Architecture of arachnoid trabeculae, pillars, and septa in the subarachnoid space of the human optic nerve: anatomy and clinical considerations

H E Killer, H R Laeng, J Flammer, P Groscurth

Aims: To describe the anatomy and the arrangement of the arachnoid trabeculae, pillars, and septa in the subarachnoid space of the human optic nerve and to consider their possible clinical relevance for cerebrospinal fluid dynamics and fluid pressure in the subarachnoid space of the human optic nerve.

Methods: Postmortem study with a total of 12 optic nerves harvested from nine subjects without ocular disease. All optic nerves used in this study were obtained no later than 7 hours after death, following qualified consent for necropsy. The study was performed with transmission (TEM) and scanning electron microscopy (SEM).

Results: The subarachnoid space of the human optic nerve contains a variety of trabeculae, septa, and stout pillars that are arranged between the arachnoid and the pia layers of the meninges of the nerve. They display a considerable numeric and structural variability depending on their location within the different portions of the optic nerve. In the bulbar segment (ampulla), adjacent to the globe, a dense and highly ramified meshwork of delicate trabeculae is arranged in a reticular fashion. Between the arachnoid trabeculae, interconnecting velum-like processes are observed. In the mid-orbital segment of the orbital portion, the subarachnoid space is subdivided, and can appear even loosely chambered by broad trabeculae and velum-like septa at some locations. In the intracanalicular segment additionally, few stout pillars and single round trabeculae are observed.

Conclusion: The subarachnoid space of the human optic nerve is not a homogeneous and anatomically empty chamber filled with cerebrospinal fluid, but it contains a complex system of arachnoid trabeculae and septa that divide the subarachnoid space. The trabeculae, septa, and pillars, as well as their arrangement described in this study, may have a role in the cerebrospinal fluid dynamics between the subarachnoid space of the optic nerve and the chiasmal cistern and may contribute to the understanding of the pathophysiology of asymmetric and unilateral papilloedema. All the structures described are of such delicate character that they cannot even be visualised with high resolution magnetic resonance imaging (MRI).

MATERIAL AND METHODS

The 12 optic nerves from nine patients used in this study were removed no longer than 7 hours after death following qualified consent for necropsy. Access to the optic nerve and the globe was obtained through the orbital roof. Before removing the globe, the subarachnoid space of the intracranial segment of the optic nerve was carefully injected with fixative using a 26 gauge needle. Subsequently, the orbital portion of the optic nerve and the globe were carefully dissected in situ from the surrounding tissues and the specimens were fixed with 2.5% glutaraldehyde (0.1 M cacodylate buffer). In two cadavers the optic nerves and globes were removed with the intact sphenoid bone in order to examine the intracanalicular segment of the nerve.

Scanning electron microscopy (SEM)

The globe and optic nerve were fixed for at least 1 week in a solution of 2% glutaraldehyde (0.1 M cacodylate buffer). For examination of the intracanalicular segment the fixed specimens were decalcified with 0.5% HNO₃, for 24 hours.
Transverse sections of the bulbar, mid-orbital, and canalicular segment were then dehydrated in an acetone series, dried by the critical point method (CO₂), mounted on aluminium stubs, and sputtered with gold (15 nm). The specimens were studied with an SEM 505 (Philips, Einthoven, Netherlands).

**Transmission electron microscopy (TEM)**

Small fragments (approximately 1 mm³) were cut from the bulbar and mid-orbital segment and postfixed for 1 day within 1% OsO₄ (0.1 M cacodylate buffer). Subsequently the specimens were dehydrated in a series of alcohol and embedded into Epon by routine procedure. Semithin sections (approximately 2 µm) were cut from each block and stained with toluidine blue to identify the meninges. Ultrathin sections (approximately 50 nm) were contrasted with uranyl acetate and lead citrate and studied with a CM 100 (Philips, Einthoven, Netherlands).

**RESULTS**

**Bulbar segment of the orbital optic nerve portion**

The bulbar segment of the optic nerve showed a distinct enlargement of the subarachnoid space (ampulla) which was detectable even macroscopically. SEM examination revealed abundant round shaped arachnoid trabeculae bridging the subarachnoid space, which were anchored in the arachnoid and pia layers without broadening (Fig 2A). The trabeculae were often branched to form a delicate network (Fig 2B). The profile of the trabeculae varied between 5 µm and 7 µm. Thin lamellae of highly flat cells were located in a veil-like pattern between adjacent trabeculae. Occasionally, larger trabeculae containing one or two blood vessels were found within the trabecular network (Fig 2C).

Each trabecula was covered by flat leptomeningeal cells with smooth surface and slender processes (Fig 2D). The continuity of the cells was occasionally interrupted by intercellular gaps and fenestrations with a diameter between 0.2 µm and 1 µm. TEM revealed that each trabecula was surrounded by a complete sheath of leptomeningeal cells (Fig 3A). Adjacent trabeculae were connected by thin cytoplasmic bridges of leptomeningeal cells running freely through the subarachnoid space. The oval shaped nucleus of the leptomeningeal cells was rich in heterochromatin and protruded
into the subarachnoid space. The leptomeningeal cells formed, with their slender processes, either single or multiple layers of surface lining cells (Fig 3B). In multiple layers the cells were attached to each other by well defined desmosomes. The size of the intercellular space varied distinctly and it was often widened forming small lacunae between leptomeningeal cells. The basal portion of the leptomeningeal cells was supported either by a fine fibrillar extracellular matrix resembling a basal lamina or by delicate collagenous fibrils in close contact with the cell membrane. The centre of the trabeculae was filled by densely packed collagenous fibrils which were arranged in small bundles (Fig 3A). Occasionally, single fibroblasts were detected within the trabeculae in close contact with the collagenous fibrils. Blood capillaries, lymphatic vessels, and nerve fibres were not present.

Mid-orbital segment of the orbital optic nerve portion
The subarachnoid space was distinctly smaller than in the bulbar segment. It contained numerous broad septa running in various directions (Fig 4A and B). They divided the subarachnoid space into chambers that were connected to each other by large perforations within the septae and by interseptal spaces. In addition, single round shaped pillars were detectable which, in contrast with trabeculae, ended with broadened ends at the dural and pial attachment of the arachnoidal layer (Fig 4B). Both septa and pillars were covered by flattened cells with smooth surfaces displaying numerous intercellular fenestrations of varying size (Fig 4D). TEM morphology of leptomeningeal cells covering septa and pillars was comparable to that of trabeculae in the bulbar segment. They formed either single or multiple layers and were connected to each other by well defined desmosomes. The centre of the septa and pillars contained numerous collagenous fibrils and single fibroblasts but no blood vessels, lymphatics, or nerve fibres.

Intracanalicular portion of the optic nerve
Within the optic canal the subarachnoid space was continuous over large distances (Fig 5A). The centre of the canal demonstrated one or two large pillars with a diameter of approximately 25 µm which expanded at the dural and pial attachment of the arachnoidal layer (Fig 5B), or delicate round shaped and slightly curved trabeculae of approximately 5 µm diameter (Fig 5C). At the orbital opening of the canal the trabeculae were more numerous running in parallel and bridging the subarachnoid space in oblique direction (Fig 5C). Again the surface of the pillars and trabeculae was covered by flat, smooth surfaced cells with small intercellular clefts. Occasionally single leucocytes were found closely attached to the leptomeningeal cells (Fig 5D).

DISCUSSION
The present study has used SEM and TEM to describe in detail for the first time both the fine anatomy of trabeculae, septa, and pillars in the subarachnoid space of the human optic nerve and their relation to location within the different portions of the nerve.

The morphology of the cranial and spinal meninges and of the subarachnoid space have been described in previous studies on dogs, cats, rats, and humans. In order to find appropriate terminology for the structures transversing the spinal subarachnoid space, Parkinson used the terms arachnoid septa, trabeculae, and “rough strands.” A classification of cranial arachnoid trabeculae in stout, columnar or sheet-like...
Some of the larger trabeculae contain blood vessels (Fig 2B) as arachnoid trabeculae are arranged in a reticular fashion. Segment of the intraorbital portion. There, a multitude of deli-
subarachnoid space of the optic nerve is widest in the bulbar present study on human optic nerves we observed that the 
anchorage of the fibroblasts that form the arachnoid trabecu-
lae in the bulbar segment and not in the canalicular portion.
Hayreh and Liu suggested that the trabeculae in the bulbar segment behind the globe, cerebrospinal fluid needs to be absorbed enough cerebrospinal fluid during a rise in fluid pressure. The subarachnoid space of the optic nerve is at the lower end of the cerebrospinal fluid chambers and because of its cul de sac anatomy the pressure gradient points
however, provide histological evidence that would support their assumptions. Concerning the architecture and location of the trabeculae and septa, our study shows that the subarachnoid space narrows in the mid-orbital segment where the delicate character of the trabeculae changes gradually into broader septa and stout pillars that subdivide the subarachnoid space into small compartments. Within the intracanalicular portion the subarachnoid space is extremely narrow. Only a few large pillars and trabeculae but no septa are observed. As described by other authors, free cells can be observed on the arachnoid surface and on the surface of the trabeculae. In consideration of this anatomy the subarachnoid space of the optic nerve cannot be regarded as a homogeneous space filled with cerebrospinal fluid, but rather as a multichambered and subdivided tubular system with a blind end (cul de sac) behind the ocular globe.
Cerebrospinal fluid is mainly produced by the choroid plexus epithelium and the ependymal cells of the ventricular system from where it flows into interconnecting chambers—namely, the cisterns and the subarachnoid spaces of the central nervous system. Cerebrospinal fluid circulation and direction of flow have been studied with radioisotopes and other tracers injected into the cerebrospinal fluid. It is generally agreed that there is a bulk circulation of fluid from the site of origin to the site of absorption—that is, from the ventricles to the arachnoid villi in the cranial subarachnoid spaces. The mechanism by which cerebrospinal fluid is propelled and its circulatory route are not fully understood but are probably influenced by the release of newly produced cerebrospinal fluid, postural effects, the ventricular pulsations, and the pulse pressure of the vascular choroid plexus.
If the intracranial pressure rises because of an increase of cerebrospinal fluid, the fast and the slow axoplasmic transport slow down and papilloedema can be observed in the ocular fundus. Since the cerebrospinal fluid is assumed to communicate between the different cerebrospinal fluid compartments a homogeneous pressure is postulated in all of these cerebrospinal fluid chambers.
However, this anatomical study of the subarachnoid space of the human optic nerve, as well as clinical observations such as asymmetric and unilateral papilloedema, raise legitimate questions as to whether or not the pressure in the subarachnoid space can indeed be expected to be equivalent to the pressure in the ventricles, the cisterns, and the cranial subarachnoid spaces. Because of the influence of the measuring process itself pressure measurements in small and non-homogeneous compartments are technically difficult to perform and the measurements are not necessarily reliable (personal experience) although other groups published their results with more confidence.

Our morphological study of trabeculae and septa within the subarachnoid space of the optic nerve provides strong anatomical evidence that the hydrodynamics between and within the different cerebrospinal fluid segments, especially within the subarachnoid space of the optic nerve, is more complex than within the ideal Bernoulli tube. Cerebrospinal fluid dynamics should therefore not simply be regarded as an undulated continuum in a series of interconnecting chambers. Because the subarachnoid space ends blindly in the bulb segment behind the globe, cerebrospinal fluid needs to reverse its direction of flow in order to return to the site of resorption. There is histological evidence for a cerebrospinal fluid outflow system into the dura of the optic nerve itself; however, this cerebrospinal fluid outflow pathway albeit helpful in maintaining the pressure within certain limits during movements of the globe and optic nerve, may not be sufficient to absorb enough cerebrospinal fluid during a rise in fluid pressure. The subarachnoid space of the optic nerve is at the lower end of the cerebrospinal fluid chambers and because of its cul de sac anatomy the pressure gradient points
unidirectionally from the chiasmal cistern to the subarachnoid space of the optic nerve in the manner of a hydraulic pump. Pressure could therefore build up in this small anatomical compartment that ends blindly behind the globe.

Morgan pointed out that the optic disc is located between two pressure compartments—the globe and the subarachnoid space. The pressure in the subarachnoid space largely determines the retrolaminar pressure. While the optic disc protrudes into the globe in papilloedema, it is excavated in glaucoma. Both diseases are associated with loss of ganglion cell axons. As axonal transport is influenced by pressure gradients, a steady transmigratory pressure is expected to be critical in order to allow optimal axonal transport.21 The retrolaminar pressure depends mainly on the cerebrospinal fluid pressure in the bulb segment. The architecture of the trabecular meshwork is believed to be important for pressure homeostasis. Since the compression and distortion of the lamina cribrosa may contribute to the development of glaucomatous optic neuropathy, likewise the anatomy of the subarachnoid space and the local cerebrospinal fluid pressure may influence both the lamina cribrosa and the retrobulbar part of the optic nerve.22,23

Because of the small size of the trabeculae and septa even high resolution magnetic resonance imaging is not useful in demonstrating their anatomy in normal or in diseased states of the optic nerve.24,25 We suggest that further studies should aim to demonstrate possible changes of the arachnoid trabeculae and septa in pathological conditions such as asymmetric and unilateral papilloedema and optic neuritis.26-31

ACKNOWLEDGEMENTS
The authors thank W Moor (Institute of Pathology, Kantonsspital Aarau, Switzerland) for careful preparation of all the optic nerves used in this study as well as P Rosenbaum, MD (Albert Einstein College of Medicine, New York, USA), and Th Resink MD (University of Basle, Switzerland) for assisting with English language problems. We also thank Mrs M Erni for excellent technical help for SEM and TEM preparation. Special thanks to Duane E Haines, (Department of Anatomy, University of Mississippi Medical Center, Jackson, MI, USA) for many helpful and inspiring comments on the manuscript. Part of this work was supported by the Rudolf and Frida Buck Foundation.

Authors’ affiliations
H E Killer, Department of Ophthalmology, Kantonsspital Aarau, Switzerland
H R Laeng, Department of Pathology, Kantonsspital Aarau, Switzerland
J Flammer, Department of Ophthalmology, School of Medicine, University of Basle, Basle, Switzerland
P Groscurth, Institute of Anatomy, University of Zürich, Switzerland

REFERENCES

www.bjophthalmol.com