/ml. No levofloxacin was found in the serum or vitreous of control cases. No adverse drug reactions were noted.

DISCUSSION

The fluoroquinolones have high oral availability, approximately 99% for levofloxacin, and are absorbed rapidly. Food does not significantly affect absorption. Maximum serum concentrations are achieved after 1 hour when fasted or 2 hours unfasted. Our results are consistent with this rapid availability in serum. We have found that levofloxacin reaches the vitreous of uninflamed phakic eyes within 2.5 hours of a single oral dose. The peak concentration measured (1.6 µg/ml) was less than half the highest reported MIC90 for Staphylococcus epidermidis, the commonest organism isolated in post-cataract...
than 2 hours after last dose), exceeding most of the MIC\textsubscript{90} values they chose. However, our own review of the literature found higher MIC values and therefore fewer organisms encountered in endophthalmitis as they include isolates obtained from blood cultures in cancer patients, many of whom had received recent antimicrobial therapy. Susceptibility data obtained from ocular isolates would provide the most relevant reference point, but none is presently published. It is likely that MIC values will rise as resistance becomes more widespread. This has been seen for other fluoroquinolones tested on isolates from bacterial keratitis.\textsuperscript{*} Discretion should be used to reduce inappropriate use of these antibiotics and limit spread of resistance.

The role of systemic antibiotics in therapy or as prevention for endophthalmitis has not been established.\textsuperscript{14} Direct intravitreal treatment remains the mainstay of treatment. Systemic agents with adequate vitreous penetration and an appropriate antimicrobial spectrum may have a valuable role. The penetration of levofloxacin and newer fluoroquinolones into the infected eye and their effect on outcome require further investigation.

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endophthalmitis (Table 1). It is well recognised that sensitivities to most antibiotics for *S. epidermidis* are very variable (as seen by the large range in Table 1) and resistance widespread. The MIC\textsubscript{90} values quoted may not be representative of the organisms encountered in endophthalmitis as they include isolates obtained from blood cultures in cancer patients, many of whom had received recent antimicrobial therapy.\textsuperscript{*} Susceptibility data obtained from ocular isolates would provide the most relevant reference point, but none is presently published. It is likely that MIC values will rise as resistance becomes more widespread. This has been seen for other fluoroquinolones tested on isolates from bacterial keratitis.\textsuperscript{13} Discretion should be used to reduce inappropriate use of these antibiotics and limit spread of resistance.

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Table 1 \(\text{MIC}_{90}\) values for organisms commonly isolated in bacterial endophthalmitis. Values in \(\mu g/ml\)

<table>
<thead>
<tr>
<th>Organism</th>
<th>% isolates in EVS\textsuperscript{*}</th>
<th>Highest MIC\textsubscript{90} (µg/ml)</th>
<th>Ref 8, 1999</th>
<th>Ref 9, 1997</th>
<th>Ref 11, 1994</th>
<th>Ref 12, 1998 (MIC\textsubscript{90} range)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Staphylococcus epidermidis</em></td>
<td>70</td>
<td>4.0 (16.0)</td>
<td>4.0</td>
<td>4.0 (16.0)</td>
<td>0.42</td>
<td>0.25–0.41</td>
</tr>
<tr>
<td><em>Staphylococcus aureus</em></td>
<td>9.9</td>
<td>0.5 (16.0)</td>
<td>0.25</td>
<td>0.25 (8.0)</td>
<td>0.41 (0.52)</td>
<td>0.25–0.5 (0.05–16)</td>
</tr>
<tr>
<td><em>Enterococcus faecalis</em></td>
<td>2.2</td>
<td>2.0</td>
<td>2.0</td>
<td>2.0</td>
<td>1.0</td>
<td>0.06–2.0</td>
</tr>
<tr>
<td><em>Bacillus cereus</em>\textsuperscript{†}</td>
<td>1.2</td>
<td>3.1</td>
<td>2.0</td>
<td>2.0</td>
<td>1.91</td>
<td>1.0–3.13</td>
</tr>
<tr>
<td><em>Pseudomonas mirabilis</em></td>
<td>1.9</td>
<td>0.25</td>
<td>0.25</td>
<td>0.25</td>
<td>0.18</td>
<td>0.06–0.25</td>
</tr>
<tr>
<td><em>Haemophilus influenzae</em></td>
<td>0.05</td>
<td>0.05</td>
<td>0.03</td>
<td>0.02</td>
<td>0.015–0.05</td>
<td></td>
</tr>
</tbody>
</table>

\(\text{MIC}_{90}\) is the minimum concentration of antibiotic required to inhibit replication in 90% of clinical isolates in vitro.

Figures in parentheses are for methicillin-resistant organisms.

\*More common in late post trabeculectomy endophthalmitis; †more common in post-traumatic endophthalmitis.

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REFERENCES