Photodynamic therapy with PhotoPoint photosensitiser MV6401, indium chloride methyl pyropheophorbide, achieves selective closure of rat corneal neovascularisation and rabbit choriocapillaris

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Aim: The new photosensitiser PhotoPoint MV6401, indium chloride methyl pyropheophorbide, was assessed as a possible ocular photodynamic therapy agent in a rat model of experimentally induced corneal neovascularisation and in choriocapillaris closure in the rabbit. Optimal drug and light activation parameters were determined.

Methods: MV6401 (Miravant Pharmaceuticals, Inc, Santa Barbara, CA, USA) was activated at 664 nm using a DD3-0665 (Miravant Systems Inc) 0.5 W diode laser. Corneal neovascularisation in rats was induced using an N-heptanol technique. The evaluated drug dosages, light dosages, and post-injection activation times ranged from 0.01–0.1 μmol/kg, 5–25 J/cm², and 10–60 minutes, respectively. The efficacy of MV6401 on normal choriocapillaris and choroidal vessels was evaluated in rabbits with indirect ophthalmoscopy, fundus photography, fluorescein angiography, and histology. In rabbits, the evaluated drug dosages, light dosages, and post-injection activation times ranged from 0.025–0.25 μmol/kg, 3.3–20 J/cm², and 10 minutes, respectively.

Results: In the rat corneal neovascularisation model, an optimal intravenous drug dosage of 0.075 μmol/kg was activated by a 20 J/cm² light dose at 10 minutes after drug administration, the results of which demonstrated early evidence of efficacy in ocular neovascularisation. In rabbits, closure of the normal choriocapillaris was selectively achieved at a drug dosage of 0.15 μmol/kg using light doses from 3.3 to 20 J/cm².

Conclusion: PhotoPoint MV6401 is a potent photosensitiser that demonstrates both efficacy and selectivity in experimental ocular models.

Photodynamic therapy (PDT) provides a viable treatment option for patients with choroidal neovascular membranes (CNVM) in disorders such as age related macular degeneration (AMD). While this new technology is welcomed, the need for improvements over current therapies is apparent by their limited efficacy and the need for re-treatment.

The primary objective of this study was to assess a new photosensitiser PhotoPoint MV6401, indium chloride methyl pyropheophorbide, for possible PDT applications in ophthalmology. Pyropheophorbide based photodynamic therapy has been well studied for utilisation in cancer applications1–3 and have a role in ophthalmic applications. These compounds are well described chemically, hydrophobic, absorb light above 600 nm for enhanced tissue penetration, have excellent photosensitising efficiency, and may not cause prolonged skin photosensitivity experienced with porphyrin derivatives, such as verteporfin.1–3 MV6401 has two main absorption peaks at 423 nm (molar extinction coefficient 101 000 M⁻³cm⁻¹) and 659 nm (74 000 M⁻³cm⁻¹) and its chemical structure is shown in figure 1. These experiments assessed MV6401 in models involving closure of experimentally induced rat corneal neovascularisation and normal rabbit choriocapillaris. Rat corneal neovascularisation1–5 and rabbit choriocapillaris5–7 models have been used previously to study other photosensitising agents.

MATERIALS AND METHODS

Photosensitiser

MV6401 (supplied by Miravant Medical Technologies, Santa Barbara, CA, USA) was formulated in egg yolk phosphatidylcholine (EYP) (purity >98%). The concentration of the ready to use formulation was approximately 1 mM. The agent was stored at 2–8°C until used. The vials were wrapped in aluminium foil to avoid light exposure at all times. The photosensitiser was administered intravenously before light treatment.

Laser

A low power (0.5 W) diode laser (Model DD3-0665; Miravant Systems, Inc, Santa Barbara, CA, USA) at 664 nm was coupled into a slit lamp adapter on a Haag Streit slit lamp to deliver the laser light following MV6401 administration.

Rat corneal neovascularisation model

Animals

Fifty five male Sprague Dawley rats weighing 0.30–0.45 kg were used in accordance with the resolutions on research animal use developed by the Association for Research in Vision and Ophthalmology as well as the guidelines developed by the Miravant Medical Technologies institutional animal care and use committee.

The corneal neovascularisation was induced using an N-heptanol (99%) corneal scrub technique. The drug and light dose ranged from 0.01–0.1 μmol/kg (lateral tail vein injection) and 5–25 J/cm², respectively. Animals were euthanised 5 minutes after light activation.

Abbreviations: AMD, age related macular degeneration; CNVM, choroidal neovascular membranes; EYP, egg yolk phosphatidylcholine; PDT, photodynamic therapy; PIAT, post-injection activation time; RPE, retinal pigment epithelium

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References


anaesthetised with ketamine hydrochloride (50.0 mg/kg) and acepromazine (0.5 mg/kg) before inducing corneal neovessels, laser PDT treatment, and fluorescein angiography.

**Induction of corneal neovascularisation**
Corneal neovascularisation was induced by N-heptanol chemical injury, a technique used previously. In brief, N-heptanol was applied to the corneal epithelium, including the limbal epithelium and accompanying stem cells. The animals were monitored daily and antibiotics were administered to avoid infection (antibiotic topical ointment with or without 2.27% enrofloxacin antibacterial injectable solution). Three to 4 weeks after N-heptanol application, corneal neovessels formed a uniform network across the entire cornea and were ready for PDT treatment.

**Illumination parameters**
A 3 mm diameter spot of laser light (664 nm; 150 mW/cm²) was delivered directly to the cornea without a contact lens. PDT treatment was applied to both eyes at light doses ranging from 5–25 J/cm². Light treatment began 10–60 minutes following MV6401 administration.

**Clinical evaluation**
Corneal fluorescein angiography was performed at baseline and up to 7 or 28 days following PDT to evaluate the neovessel closure. Approximately 0.3 ml sodium fluorescein (10%) was administered intravenously at the time of angiography.

**Histology**
A group of nine Sprague Dawley rats was treated with MV6401 PDT and killed at 1, 7, or 28 days following PDT for histological evaluation. The animals received an intravenous dose of MV6401 at 0.075 μmol/kg and light treatment 10 minutes later with 20 J/cm². Immediately following euthanasia, the eyes were enucleated and submerged in 10% buffered formalin. The cornea was dissected and embedded in glycol methacrylate, sectioned, and stained with haematoxylin and eosin.

**Group assignments**
The study was divided into several phases to elucidate the optimal PDT parameters in the corneal neovascularisation model. The first phase evaluated the drug and light dose-response while the time interval between drug injection and laser light treatment remained constant. The second phase maintained a constant drug dose, while the laser light dose and time interval between photosensitiser injection and laser light treatment were varied. The third phase followed the PDT treatment up to 28 days post-treatment to evaluate lesion evolution; histological analysis was carried out at selected time points in addition to corneal fluorescein angiography.

**Drug and light dose response**
MV6401 was intravenously administered to 19 animals at 0.01–0.10 μmol/kg 10 minutes before light treatment and the light dose was varied from 5 to 25 J/cm² (see table 1). Both eyes were treated at different light doses. Fluorescein angiography was performed at baseline, 1 day, and 7 days post-PDT in this phase of the study.

**Post-injection activation time responses**
In this phase of the study the post-injection activation time (PIAT) interval between MV6401 injection and laser light application was increased; specified times up to 60 minutes were investigated (see table 2). MV6401 was intravenously administered to 17 animals at a fixed dosage of 0.05 μmol/kg and PDT treatment was applied at light dosages of 5–25 J/cm². Fluorescein angiography was performed at baseline, 1 day, and 7 days following PDT.

**Post-treatment evaluations**
Evaluation of ongoing changes in corneal tissues following MV6401 PDT was monitored in 19 animals for periods up to 28 days; eyes from nine of these 19 animals underwent histological processing and analysis at 1, 7, or 28 days post-PDT (see table 3). The animals were intravenously administered 0.075 or 0.10 μmol/kg of MV6401 and PDT treated 10 minutes later with light dosages that ranged from 15–25 J/cm². Fluorescein angiography was performed at baseline, 1, 7, 14, 21, and 28 days following PDT for the first group of animals. The histology group of animals underwent fluorescein angiography at baseline, 1, 7, and 28 days.

### Rabbit choriocapillaris model

**Animals**
Twenty four female Dutch Belted rabbits weighing 1.5–3.0 kg were used in accordance with the resolutions on research animal use developed by the Association for Research in Vision and Ophthalmology as well as the guidelines for proper animal care and use.

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**Table 1** Rat drug and light dosage regimen for the dose-response study in experimentally induced corneal neovascularisation (time interval = 10 minutes)

<table>
<thead>
<tr>
<th>MV6401 dose (μmol/kg)</th>
<th>No. per MV6401 dose</th>
<th>Light dose (J/cm²)</th>
<th>No. per drug/light combination</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.01</td>
<td>3</td>
<td>15, 20, 25</td>
<td>2</td>
</tr>
<tr>
<td>0.025</td>
<td>2</td>
<td>20, 25</td>
<td>2</td>
</tr>
<tr>
<td>0.05</td>
<td>5</td>
<td>5, 10, 15, 20</td>
<td>2</td>
</tr>
<tr>
<td>0.075</td>
<td>5</td>
<td>5, 10, 15, 20</td>
<td>2</td>
</tr>
<tr>
<td>0.10</td>
<td>4</td>
<td>5, 10, 15</td>
<td>2</td>
</tr>
</tbody>
</table>

*Number of animals. †Number of eyes.*
developed by the Indiana University institutional animal care and use committee. Rabbits received intravenous injections ranging from 0.025–0.25 μmol/kg of MV6401 via marginal ear vein. Following photosensitisiser injection, animals were housed in subdued lighting conditions before and after treatment to minimise potential photosensitivity reactions. Control animals were injected with EYP alone using a volume equivalent to 1.0 μmol/kg formulated MV6401. All rabbits were fasted for at least 12 hours before being given anaesthesia. All rabbits were anaesthetised with an intramuscular injection solution of ketamine hydrochloride (50 mg/kg) and xylazine hydrochloride (5 mg/kg). Maintenance amounts (10–15% of the original dosage) were administered at 45 minute intervals as needed.

Illumination parameters
Before light application, a Goldmann plano fundus contact lens (OGFA, Ocular Instruments, Inc, Bellevue, WA, USA) was placed on the animal’s eye. Laser light (664 nm) was delivered to the fundus transcorneally. Power was verified at the cornea by a power meter (International Light Radiometer Model IL1400A, Detector Head Model SPL024F, Newburyport, MA, USA). A power density of 350 mW/cm² was used for all light treatments (1200 μm spot size). For each rabbit, only one eye was irradiated, while the fellow eye served as a drug only control. Laser irradiation started 10 minutes following injection of either MV6401 or EYP alone. Animals were exposed in four regions to the following photodynamic light dosing schedule (nasal to temporal, 10 minutes following injection of either MV6401 or EYP alone). Animals were housed in subdued lighting conditions before and after treatment to minimise potential photosensitivity reactions. Control animals were injected with EYP alone using a volume equivalent to 1.0 μmol/kg formulated MV6401. All rabbits were fasted for at least 12 hours before being given anaesthesia. All rabbits were anaesthetised with an intramuscular injection solution of ketamine hydrochloride (50 mg/kg) and xylazine hydrochloride (5 mg/kg). Maintenance amounts (10–15% of the original dosage) were administered at 45 minute intervals as needed.

Clinical evaluation
Before the day of PhotoPoint MV6401 photodynamic treatment, all rabbits underwent an initial baseline examination to rule out any pre-existing pathology and to document the baseline condition of the rabbits assigned to the study. Clinical examinations were conducted by slit lamp biomicroscopy or indirect ophthalmoscopy. Baseline fundus and fluorescein angiographic photography was also performed. Following MV6401 injection, any changes in tissue appearance from the baseline examination were documented. Adverse changes that were monitored included but were not limited to exudation, infiltration, opacification, optical distortion, oedema, cell loss, pigmentary disruption, vascular anomaly haemorrhage, atrophy, and dislocation or detachment of the retina.

Photodocumentation
Fundus photography and fluorescein angiography were performed before photosensitisiser injections and after photodynamic light treatments. Angiography was performed with 0.1 ml/kg body weight of 25% sodium fluorescein via marginal ear vein injection.

Histology
Immediately following the last scheduled ophthalmological examination, animals were euthanised by intravenous injection. Both eyes were excised immediately and fixed in phosphate buffered 4% paraformaldehyde solution (overnight at room temperature) for histological processing. Tissues were embedded in paraffin, cross sectioned for light microscopy, and stained with haematoxylin and eosin. Ophthalmic tissues were evaluated by an investigator (TAC or MHC) masked to the study groups.

Drug and light dosage responses
In order to establish an appropriate therapeutic MV6401 dosage range, rabbits were divided into six dose-response groups (see table 4), with laser photodynamic treatment sites within the test eye receiving light dosages of 5, 10, and 20 J/cm². Subsequently, eyes were evaluated at days 1 and 7 post-treatment, and were excised at day 7 after euthanasia.

<table>
<thead>
<tr>
<th>Table 2</th>
<th>Rat post-injection activation time (PIAT) and light dosage regimen</th>
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<tbody>
<tr>
<td>Time interval (minutes)</td>
<td>No* per time interval</td>
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<tr>
<td>20</td>
<td>5</td>
</tr>
<tr>
<td>30</td>
<td>6</td>
</tr>
<tr>
<td>60</td>
<td>6</td>
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*Number of animals.
†Number of eyes.
(Animals were intravenously administered 0.05 μmol/kg of MV6401.)

<table>
<thead>
<tr>
<th>Table 3</th>
<th>Rat drug and light dosage regimen used in post-PDT evaluations in experimentally induced corneal neovascularisation (time interval = 10 minutes)</th>
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</thead>
<tbody>
<tr>
<td>MV6401 dose (μmol/kg)</td>
<td>Days post-treatment (euthanasia)</td>
</tr>
<tr>
<td>0.075</td>
<td>1</td>
</tr>
<tr>
<td>7</td>
<td>20</td>
</tr>
<tr>
<td>28</td>
<td>3†</td>
</tr>
<tr>
<td>28</td>
<td>8†</td>
</tr>
<tr>
<td>0.10</td>
<td>28</td>
</tr>
</tbody>
</table>

*Number of animals.
†Number of eyes.
‡Number of animals evaluated histologically (note: only one eye per animal was treated; the fellow eye served as a drug only control).
*In this group 10 out of 16 eyes were treated.
Lesion development following PDT

In a second set of experiments, the evolving histopathological responses of ocular tissues to MV6401 PDT at different light dosage levels (3.3, 5, 10, and 20 J/cm²) were evaluated, with particular emphasis on the possible post-treatment development of lesions (see table 5). After the results from the earlier dose-response study suggested that an MV6401 dosage of 0.15 μmol/kg was therapeutically most appropriate, this dose was then used to evaluate possible lesion development at various post-treatment time points up to 28 days.

RESULTS

Rat corneal neovascularisation model

A dose-response was observed as MV6401 dosage levels were escalated from 0.01 μmol/kg to 0.1 μmol/kg in Sprague Dawley rats. Fluorescein angiography did not show any closure of the corneal neovascularity at the lowest drug dose (0.01 μmol/kg) when treated at the highest light doses (15–25 J/cm²). The 0.025 μmol/kg dose resulted in an area of neovessel closure the size of the treatment area 1 day following photodynamic treatment, but no effect was evident at 7 days post-treatment. The 0.05 μmol/kg dose demonstrated a light dose dependent response with the most effective drug and light combination at 20 and 25 J/cm² showing sustained neovessel closure at 7 days post-treatment. The highest MV6401 doses (0.075 μmol/kg and 0.10 μmol/kg) resulted in the largest areas of neovessel closure as well as sustained closure at 7 days (see figs 3 and 4). The optimal dosage scheme was 0.075 μmol/kg MV6401 with a light dosage of 20 J/cm², because the neovessel closure at the highest MV6401 and light dose (0.10 μmol/kg and 20 J/cm²) was accompanied by severe chemosis and loss of selectivity for the neovessels.

The time interval was evaluated for periods from 10–60 minutes between initial drug injection and light dose administration, and the optimal time interval was determined to be 10 minutes based on the maximum closure of the neovascularature (as evidenced by fluorescein angiography).

Effects subsequent to treatment at the highest drug doses (0.075 μmol/kg and 0.10 μmol/kg) were monitored up to 28 days following photodynamic treatment (see fig 3). Corneal fluorescein angiography of the majority of treatment responses showed regrowth or recanalisation of the corneal neovessels by 21 days post-PDT. Histology (fig 4) revealed evidence of neovessel collapse and extensive thrombosis at 1 day post-PDT. Thrombosis was less extensive at 7 days. By 28 days, patent neovessels dominated the treatment field with occasional evidence of thrombosis.
Rabbit choriocapillaris dose-response experiments

The MV6401 dose-response experiments showed no acute fundus lesions, suggesting that laser treatment at the fluences used had no thermal effect, and that the ocular effects observed later were photochemical in origin. At the lowest MV6401 dose (0.025 μmol/kg) there was only minimal retinal pigment epithelium (RPE) mottling, noted variably at 1 day post-treatment, and these findings resolved on examination and angiography by 7 days post-treatment. At MV6401 doses of 0.1 μmol/kg and higher, the lesions exhibited subretinal fluid at post-treatment day 1, and this finding also resolved by 7 days post-treatment, except at sites where the highest dose was used (0.25 μmol/kg). There were no effects observed on the optic nerve or major retinal vessels outside of the treatment zones. There were no effects observed in the cornea, iris, lens, or vitreous at any of the doses used in this study.

Figure 4  Rat corneal sections obtained (A) 1 day, (B) 7 days, and (C) 28 days after PDT with 0.075 μmol/kg MV6401 and 20 J/cm² light (PIAT interval = 10 minutes).

Figure 5  Fundus photographs of laser induced PDT sites (left column: temporal retina containing 20 and 10 J/cm² PDT sites, middle column: optic disc, right column: nasal retina containing 5 and 3.3 J/cm² PDT sites) at 1 day (top row), 7 days (second row), 14 days (third row), and 28 days (fourth row) post-treatment (MV6401 dosage: 0.15 μmol/kg). All photographs represent the same eye in one animal. The fifth row illustrates fundus photographs from a control eye (different animal), 1 day following laser light treatment using the same laser parameters described previously. Tissues appeared unaffected here and in later histological sections.
With regard to the desired end point of choriocapillaris closure, there was subthreshold dosing at 0.025, 0.1, and 0.125 μmol/kg MV6401, as fluorescein angiography revealed perfusion of the choroid at 7 days post-treatment. Toxicity was observed on histopathological analysis of photodynamically treated tissues that received 0.20 or 0.25 μmol/kg dosages of MV6401, with RPE atrophy outside of the treatment zones and severe disorganisation of the retinal outer segments.

Photodynamic treatments using an intermediate dosage of MV6401 (0.15 μmol/kg) produced the desired end point, although this result may represent a narrow therapeutic range between 0.125 and 0.20 μmol/kg, at least within the rabbit. There were no fundus lesions acutely discernable. At 1 day post-treatment, examination revealed prominent grey-white elevated lesions at each of the three treatment sites, consistent with subretinal fluid. These varied in size and intensity, correlating with the laser light dosage used. Fluorescein angiography showed absolute homogeneous hypofluorescence at each of the lesions, consistent with non-perfusion of the choriocapillaris or with blockage by turbid subretinal fluid/opaque RPE, or both.

By 7 days post-treatment (at the 0.15 μmol/kg dosage), there were three discrete fundus lesions, each consisting of relative (not absolute) RPE atrophy. Compared to post-treatment day 1, these lesions no longer showed the homogeneous grey-white colour or elevation, suggesting resolution of subretinal fluid. These lesion sites no longer exhibited any adjacent RPE changes outside of the treatment zones. Fluorescein angiography showed homogeneous hypofluorescence at the lesions, suggestive of choriocapillaris non-perfusion. Except for some mild staining at the margins of the lesions there were no other angiographic abnormalities. There were no corneal, iris, lenticular, or vitreous lesions seen at any time point.

At an MV6401 dosage of 0.15 μmol/kg, histopathological analysis at 1 week post-treatment demonstrated significant attenuation of the choroidal layer/vasculature, consistent with closure of the choriocapillaris and moderate sized choroidal vessels at each of the photodynamic treatment sites, and this result was consistent with the above mentioned fluorescein angiography findings. There was also attenuation of the photoreceptor layer within the laser treatment sites, which was directly correlated to the laser light dosages. At the highest light dose only (20 J/cm²) there was subretinal fibrosis.

**Rabbit choriocapillaris lesion development with 0.15 μmol/kg MV6401**

Given the findings noted above, further study of the photodynamic response to MV6401 was carried out with

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**Figure 6** Fluorescein angiography of PDT sites in the (left column) temporal retina (20 and 10 J/cm² sites) and (right column) nasal retina (5 J/cm² and 3.3 J/cm² sites) at 1 day (top row) and again at 28 days (bottom row) post-treatment. This was the same PDT treated eye as that shown in fundus photographs of figure 5.

**Figure 7** Photomicrographs of retinal/choroidal tissues (radial view) following PDT at duration dependent energy doses of (A) 10 J/cm² and (B) 5 J/cm² that showed closure of choroidal vessels and attenuation (5 J/cm²) and partial ablation (10 J/cm²) of photoreceptors. In each case, MV6401 dosage was 0.15 μmol/kg. Control tissues (C), representing a laser site at the 5 J/cm² light dosage level, received laser light treatment without MV6401 infusion. Here retinal structures remained completely intact and choroidal vessels remain open.
the 0.15 μmol/kg MV6401 dosage and included an additional lower laser light dosage of 3.3 J/cm².

All treatment lesions (fig 5) showed the presence of subretinal fluid at day 1 post-treatment, although this finding resolved by day 7 (except at the highest light dose (20 J/cm²) in which fluid was variably present, but resolved by 14 days post-treatment). The subretinal exudate variedly evolved to subretinal fibrosis within the higher light dosage sites (10 and 20 J/cm²) and this effect appeared most prominently in rabbits which demonstrated relative RPE hypertrophy at baseline. It is possible that the more prominent RPE mounts a more aggressive metaplastic/fibroblastic response to photochemical injury. In general, examination showed that the remaining (lower light dosage) lesion sites evolved to RPE mottling without subretinal exudate or fibrosis at the later time points.

With regard to photodynamic closure of the choroidal vessels, fluorescein angiography (fig 6) revealed homogenous hypofluorescence at the lesions, suggestive of choriocapillaris non-perfusion, at day 7 post-treatment. This effect further evolved to only mottled hypofluorescence with marginal staining by 28 days. One interpretation of this change would be the possible reperfusion of the choroidal vessels. However, histopathological analysis (fig 7) showed significant attenuation of the choroidal layer/vasculature, suggestive of primary closure of the choriocapillaris and of moderate sized choroidal vessels. There was also attenuation of the photoreceptor layer following treatment at the 3.3 and 5.0 J/cm² laser light dosages with improvement noted by 28 days. At the higher (10 and 20 J/cm²) laser light dosages, generalised damage to the photoreceptors was observed. Control eyes, receiving laser light treatments without infusion of MV6401, did not exhibit any vessel closure or damage to the retina.

DISCUSSION

This study evaluated the new photosensitiser PhotoPoint MV6401, indium chloride methyl pyropheophorbide. As noted previously, pyropheophorbide based photodynamic therapy has been well studied for utilisation in cancer applications. These compounds are well described chemically, are hydrophobic, absorb light above 600 nm for enhanced tissue penetration, have excellent photosensitising efficiency, and may not cause prolonged skin photosensitivity experienced with porphyrin derivatives, such as verteporfin. Verteporfin, an efficient generator of singlet oxygen, shows peak absorption in the ultraviolet A range, as well as a secondary absorption peak between 680 and 695 nm; in ophthalmic applications, it is activated by low power, non-thermal laser light at 689 nm, which is thought to penetrate blood, melanin, and fibrous tissue. Like verteporfin, MV6401 possesses two main absorption peaks in the ultraviolet and red ranges, at 423 nm (molar extinction coefficient 101 000 M⁻¹ cm⁻¹) and 659 nm (74 000 M⁻¹ cm⁻¹).

The current study, which demonstrates efficacy and selectivity in experimental ocular models of rat corneal neovascularisation closure and of rabbit choriocapillaris closure, suggests that PhotoPoint MV6401 could potentially be used as a photosensitiser in ophthalmic applications. In the rat corneal neovascularisation model, effective closure of these vessels was achieved with an MV6401 dosage of 0.075 μmol/kg and a laser light dosage of 20 J/cm². In the rabbit choriocapillaris closure experiments, a 0.15 μmol/kg dosage produced the most desirable end point with preferential effect on the choriocapillaris and choroid and minimal retinal toxicity. Specifically, angiography and histology demonstrated primary closure of the choriocapillaris and moderate sized choroidal vessels. However, limitations of this study include that lack of power to make definitive claims about dosing and the limited ability to model human choriocapillaris and choroidal neovascularisation with the rat and rabbit model; further study with this agent in a primate model has recently been completed.

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Proprietary interest: Drs Snyder and Small are employees of Miravant Medical Technologies Inc.

This research investigation was performed at both locations.

PhotoPoint is a trademark of Miravant Medical Technologies, Santa Barbara, CA, USA.

REFERENCES