Characteristic clinical features as an aid to the diagnosis of suppurative keratitis caused by filamentous fungi

P A Thomas, A K Leck, M Myatt

Aim: To assess whether the presence of characteristic clinical features can be used as a diagnostic aid for suppurative keratitis caused by filamentous fungi.

Methods: Patients presenting with suppurative keratitis in India underwent detailed clinical examination followed by microbiological investigation of corneal scrapes. A partial diagnostic score based upon the strength of the association, as estimated by the odds ratio, between reported clinical features and laboratory confirmed diagnoses was devised and subsequently tested using a case series from Ghana.

Results: Serrated margins, raised slough, dry texture, satellite lesions and coloration other than yellow occurred more frequently in cases of filamentous fungal keratitis than bacterial keratitis (p<0.05). Hypopyon and fibrinous exudate were observed more frequently in bacterial keratitis (p<0.05). When incorporated into a backwards stepwise logistic regression model only serrated margins, raised slough, and colour were independently associated with fungal keratitis; these features were used in the scoring system. The probability of fungal infection if one clinical feature was present was 63%, increasing to 83% if all three features were present.

Conclusions: Microbiological investigations should be performed whenever possible; however, where facilities are not available, a rapid presumptive diagnosis of suppurative keratitis may be possible by scoring clinical features.

Methods
A prospective study of suppurative keratitis (defined as loss of corneal epithelium with underlying stromal infiltrate and suppuration associated with signs of inflammation, with or without hypopyon) was conducted at three centres in southern India and in three centres in Ghana between June 1999 and May 2001. The aetiology of the infection in 1090 consecutive cases (800 from India and 290 from Ghana) has already been reported.9

Clinical findings
Ophthalmologists examined patients at the different centres using a standardised protocol and proforma. A detailed examination was performed on each patient at the slit lamp; clinical features were documented, drawings made for patient records, and a photograph was taken of the affected eye. The length of the slit beam was used to assess the vertical diameter of the corneal ulceration; this was then turned 90° to assess the horizontal diameter. Using the slit beam parallelogipied the depth of ulceration and infiltrate were assessed. The presence or absence and form of the following clinical features were documented: elevation of slough (raised, flat), texture of slough (wet, dry), ulcer margins (serrated, well defined), satellite lesions, immune ring, hypopyon, keratic precipitates, or peripheral infiltrate, fibrin, flare or cells in the anterior chamber (AC), and deep lesions (posterior corneal abscess, endothelial plaque).

Microbiological investigations
Laboratory diagnosis was determined using microscopy and culture.7 Microbial cultures were considered to be significant if (i) growth of the same organism was demonstrated on two or more solid culture media; (ii) there was semi-confluent growth at the site of inoculation or growth on one solid medium consistent with microscopy (that is, appropriate staining and morphology with Gram stain); or (iii) semi-confluent growth at the site of inoculation on one solid medium consistent with microscopy.

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Suppurative keratitis is an important cause of monocular blindness worldwide, in particular in developing countries in tropical latitudes. Most ophthalmologists in the developing world do not have recourse to specialised ocular microbiological laboratory facilities and may have to depend on their own clinical acumen for diagnosis, with empirical treatment of patients with these infections. Kaufman and Wood described certain clinical manifestations that they believed to be characteristic of mycotic keratitis, including a severe ocular reaction; folds in Descemet’s membrane; “hyphate” margin; firm and elevated lesions; hypopyon; endothelial plaque and immune ring.1 However, these descriptions were based on a study of only 15 patients with laboratory diagnosed fungal keratitis. According to Jones, fungal infection is likely in a suppurative corneal lesion with one or more of the following features: a dry, raised ulcer with crenate, spiculate or pseudohyphate border; satellite lesions; recurrent hypopyon or posterior chamber endophthalmitis with progressive shallowing of the anterior chamber; failure to respond to antibacterial treatment.2 These guidelines were based on observations in just 29 patients with laboratory diagnosed fungal keratitis.

Since then these clinical features have generally been accepted as being diagnostic of filamentous fungal keratitis. Keratitis caused by yeast-like fungi such as Candida spp has been described as resembling bacterial keratitis.3 Recent studies on large numbers of patients with culture proved mycotic keratitis have tended to document only the frequency of occurrence of these features and there has hitherto been no attempt to compare the occurrence of these features in a large series of patients with bacterial and fungal keratitis with a view to assessing their suitability as diagnostic aids for suppurative keratitis.5 In this study we have attempted to determine to what extent the presence of certain clinical features may be used to provide a rapid, presumptive diagnosis of suppurative keratitis caused by filamentous fungi.
medium (if bacteria); or (iv) growth of the same organism on repeated scraping. Culture positivity was the “gold standard” used to establish the diagnosis of the bacterial ulcers. If fungal hyphae were observed in corneal tissue, but failed to grow in culture, the causative organism was reported as fungal.

Verbal patient consent was obtained but written consent was not considered appropriate as the study did not involve any deviation from routine diagnostic or treatment protocol. Patients were excluded from the study if they did not agree to investigation and treatment.

Cases included in this study
In this study only patients with confirmed bacterial or fungal infection were included in the analysis. The following patients were excluded from the 1090 consecutive cases: those with suspected or confirmed viral keratitis, corneas at risk of perforation (where corneal scrapes were not possible) and small ulcers; defined as an epithelial defect <2 mm (greatest diameter) and/or an infiltrate of <4 mm (greatest diameter) (128) as these comprised early stage corneal ulcers which may present with non-specific signs, and characteristic clinical features may not be present. Those with a mixed infection (44), Acanthamoeba keratitis (seven), unconfirmed laboratory diagnosis (249), or where clinical features had not been adequately recorded (12) were excluded. The clinical features of 360/800 patients in India with confirmed bacterial or fungal infection were analysed further to devise a diagnostic score. The diagnostic score was subsequently applied to 115/290 cases of bacterial or fungal infection from the Ghanaian patient dataset (after excluding patients according to the criteria applied to the Indian dataset).

Analysis of data
From the Indian dataset pairwise associations between clinical features and diagnosis were investigated using SPSS (SPSS for Windows, Release 10.0.5, SPSS Inc, 1989–2001) and EpiCalc.11 Significant associations were entered into a logistic regression model and non-significant associations removed using backwards elimination. A score was created from a count of signs positively and independently associated with a fungal aetiology. An operating characteristic (OC) curve for the diagnostic score was created by computer based simulation with the prevalence of fungal infection at each score estimated in 1000 bootstrap (that is, with replacement) samples of size 1000 taken from the clinical dataset. The probability of fungal infection was estimated from the median prevalence found at each score in the bootstrap samples. A 95% confidence interval around the probability of fungal infection was estimated using the 2.5% and 97.5% quantiles of the distribution of the prevalences found at each score in the bootstrap samples. Simulations were performed using the R language for data analysis and graphics.12–14 Data were plotted using R. The Ghanaian patient dataset was tested using the diagnostic score devised from the Indian data, and an OC curve was created by computer based simulation, as for the Indian data, and compared.

RESULTS
Clinical data for 360 patients from India with confirmed fungal (228) and bacterial (132) keratitis were analysed. Features which occurred significantly (p<0.05) more frequently in fungal than in bacterial corneal ulcers by univariate analysis were as follows: serrated margins, raised slough, dry textured slough, satellite lesions, and colour

Table 1  Univariate analysis of clinical features occurring in fungal and bacterial keratitis

<table>
<thead>
<tr>
<th>Clinical feature</th>
<th>Frequency (%) fungal</th>
<th>Frequency (%) bacterial</th>
<th>OR (CI)</th>
<th>p Value</th>
<th>Sens</th>
<th>Spec</th>
<th>PPV</th>
</tr>
</thead>
<tbody>
<tr>
<td>Serrated margins</td>
<td>180/228 (79%)</td>
<td>63/132 (48%)</td>
<td>37.14</td>
<td>4.09 (2.57 to 6.56)</td>
<td>0.00</td>
<td>0.79</td>
<td>0.52</td>
</tr>
<tr>
<td>Raised slough</td>
<td>135/228 (59%)</td>
<td>52/132 (39%)</td>
<td>13.50</td>
<td>2.23 (1.44 to 3.55)</td>
<td>0.00</td>
<td>0.59</td>
<td>0.61</td>
</tr>
<tr>
<td>Dry texture of slough</td>
<td>101/228 (44%)</td>
<td>37/132 (28%)</td>
<td>9.36</td>
<td>2.04 (1.29 to 3.26)</td>
<td>0.00</td>
<td>0.44</td>
<td>0.72</td>
</tr>
<tr>
<td>Satellite lesions</td>
<td>51/222 (23%)</td>
<td>17/132 (13%)</td>
<td>4.91</td>
<td>1.95 (1.08 to 3.61)</td>
<td>0.04</td>
<td>0.22</td>
<td>0.87</td>
</tr>
<tr>
<td>Hypopyon</td>
<td>105/219 (48%)</td>
<td>83/128 (65%)</td>
<td>9.29</td>
<td>0.50 (0.32 to 0.78)</td>
<td>0.00</td>
<td>0.48</td>
<td>0.35</td>
</tr>
<tr>
<td>Fibrin</td>
<td>21/210 (10%)</td>
<td>28/125 (22%)</td>
<td>9.65</td>
<td>0.38 (0.20 to 0.70)</td>
<td>0.00</td>
<td>0.10</td>
<td>0.78</td>
</tr>
<tr>
<td>Colour (not yellow)</td>
<td>213/228 (93%)</td>
<td>106/132 (80%)</td>
<td>14.26</td>
<td>3.47 (1.77 to 6.98)</td>
<td>0.00</td>
<td>0.93</td>
<td>0.20</td>
</tr>
</tbody>
</table>

PPV, positive predictive value.

Table 2  Multivariate analysis of clinical features occurring in fungal and bacterial keratitis

<table>
<thead>
<tr>
<th>Clinical feature</th>
<th>OR (CI)</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Serrated margins</td>
<td>3.45 [2.12 to 5.64]</td>
<td>0.00</td>
</tr>
<tr>
<td>Raised slough</td>
<td>2.32 [1.43 to 3.74]</td>
<td>0.00</td>
</tr>
<tr>
<td>Fibrin</td>
<td>0.39 [0.20 to 0.77]</td>
<td>0.01</td>
</tr>
<tr>
<td>Colour (not yellow)</td>
<td>2.85 [1.34 to 6.03]</td>
<td>0.01</td>
</tr>
</tbody>
</table>
Operating characteristic (OC) curve for diagnostic score

Table 3 Screening test indices at each level OC curve

<table>
<thead>
<tr>
<th>Score</th>
<th>Sensitivity</th>
<th>Specificity</th>
<th>PPV</th>
<th>NPV</th>
</tr>
</thead>
<tbody>
<tr>
<td>&gt;0</td>
<td>0.99 (0.97, 1.00)</td>
<td>0.08 (0.04, 0.14)</td>
<td>0.65 (0.60, 0.70)</td>
<td>0.83 (0.51, 0.97)</td>
</tr>
<tr>
<td>&gt;1</td>
<td>0.88 (0.83, 0.92)</td>
<td>0.41 (0.33, 0.50)</td>
<td>0.72 (0.66, 0.77)</td>
<td>0.66 (0.54, 0.76)</td>
</tr>
<tr>
<td>&gt;2</td>
<td>0.45 (0.38, 0.51)</td>
<td>0.84 (0.76, 0.90)</td>
<td>0.83 (0.75, 0.89)</td>
<td>0.47 (0.40, 0.53)</td>
</tr>
</tbody>
</table>

PPV, positive predictive value; NPV, negative predictive value.

Table 4 Screening test indices at each level OC curve (Ghana test data)

<table>
<thead>
<tr>
<th>Score</th>
<th>Sensitivity</th>
<th>Specificity</th>
<th>PPV</th>
<th>NPV</th>
</tr>
</thead>
<tbody>
<tr>
<td>&gt;0</td>
<td>0.94 (0.87, 0.98)</td>
<td>0.15 (0.05, 0.36)</td>
<td>0.79 (0.70, 0.86)</td>
<td>0.44 (0.15, 0.77)</td>
</tr>
<tr>
<td>&gt;1</td>
<td>0.64 (0.53, 0.74)</td>
<td>0.46 (0.27, 0.66)</td>
<td>0.80 (0.69, 0.88)</td>
<td>0.27 (0.15, 0.43)</td>
</tr>
<tr>
<td>&gt;2</td>
<td>0.25 (0.16, 0.35)</td>
<td>0.92 (0.73, 0.99)</td>
<td>0.92 (0.72, 0.99)</td>
<td>0.26 (0.18, 0.37)</td>
</tr>
</tbody>
</table>

PPV, positive predictive value; NPV, negative predictive value.

No significant differences were observed between the frequency of occurrence of an immune ring, keratic precipitates, perineural infiltrates, endothelial plaque, and flare or cells in the AC. Clinical features found to be characteristic of fungal or bacterial infection were entered into a logistic regression model.

Serrated margins, raised slough, coloration other than yellow, and fibrin were statistically independent features as determined by the logistic regression model (table 2).

Using the three clinical features associated with fungal infection a score was devised, with the presence of a significant feature scoring +1. The higher the score, the greater the probability of fungal infection (fig 1/table 3).

The probability of fungal infection if one of the clinical features was present; either a serrated margin, raised slough or coloration other than yellow; was 63%; the presence of all three signs indicated an 83% chance of fungal infection.

The results obtained from testing the cases from Ghana closely correlated with those from the Indian dataset and the presence of all three signs resulted in a probability of >90% that the infection was fungal (fig 2/table 4).

Approximately 17% of the corneal ulcers in the Indian dataset and >50% of corneal ulcers in Ghana scored zero. These comprised bacterial corneal ulcers and fungal corneal ulcers for which all of the scored clinical features were absent.

DISCUSSION

Filamentous fungi are the commonest cause of mycotic keratitis in many countries in tropical latitudes and it is vital that a specific diagnosis is made as quickly as possible to ensure prompt institution of antifungal therapy. Although a detailed clinical examination may help to reach a rapid presumptive diagnosis, fungal keratitis continues to be confused with other causes of inflammatory keratitis. Certain clinical characteristics of corneal ulcers may suggest a specific pathogen, but it is now generally accepted that a reliable diagnosis cannot be made by clinical appearance alone and that microbiological investigations should be performed. Unfortunately, many ophthalmologists working in developing countries do not have access to basic ocular microbiological investigations such as microscopy or culture of corneal scrapes. Even in the United States, patients with corneal ulcers are frequently not referred for microbiological investigations and ophthalmologists tend to depend on their clinical acumen when prescribing. Thus, it is imperative to assess the reliability of what are considered to be “characteristic” clinical features in the diagnosis of suppurative keratitis.

The observations of Kaufman and Wood and Jones are now firmly established in the ophthalmic literature, however, both of these and other similar observational studies have limitations. There was no comparison of the frequency of occurrence of these clinical features in fungal, bacterial, or other types of microbial ulcers and there was no assessment of the relative importance of these different characteristics in establishing a diagnosis. It is essential to determine the validity of these concepts, an issue which most published case series of microbial keratitis have not attempted to address.

Only in recent studies have authors compared clinical features in fungal and bacterial keratitis. In a study of 142 patients with suppurative keratitis in Bangladesh, patients with culture proved fungal keratitis (almost all caused by filamentous fungi) reported a longer history of symptoms than bacterial ulcers (p<0.01) and a dry, raised, necrotic or fluffy surface were more frequent (p<0.01). Endothelial rings were also more frequent in fungal than in bacterial ulcers, whereas dacryocystitis was significantly more common in bacterial ulcers. Wong et al compared fungal and bacterial keratitis in a hospital based retrospective study in Singapore and reported anterior chamber involvement to be more common in fungal ulcers (45% v 35%). In many of the
studies serrated margins are commonly reported in association with fungal infection.\textsuperscript{1–3} In our study 79% of fungal ulcers had serrated margins, but this sign was not pathognomonic, as 48% of bacterial ulcers also had serrated margins.

Another important feature of fungal corneal ulcers is believed to be raised, dry, necrotic slough.\textsuperscript{2,4} In our study dry textured slough was seen more frequently in fungal corneal ulcers. Satellite lesions, which are reported to occur frequently in fungal corneal ulcers, were also seen in cases of bacterial keratitis although less frequently occurring. An immune ring is believed to be a frequent occurrence in a filamentous fungal corneal ulcer; in the present study, an immune ring was only observed in very few cases. Anterior chamber pathology (hypopyon, fibrin) was more frequently observed in bacterial than in fungal ulcers. This is in contrast with the observations of some authors but consistent with the findings of others.\textsuperscript{1–3,14–17,25,26}

An ophthalmologist confronted with a patient with suppurative keratitis may be uncertain as to which clinical features should be given more importance in differentiating between fungal and bacterial infection. Although certain clinical features appear to be more strongly associated with fungal infection we do not believe it is wise to depend on a single clinical feature to rule out a presumptive diagnosis. The rationale for devising a scoring scheme was based on the premise that the simultaneous presence of several “characteristic” clinical features in a corneal ulcer should permit a more precise diagnosis of fungal infection than individual characteristics considered in isolation. If a high score is obtained then the observer can be 83% certain of fungal infection. Conversely, a low score (including zero) is associated with increased probability of bacterial aetiology.

The greatest benefit of a clinical score such as this may be to clinicians who are working in eye centres where there are no facilities for laboratory investigation. The score may be used to provide a rapid indication of the type of infection, essential in guiding treatment choices. The management of corneal ulcers using this scheme will vary depending on the setting in which it is used. For example, a clinician working in a region where fungal keratitis is thought to be common may decide to use antifungals to treat ulcers with a score of 1 or 2, whereas a clinician working in an area in which fungal keratitis is less common may reserve antifungals for ulcers with a maximum score of 3. Choosing a low score as a guide for management as a fungal ulcer may result in unnecessary treatment of bacterial ulcers with antifungals; however, patients with very early disease would not be missed. In contrast, the selection of a highest score as a cut-off point would permit more specific diagnosis, and a more selective institution of antifungal therapy; however, this may mean that for patients with very early disease, where the clinical features may be vague and non-specific, appropriate treatment may be delayed. For ophthalmologists living in areas where fungal keratitis is a common or frequent cause of keratitis (southern India, sub-tropical Africa, and southern Florida), low scoring ulcers may be given antifungals whereas for those working in more temperate climates (the United Kingdom, northern United States) the decision to treat with antifungals might be confined to those cases which score highly. The decision as to when to treat an ulcer with antifungals may also be influenced by other factors such as financial constraints and availability of antifungal agents. It is recommended that fungal infections are treated with a combination of an antibiotic and an antifungal agent in case of mixed infection.

We did not stratify clinical presentation based on the duration of the symptoms as this information was thought to be unreliable as the majority of patients reported short duration of symptoms that did not concur with the clinical picture. It is possible that if duration of symptoms were included the results might have been different. We have also used only two broad categories—namely, culture proved bacterial keratitis, and microscopy or culture proved fungal keratitis, and not introduced subsets based on the infecting fungal or bacterial genus in each category, as a larger number of cases would have been needed. However, from our results it is clear that bacterial and fungal ulcers can exhibit the same features and therefore analysing the data based on genera may reduce the usefulness of such a test. Further investigations to test and refine this scoring scheme are currently under way and there are plans to validate the proposed scheme in different settings.

CONCLUSIONS

The clinical features of microbial keratitis may vary considerably and no one clinical feature can be considered as absolutely pathognomonic of a particular type of aetiological agent.

Ophthalmologists are urged to send corneal scrapes for microbiology examination where facilities for ocular microbiology are available. However, where such facilities are not available, a rapid presumptive clinical diagnosis of filamentous fungal keratitis may be possible using a tool such as the scoring scheme presented here. When fungal infection is suspected a combination of antibiotic and antifungal therapy is recommended.

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Competing interests: none declared

Ethical approval was obtained from the ethics committees at Moorfields Eye Hospital, Joseph Eye Hospital (India) and Korle Bu Teaching Hospital (Ghana).

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